

Phytoremediation, a novel strategy for the removal of toxic metals from the environment: biochemical and molecular mechanisms

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Outline

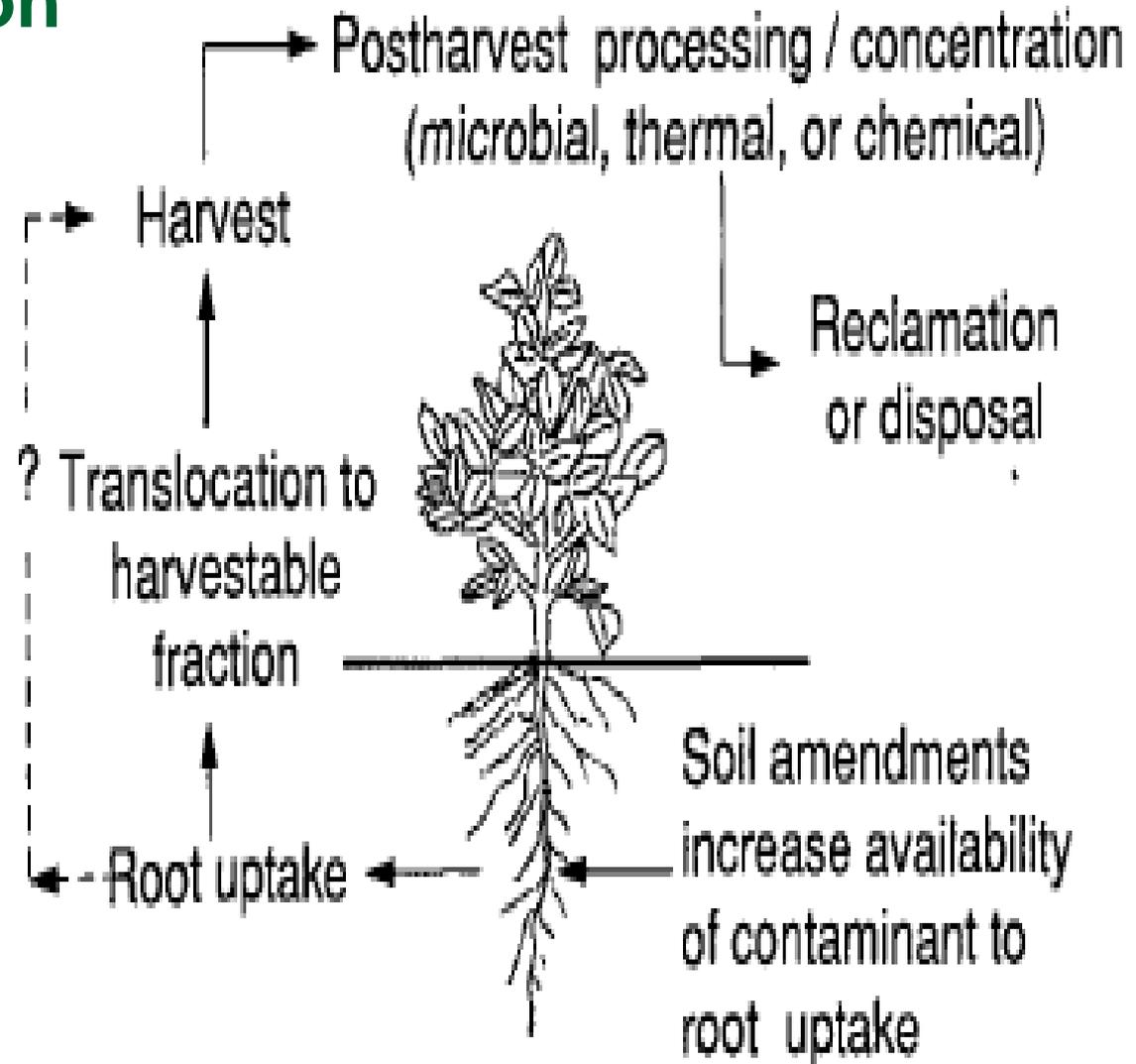
- Introduction of phytoremediation
 - *Sesbania drummondii*
 - Metal uptake
 - Microscopic evidence of metal transport
 - Biotransformation of metal compounds
 - Stress enzymes
 - Gene identification/expression
 - Long term goal
 - Conclusion
-

Phytoremediation

Use of vegetation for the *in situ* treatment of contaminated sites

- A fast emerging environmental clean up strategy
 - Immense promise for remediation of contaminated sites (soil, ground water, waste water)
 - Effective against
 - **inorganic** (toxic metals and nutrients)
 - **organic pollutants** (BTEX)
 - chlorinated solvents, ammunition wastes
-

Phytoextraction Process



Background

■ Sources of pollution:

- ❑ Mining and smelting, municipal wastes, sewage sludge, landfill leachates, fertilizers, pesticides, nuclear accidents

■ Dimension of the problem:

- ❑ 1980 Statute recognized over 40,000 Superfund sites endangering human health
 - ❑ >10,000 sites remain active today (Superfund Accomplishment Figures-FY 2003)
 - ❑ 40% of these sites have problems of heavy metal (Pb, Cd, Cr, As, Zn etc.) contamination
-

Conventional remediation strategies against metal contaminations

- Excavation and reburial of contaminated soils to another site
 - Soil flushing/washing
 - Solidification/stabilization
 - Vitrification
 - Electro-kinetics
-

Cost Analysis

- Conventional engineering technology v/s Phytoremediation (TIBTECH, 13, 1995)

Contaminants	Conventional Technology	Phytoremediation
Water soluble/ volatile compounds	\$10-100 per m ³ soil	\$ 0.02-1.00 per m ³ soil (\$200-10,000 per hectare) of cropping
Compounds requiring land-filling or low temp. thermal treatments	\$ 60-300 per m ³ soil	
Materials requiring special land-filling or high temp. thermal treatment	\$ 200-700 per m ³ soil	
Incineration	\$ 100 per m ³ soil	
Radionucleides	\$ 1000-3000 per m ³ soil	

Benefits

- Economically feasible
 - Socially desirable
 - Environment friendly
 - Improves soil health
 - Effective
-

Phytoremediation approaches

1. **Phytoextraction**: to remove contaminants directly from soil/water
 2. **Phytostabilization**: use of vegetation and soil amendments to reduce contaminant availability and movement.
 3. **Rhizofiltration**: plant root system is directed to extract pollutants from water bodies
 4. **Phytomining**: for extraction and concentration of valuable metals
-

Prerequisites for Phytoremediation

Hyperaccumulators

- Accumulate 100 times more metals than the non-accumulators
 - Conc. Criterion (% Shoot DW)
Cd (>0.01), Co, Cu, Cr and Pb (>0.1),
Ni and Zn (>1), Hg (0.001)
 - Should have good biomass
-

Terrestrial Hyperaccumulators (Brooks, 1998)

Plant	Metal	% metal in shoot (DW)
<i>Thlaspi caerulescens</i>	Zn, Cd	>2% Zn, >0.1% Cd,
<i>Thlaspi spp.</i>	Zn	>2%
<i>Cardaminopsis hallerii</i>	Zn	>1%
<i>Brassica spp.</i>	Se	
<i>Astragalus spp.</i>	Se	0.1-1%
<i>Atriplex spp.</i>	Se	
<i>Thlaspi rotundifolium</i>	Pb	<1% (~0.8%)
<i>Aelloanthus subacaulis</i>	Cu	1.3%
<i>Haemaniastrum spp.</i>	Co	Up to 1 %
Brake fern	As	>1.5% (Nature,409,2001)

Aquatic Hyperaccumulators

The water hyacinth (*Eichhornia crassipes*)

Rate of removal of heavy metals from aqueous phase		
Element	mg/g DW biomass/day	g/ha/day
Cd	0.67	400
Co	0.57	340
Pb	0.18	90
Hg	0.15	110
Ni	0.50	300
Ag	0.44	260

Gold Hill Mesa Corp. (CO Springs) - water hyacinth for removal of Au from Au tailings.

Sesbania drummondii



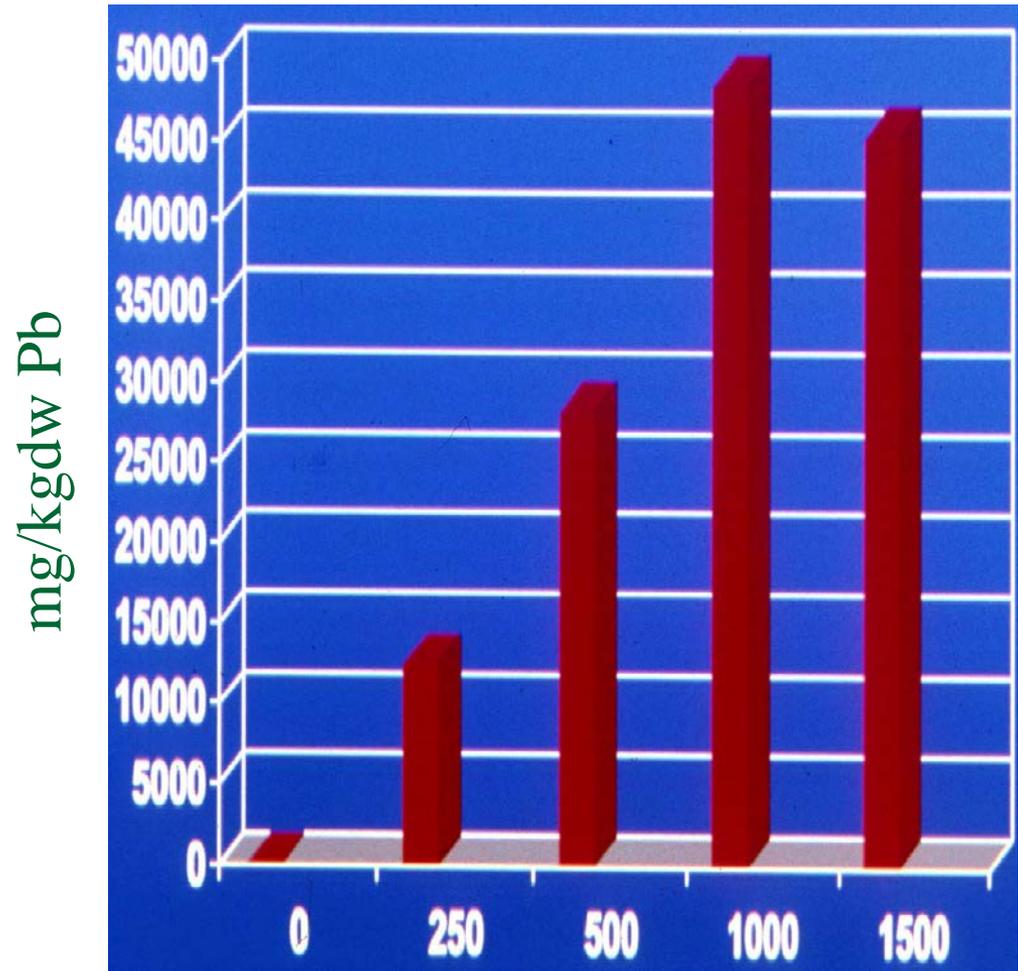
- **A high biomass plant**
- **Common name: Rattlebox**
- **Native to Southeastern U.S.**



Sesbania drummondii & Lead

Demonstrated as lead hyperaccumulator

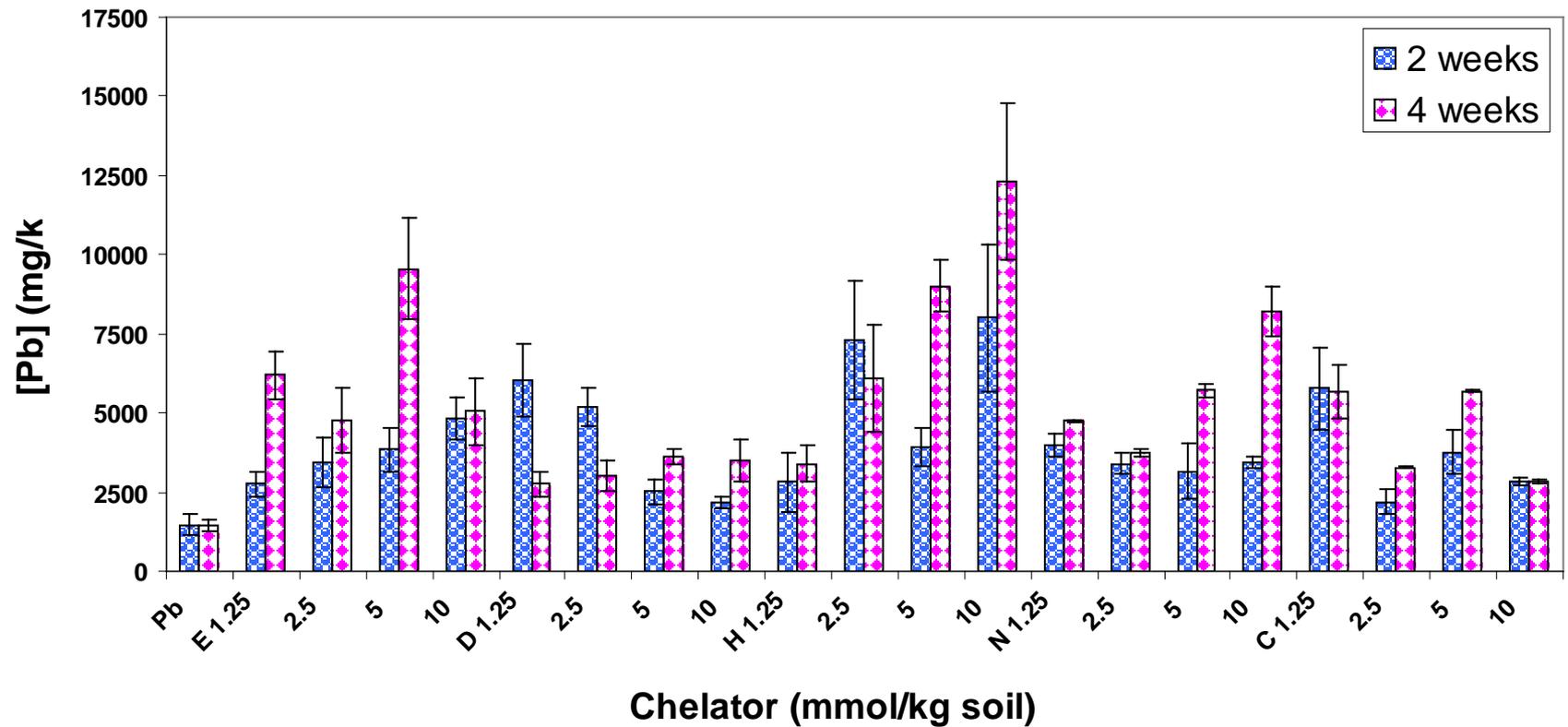
- Tolerates up to 1,000 ppm in hydroponic solution
 - Accumulated >4% (DW) Pb in shoots in hydroponic conditions
 - Roots showed 6% (DW) accumulation
 - EDTA and low pH increased accumulation further
- (EST 36, 4676-4680, 2002).



mg/L Pb

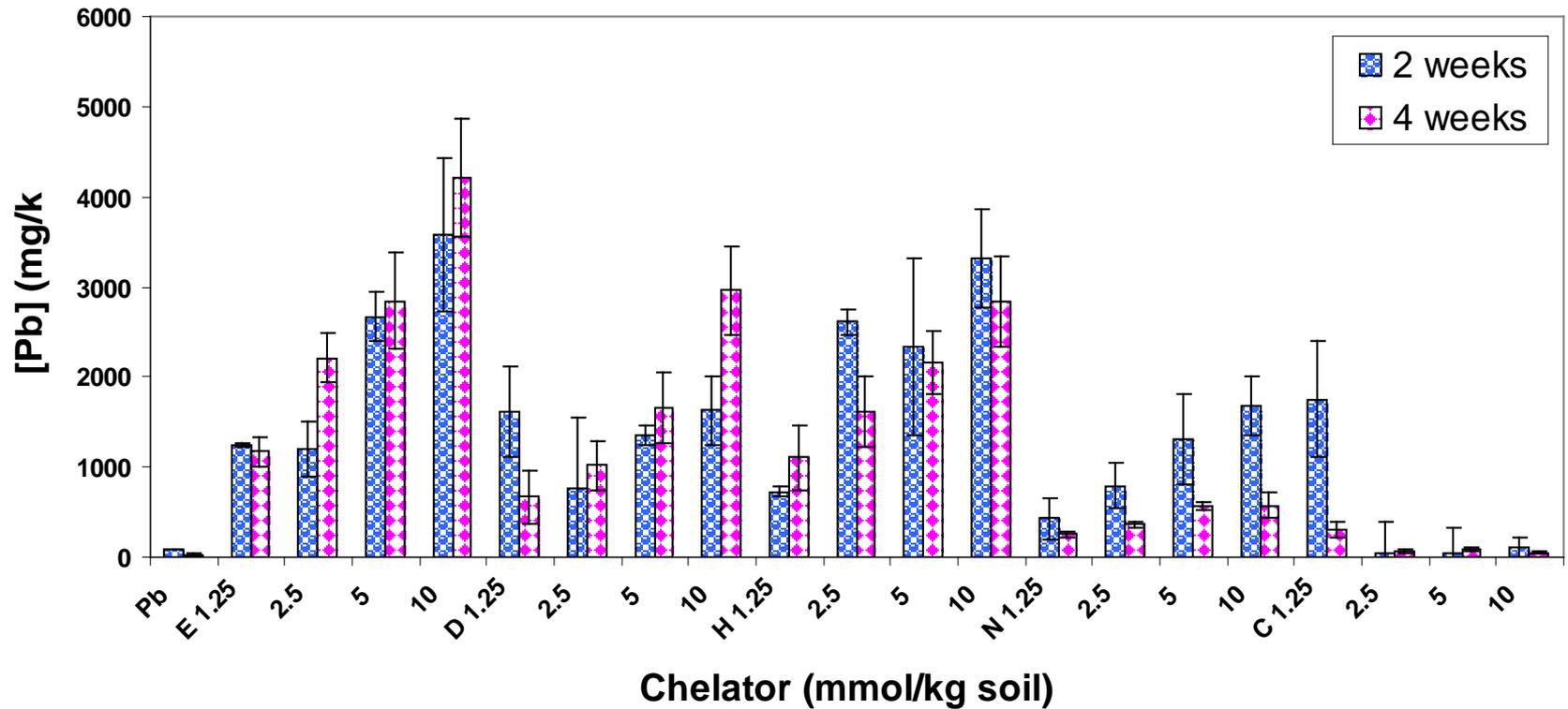
Sesbania in soil supplemented with Pb

Root Pb



Sesbania in soil supplemented with Pb

Shoot Pb

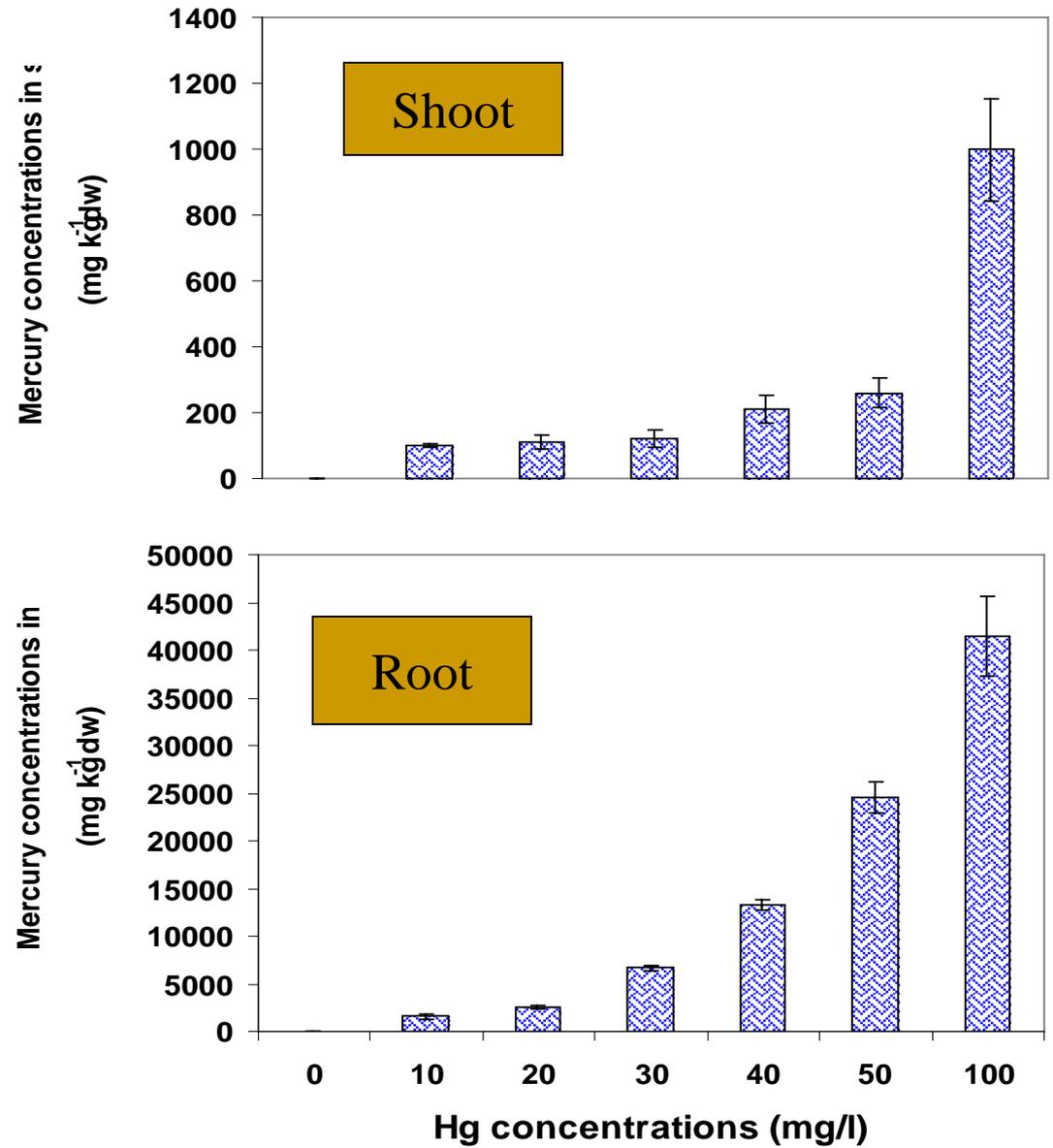


Estimated total Pb removed from soil by several plants

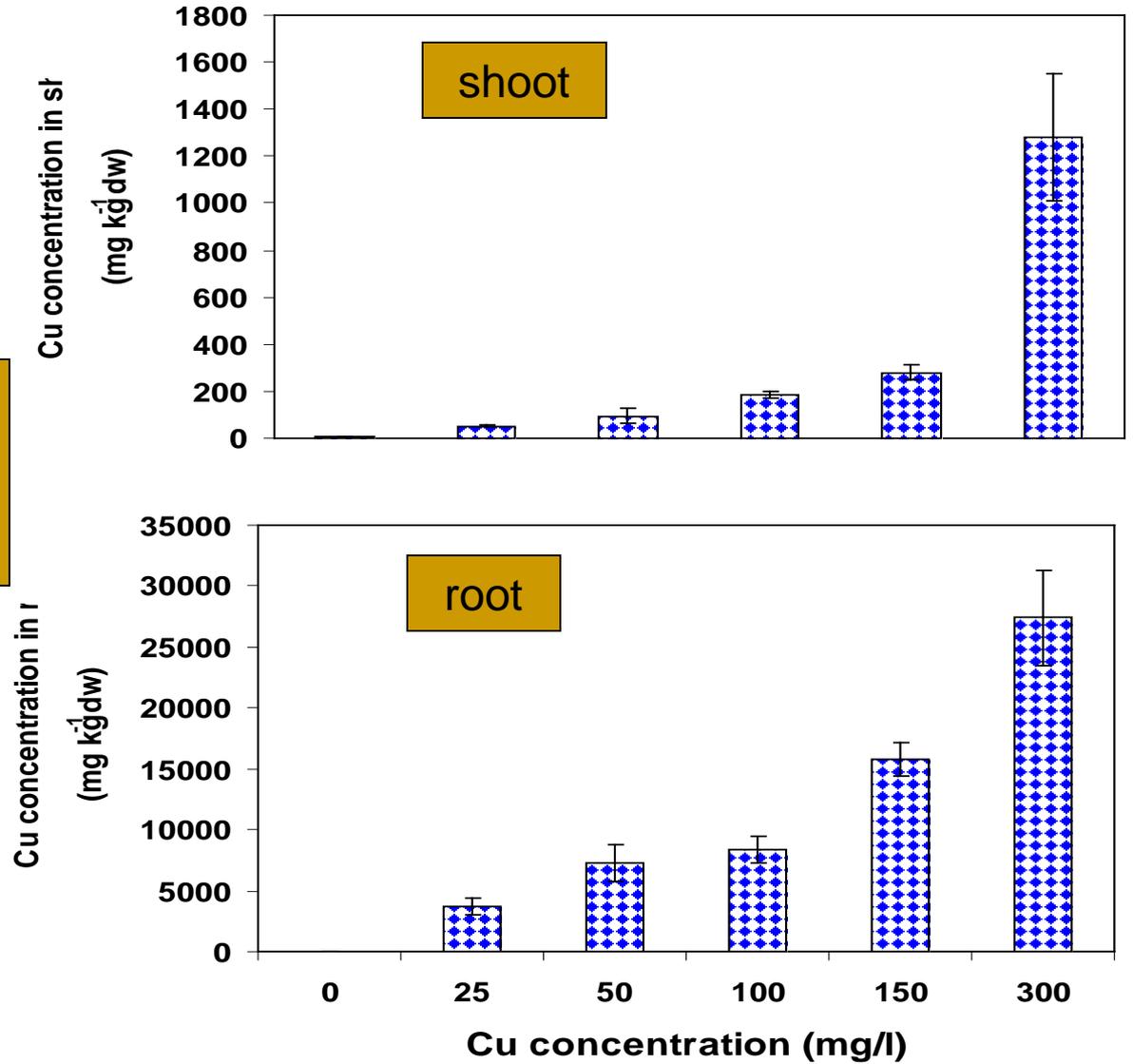
(Ruley 2004)

Species	Chelators	Soil Pb (mg/kg)	Shoot Pb (%)	Biomass (t/ha/yr)	Est. total Pb extr. (kg/ha/yr)	Source
<i>Zea mays</i>	5.8 mmol/kg HEDTA	2500	1.06	5-6	53-64	Huang et al. 1997
<i>Pisum sativum</i>	1.34 g/kg EDTA	2450	0.897	3-4	27-36	Huang et al. 1997
<i>Sesbania drummondii</i>	10 mmol/kg EDTA	7500	0.42	10-15	43-63	
<i>Brassica juncea</i>	10 mmol/kg EDTA	600	1.6	1-1.5	16-24	Blaylock et al. 1997
<i>Triticum aestivum</i>	5 mmol/kg EDTA+5 mmol/kg acetic acid	2000	0.92	2.5	23	Begonia et al. 2002

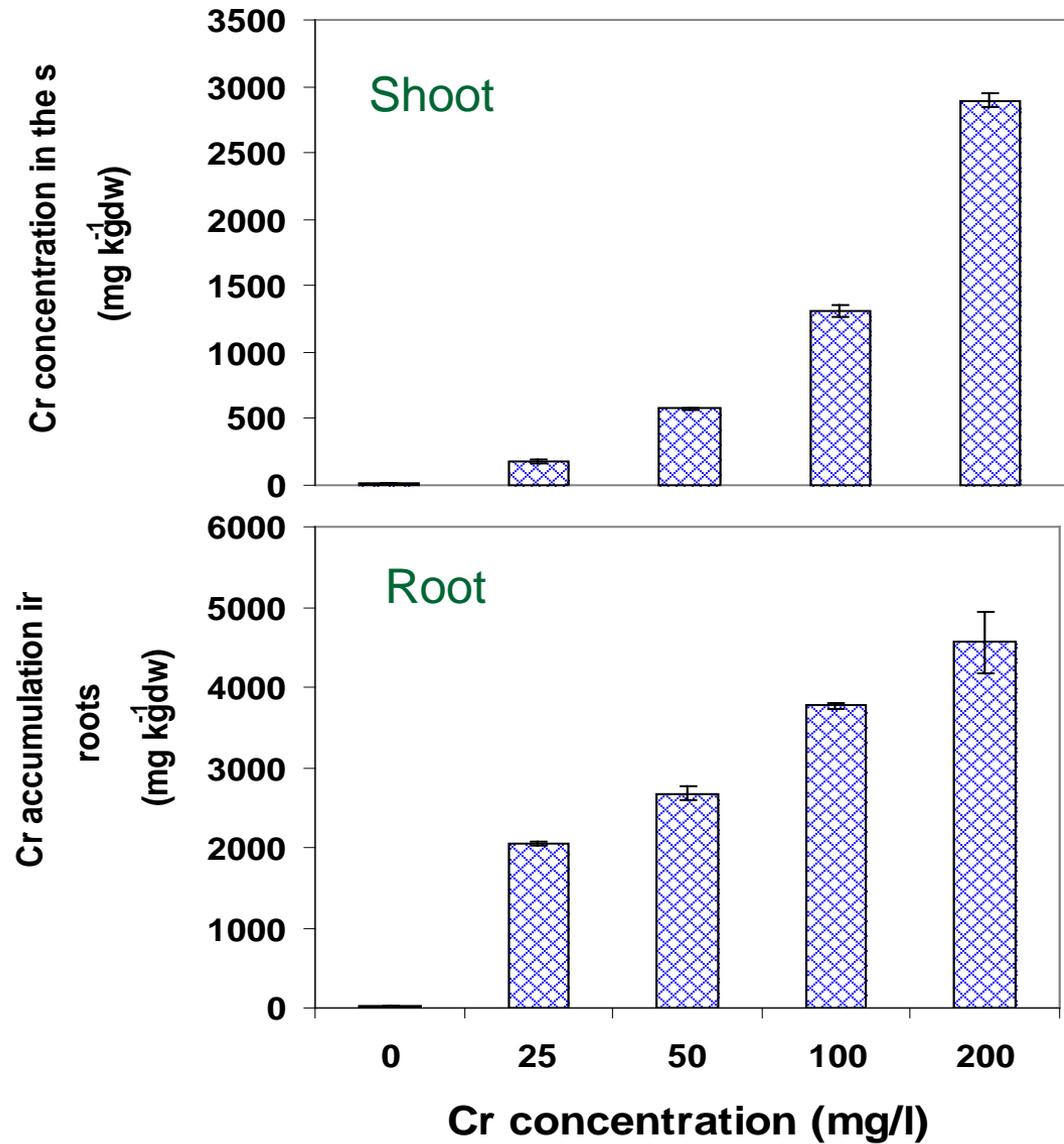
Mercury uptake by *Sesbania*



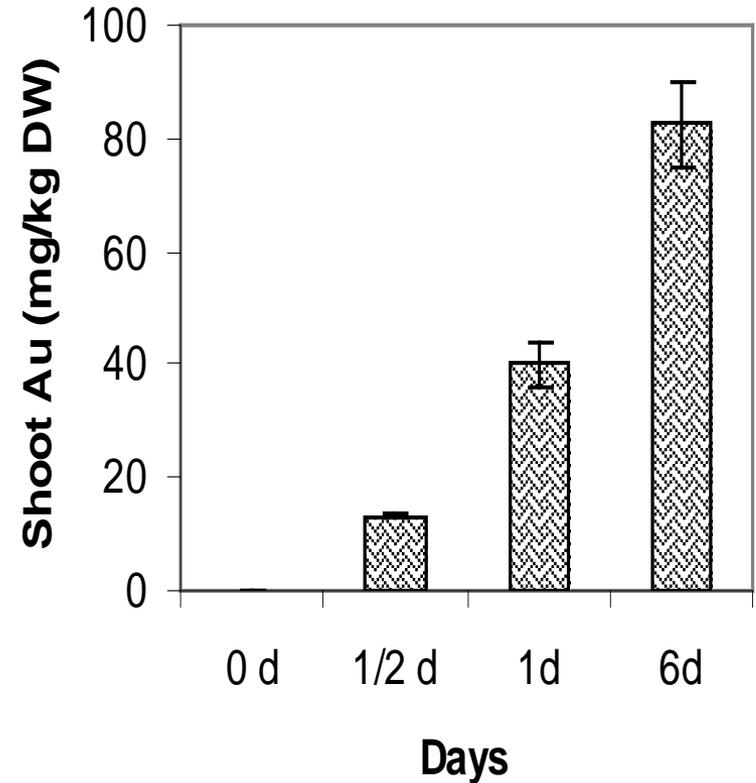
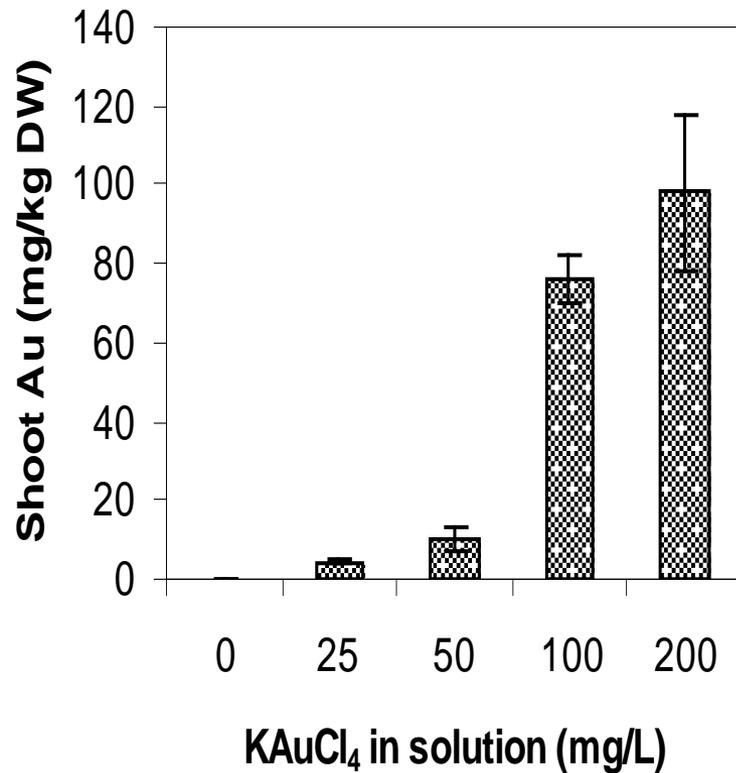
Copper uptake By *Sesbania*



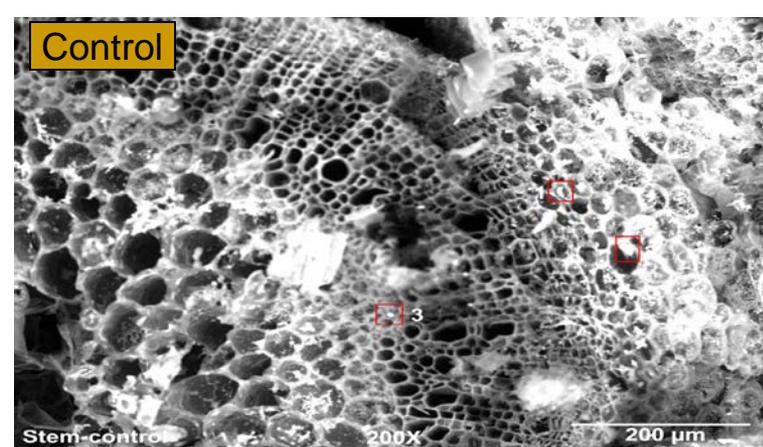
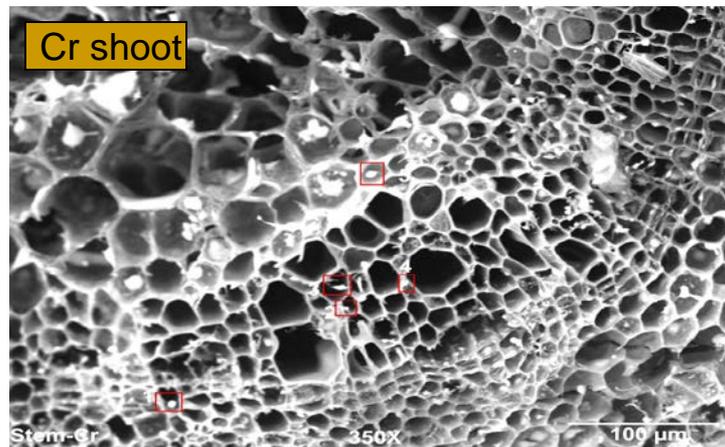
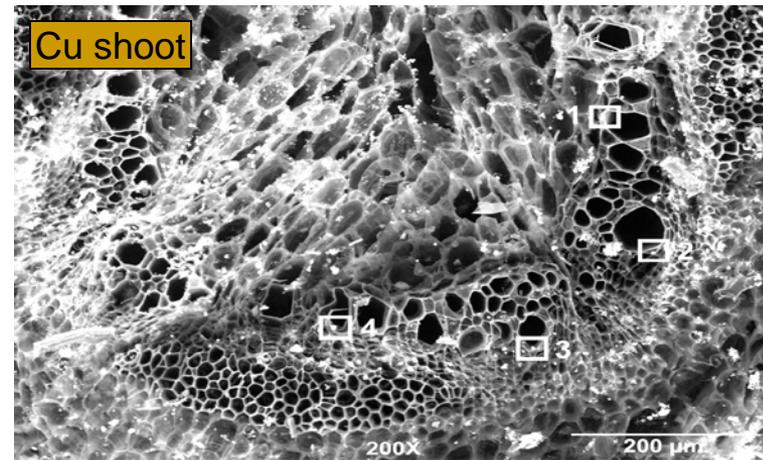
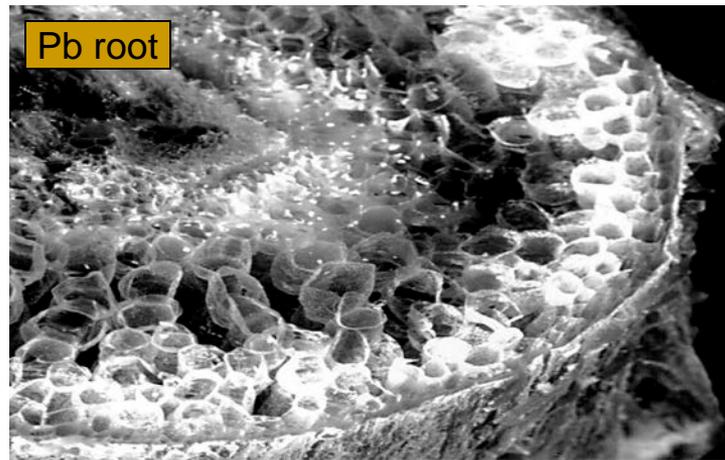
Chromium uptake By *Sesbania*



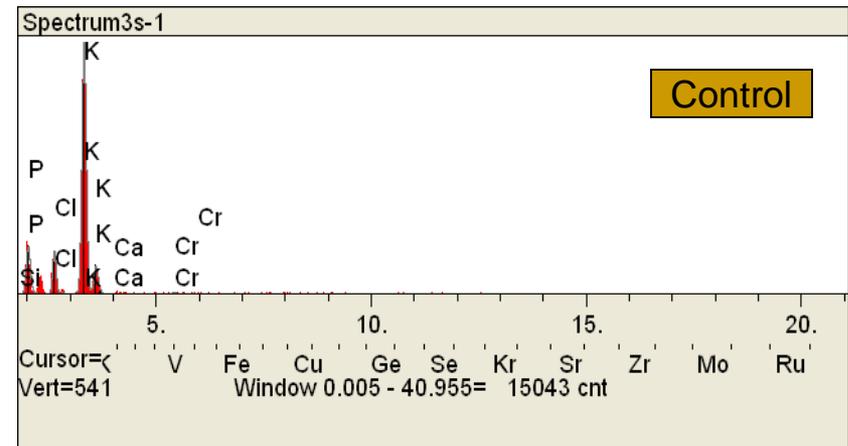
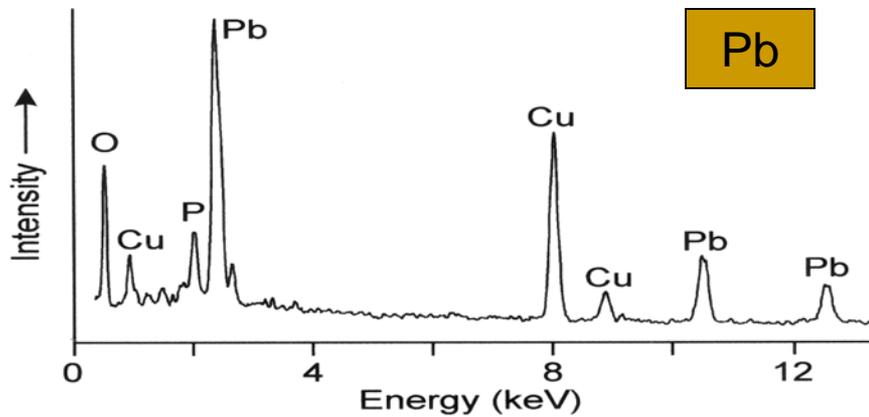
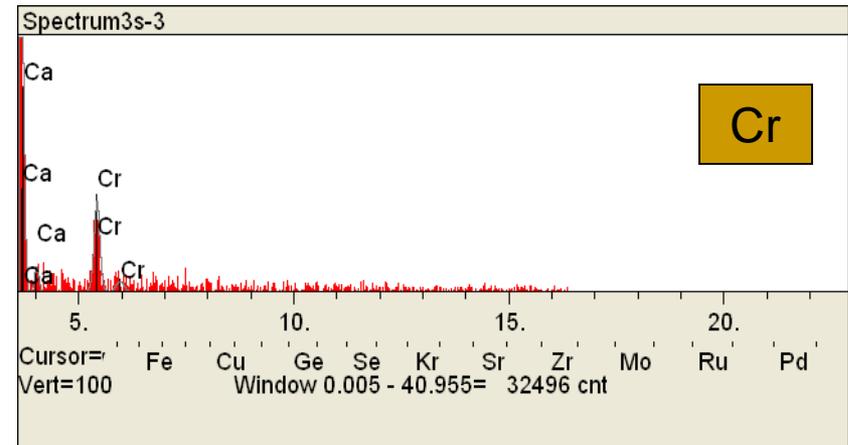
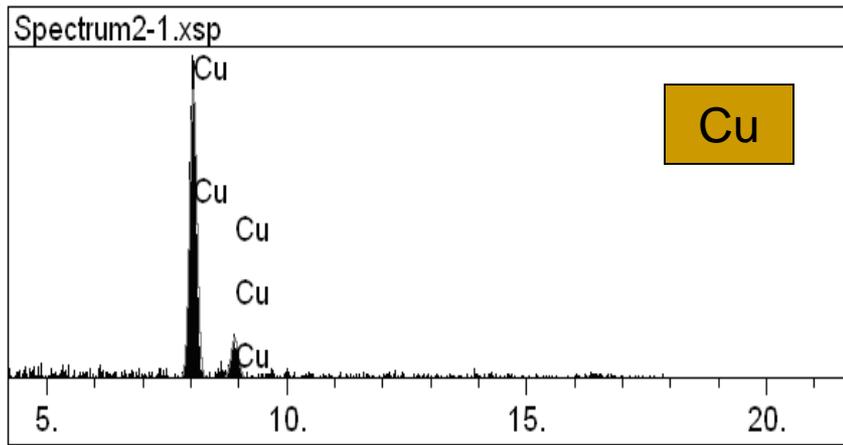
Gold uptake by *Sesbania*



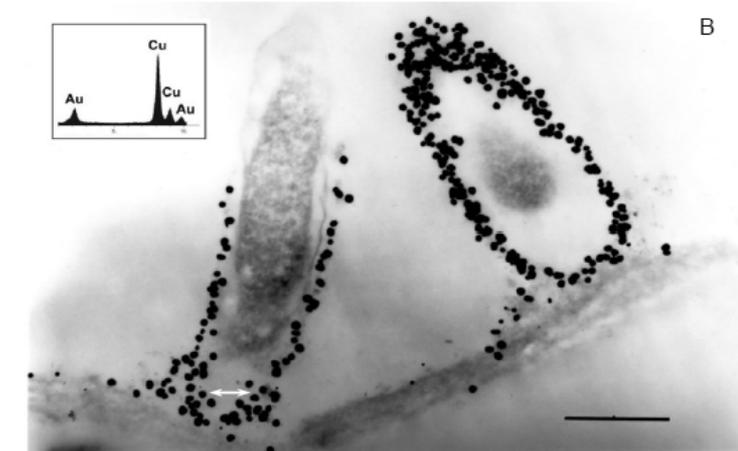
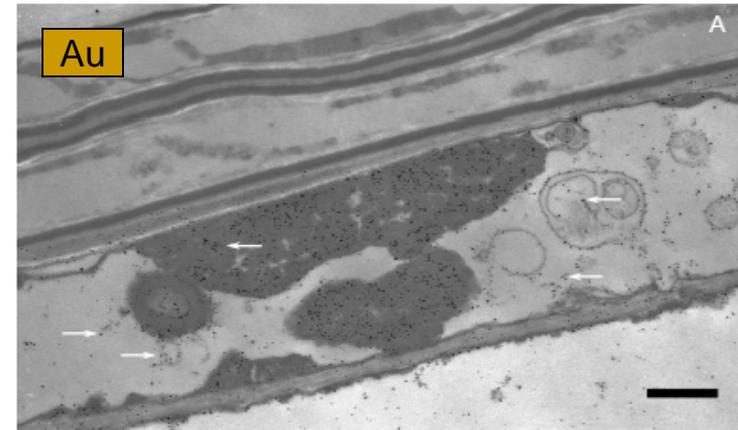
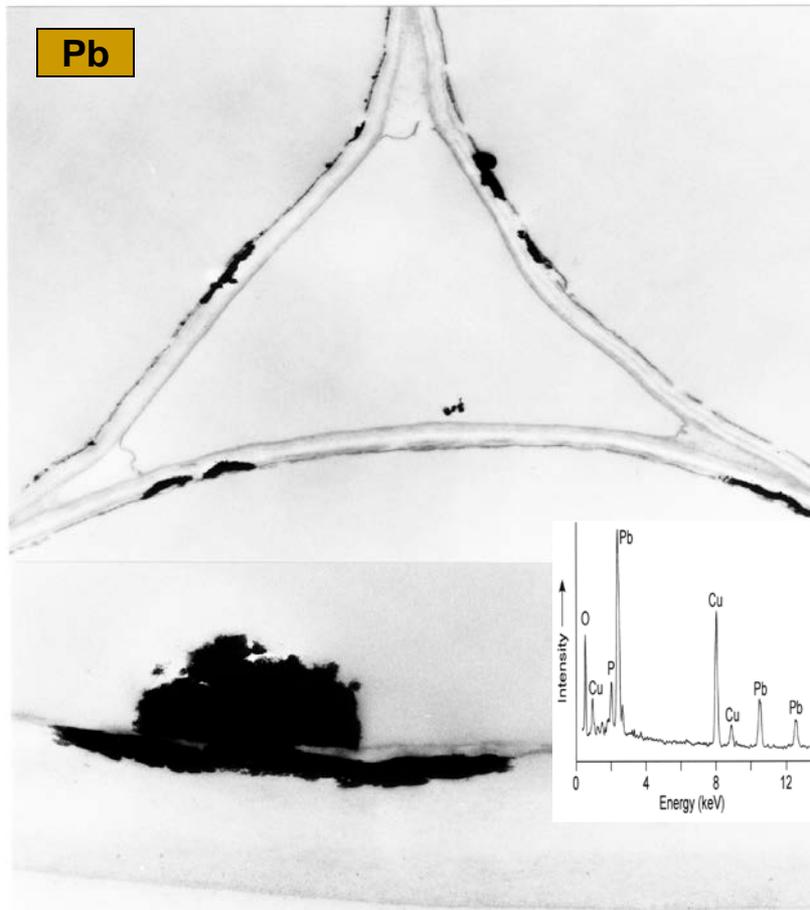
Scanning Electron Microscopy of Plant shoot grown in metals



X-ray microanalysis (EDS) of *Sesbania* tissue

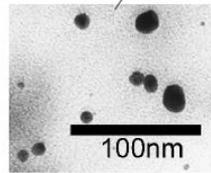
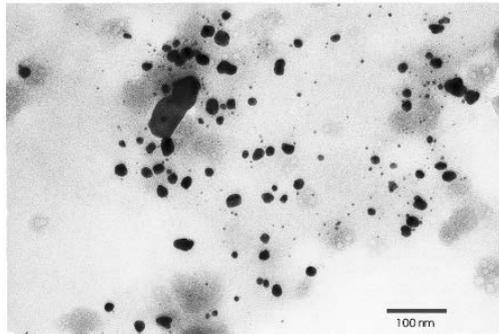


Transmission Electron Microscopy of *Sesbania* tissues with metals

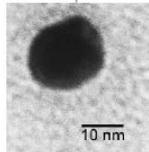
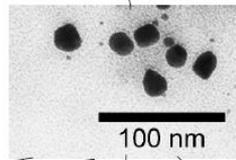


Gold nanoparticles in *Sesbania*

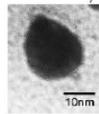
Gold Extraction



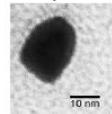
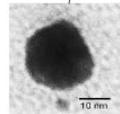
size distribution



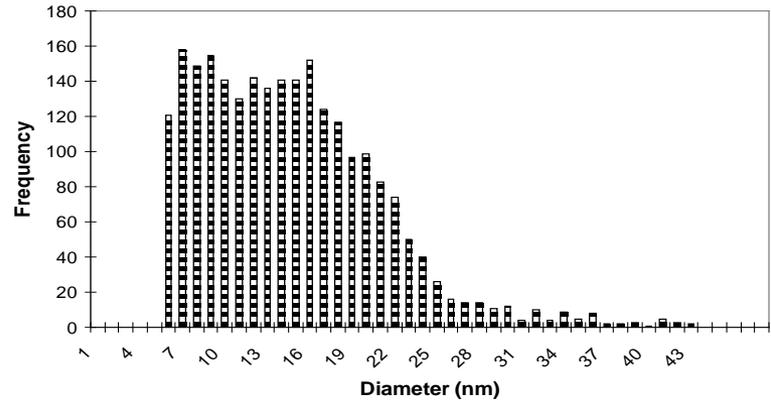
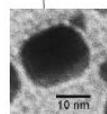
round



round



square



ES & T (accepted)

Transport of Pb in *Sesbania*

- Scanning Electron Microscopy
 - Transport of metals via different cell types
 - Transmission Electron Microscopy
 - Pb particles in intercellular spaces, cell membranes and cell walls.
 - Au particles (nanoparticles) are inside the cell.
 - Some deposits were also located within the tonoplast.
-

Biotransformation of Metals

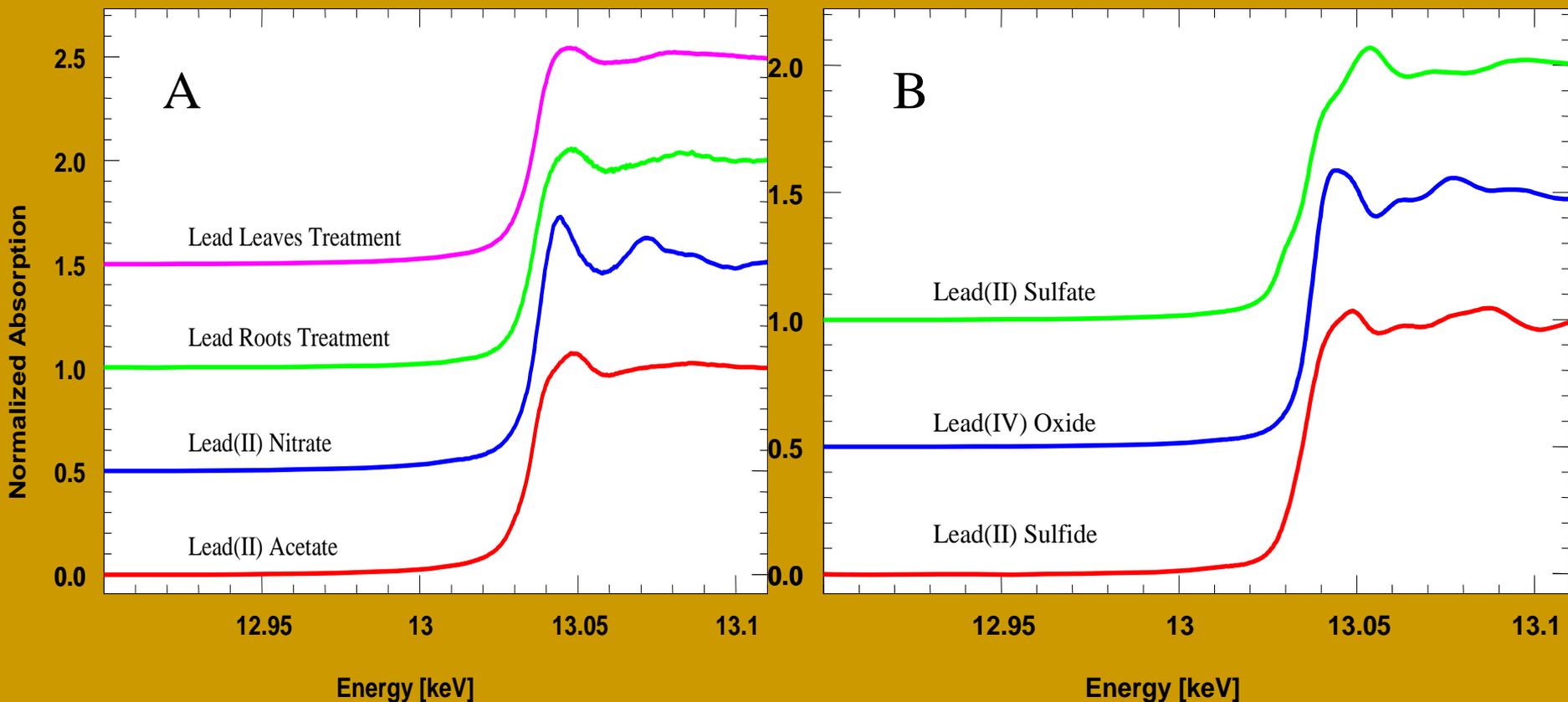
(Using XAS Technology)

Types of XAS

- **XANES** (X-ray absorption near edge structure)
 - determines the oxidation state and atomic geometry of a bound metal.
 - **EXAFS** (Extended X-ray absorption fine structure) – traces the ligand involved in the metal binding by measuring the distance from the X-ray-absorbing atom to the next nearest atom.
-

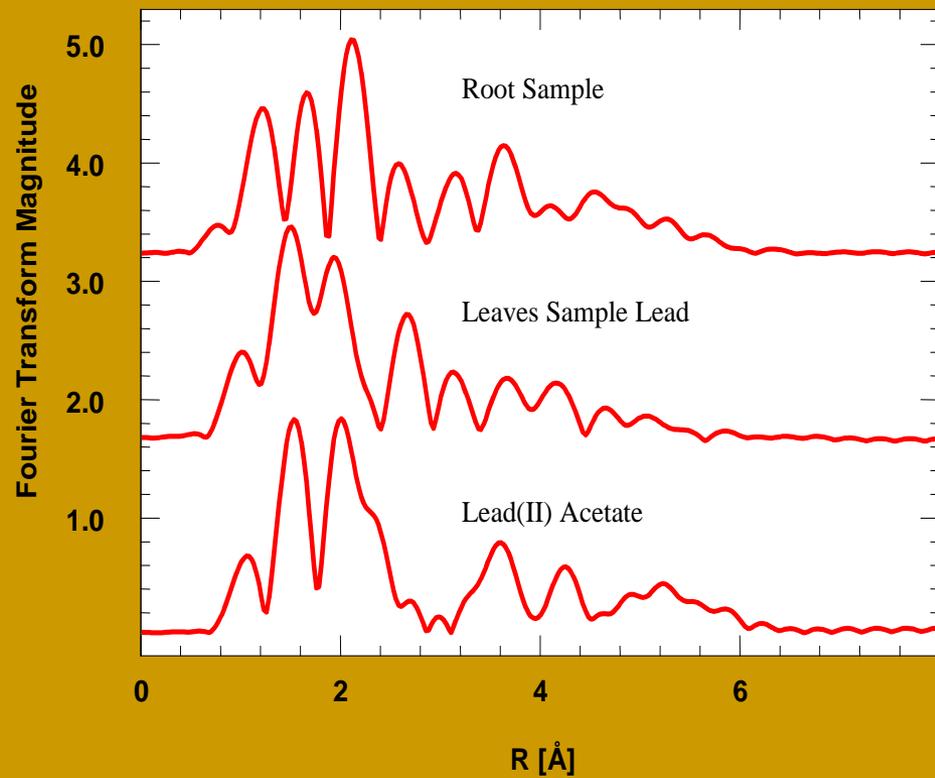
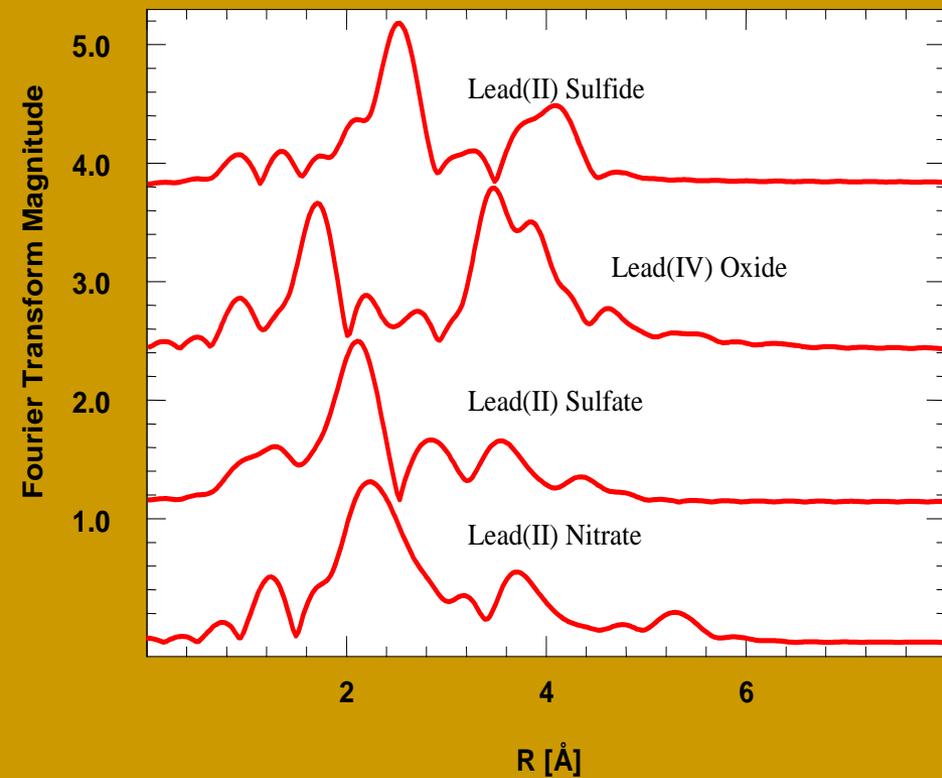
XANES Spectra of *Sesbania*

(ET & C 23, 2068, 2004)



A) L_{III} XANES of lead laden plant samples, lead(II) nitrate, and lead(II) acetate. L_{III} XANES of lead model compounds lead(II) sulfide, lead(II) sulfate, and lead(IV) oxide. **B)**

EXAFS Spectra of *Sesbania*



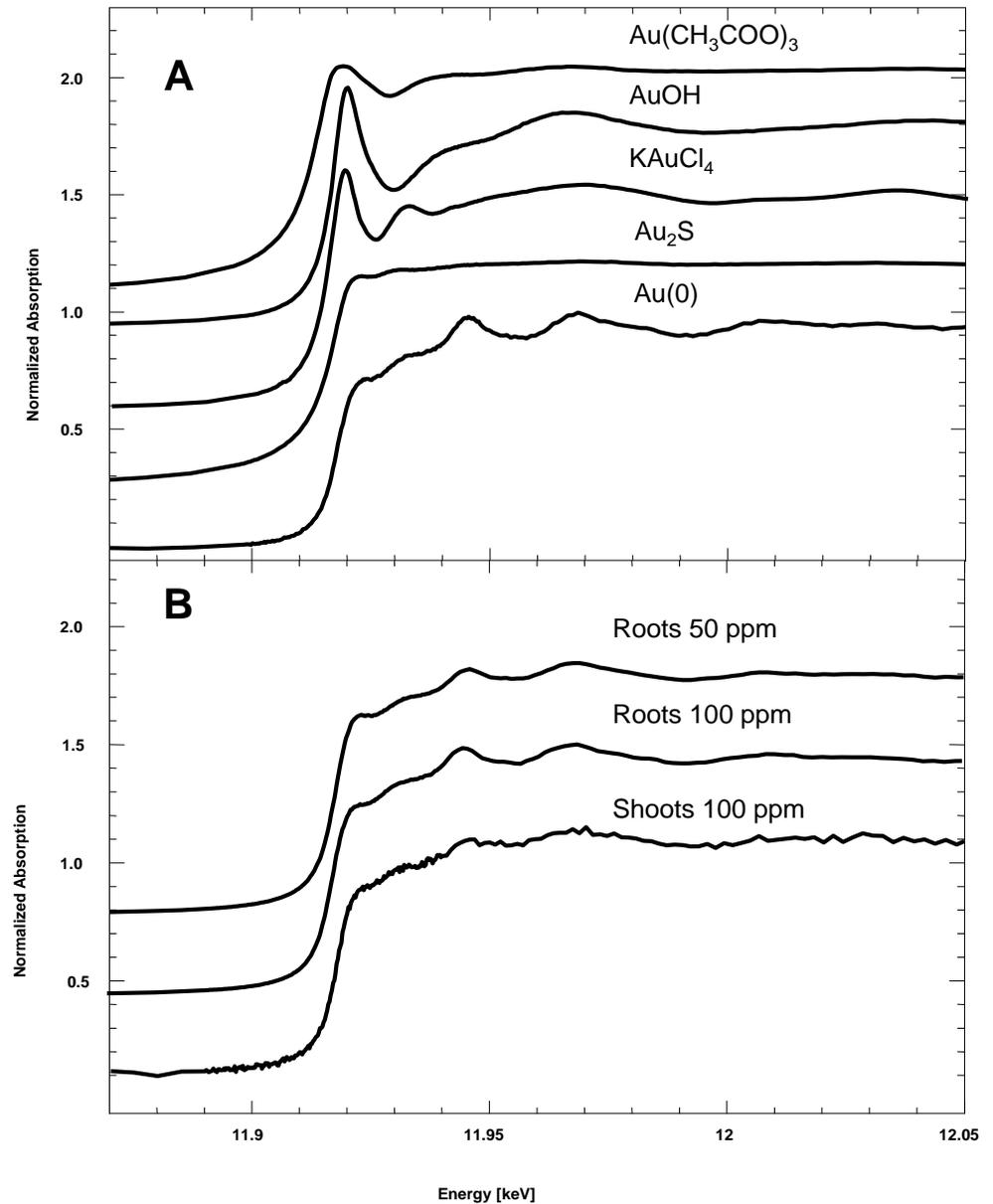
XANES and EXAFS data of Pb-treated *Sesbania*

(Environ. Toxicol. Chem. 23, 2068-2073, 2004)

Samples	Pb(NO ₃) ₂ %	PbSO ₄ %	Pb metal %	PbS %	Pb acetate %
Leaves	7.6	25.8	0	14.2	52.4
Roots	10.1	0	8.8	20.2	60.9

A. XANES of gold model compounds: gold acetate, gold hydroxide, potassium tetrachloroaurate, gold sulfide, and gold metal.

B. XANES of gold-laden plant samples.



XANES of gold accumulated in *Sesbania*

Samples	% KAuCl₄	% Au(CH₃COO)₃	% Au₂S	% Au(0)	% AuOH
Au 50 ppm roots	0	0	18.4	81.6	0
Au 100 ppm roots	0	0	16.4	83.6	0
Au 100 ppm shoots	0	0	14.2	84.4	1.4

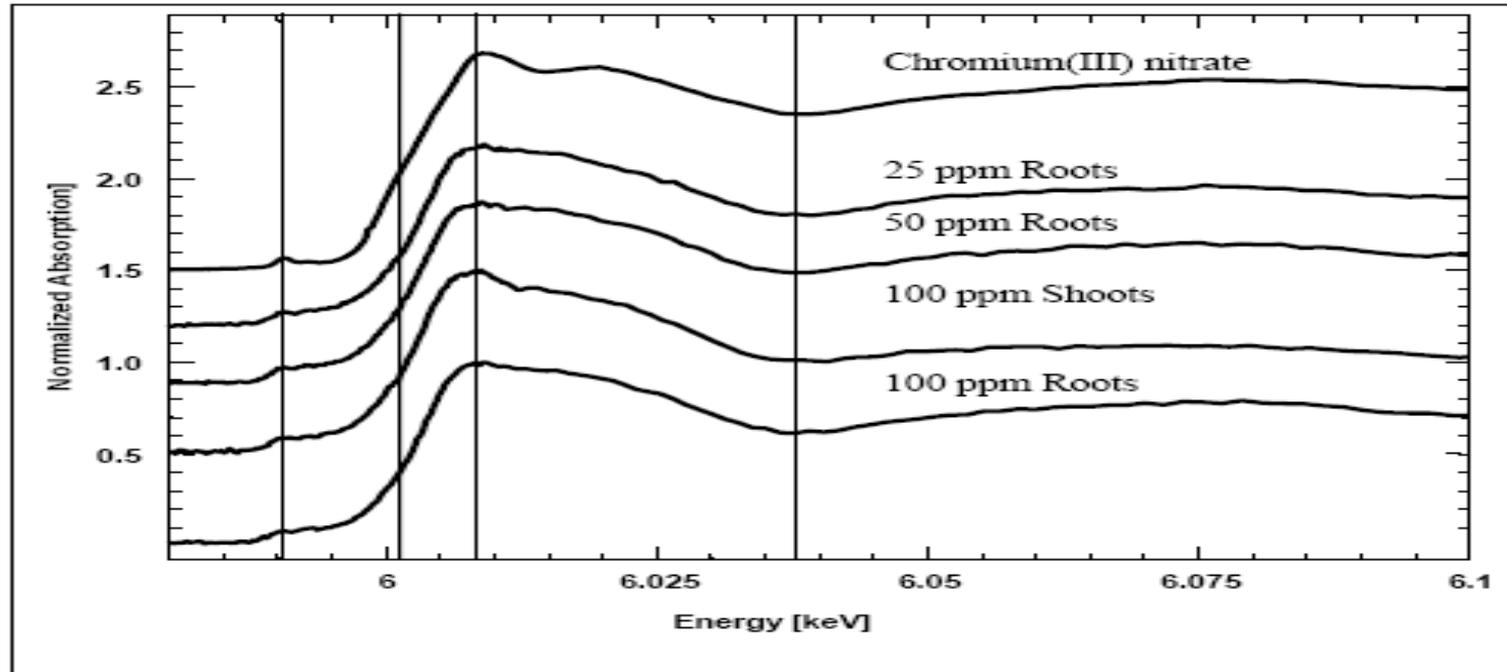
XANES of Cu accumulated in *Sesbania*

(Chemosphere 67, 2257-66, 2007)

Samples	% copper(II) acetate	% copper(II) nitrate	% copper(II) phthalocyanine	% copper(II) gluconate	% copper(II) Oxide
Cu 25 mg l⁻¹	0	20	2	57	21
Cu 100 mg l⁻¹	30	14	5	18	33

Biotransform cupric sulfate to copper(II) sugar/small organic acid complex and acetate in its tissue

XANES analysis of Cr in *Sesbania*

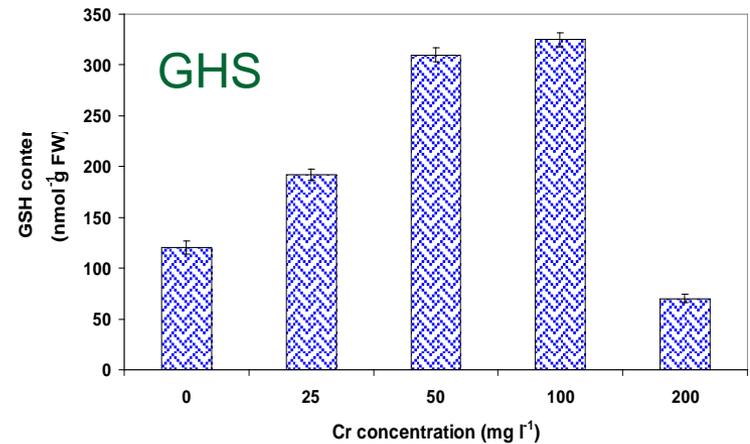
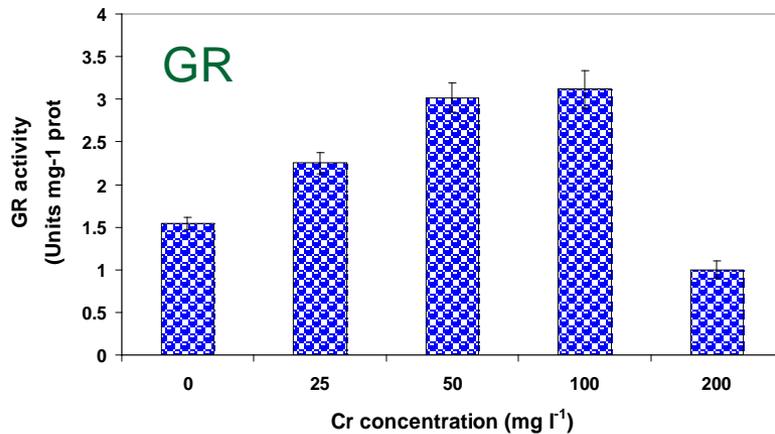
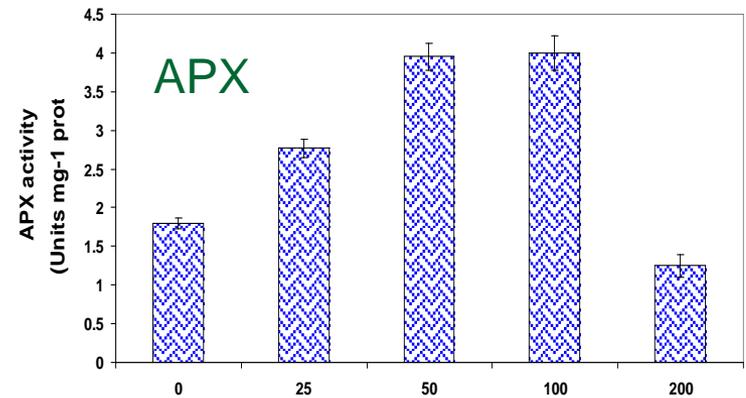
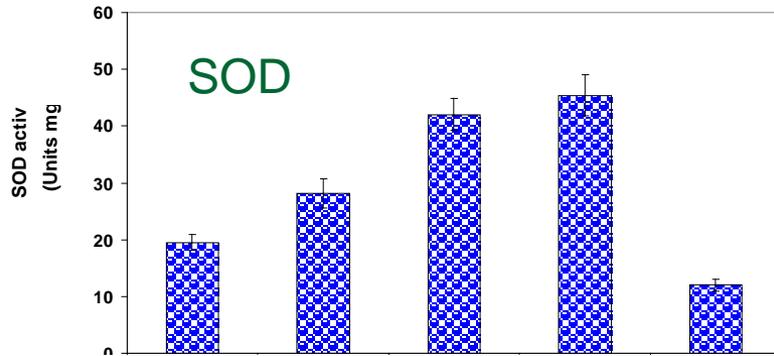


Sesbania has capability to biotransform
Cr(VI) in to Cr(III) in its tissue

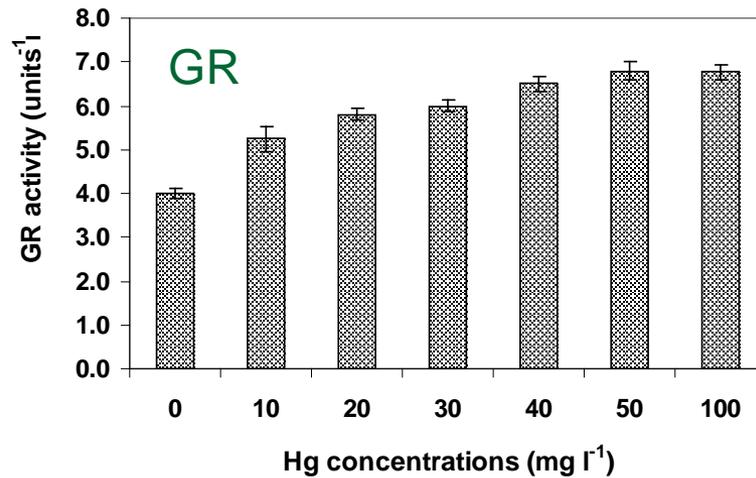
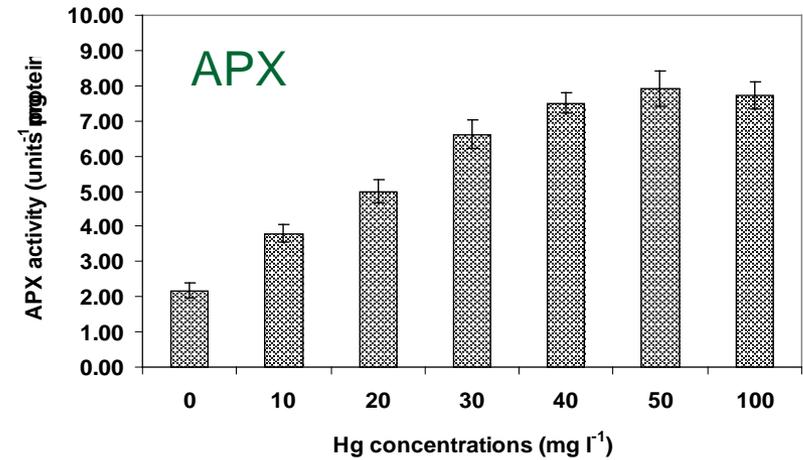
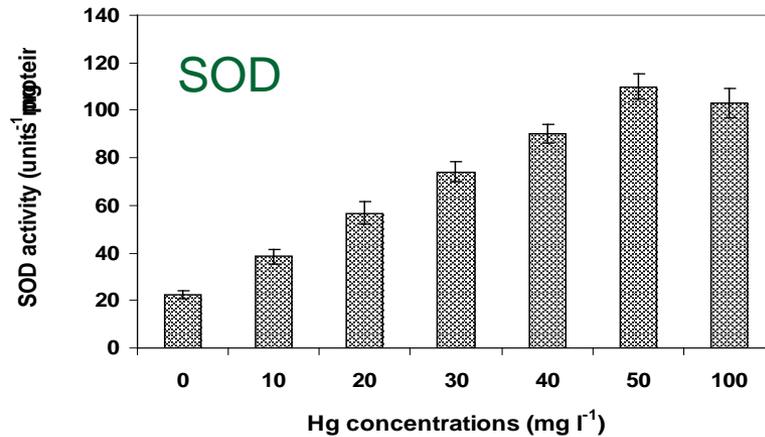
Antioxidant Reactions & metal Stress in *Sesbania*

- Generally metal exposure triggers an increase in activity of antioxidant enzymes.
 - **Superoxide dismutase (SOD)** – catalyze dismutation of superoxide radicals to hydrogen peroxide & oxygen
 - **Catalase (CAT)** – catalyzes decomposition of hydrogen peroxide to water and oxygen
 - **Ascorbate peroxidase (APX)** – detoxifies hydrogen peroxide to water using ascorbate as substrate
 - **Glutathione reductase (GR)** – reduces oxidized glutathione (GSSG) to reduced glutathione (GSH)
 - maintains high GSH/GSSH to sustain role of GSH as anti-oxidant
 - also incorporating into phytochelatins
 - GSH also function as free radical scavenger

Stress enzymes in *Sesbania* in Cr

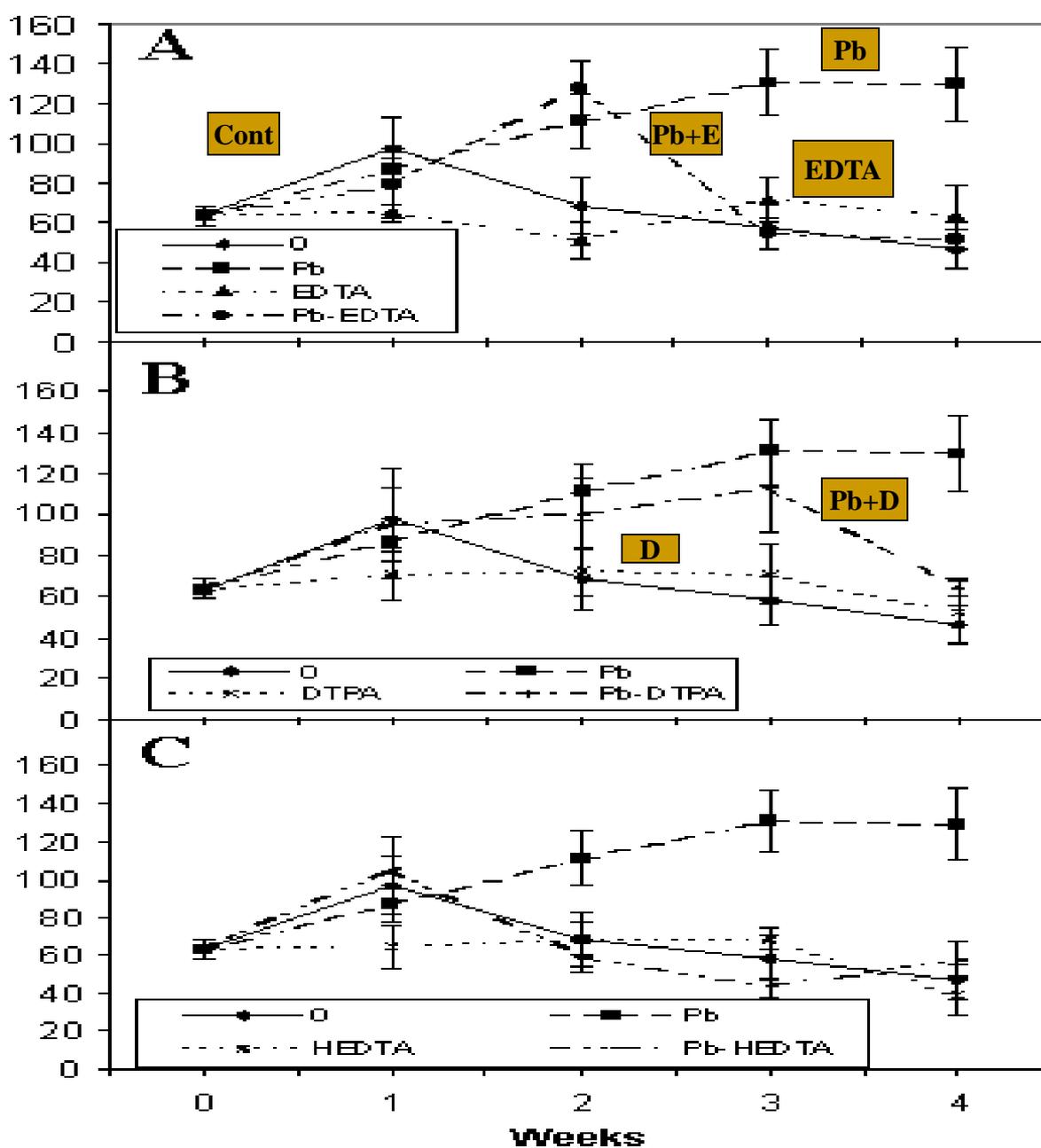


Stress enzymes in *Sesbania* in Hg

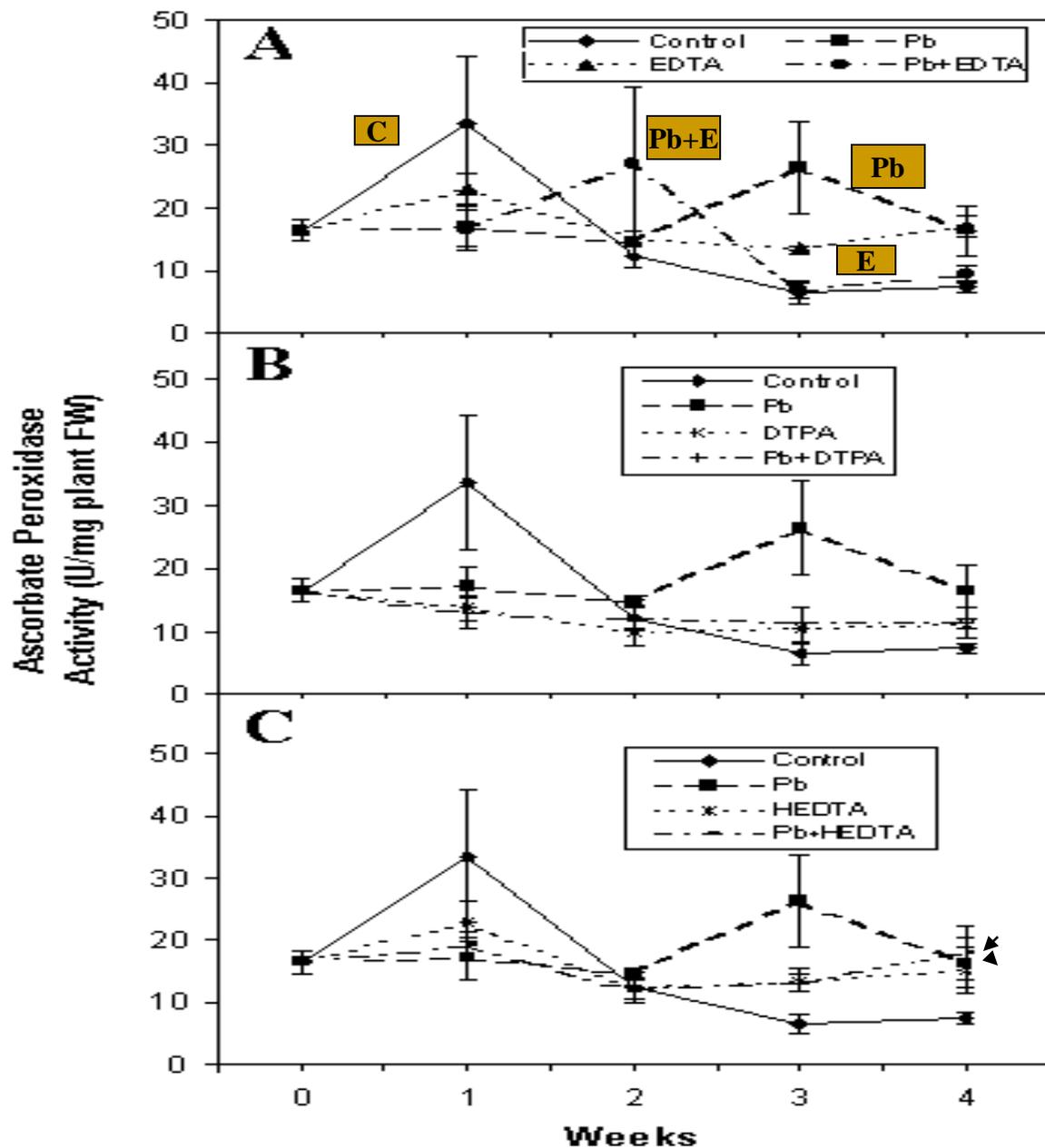


Superoxide dismutase (SOD)

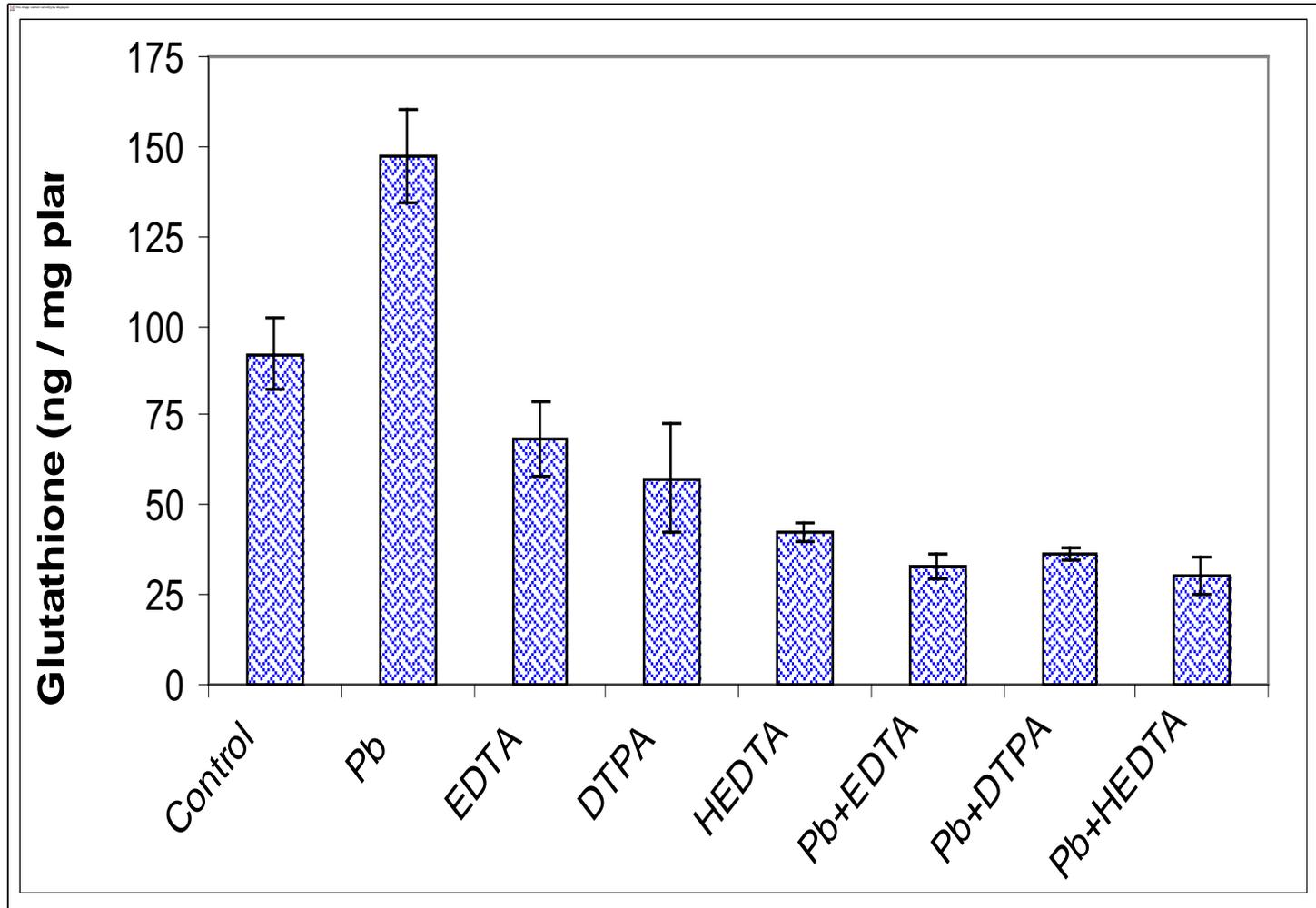
Superoxide Dismutase Activity (U/mg plant FW)



Ascorbate Peroxidase (APX)

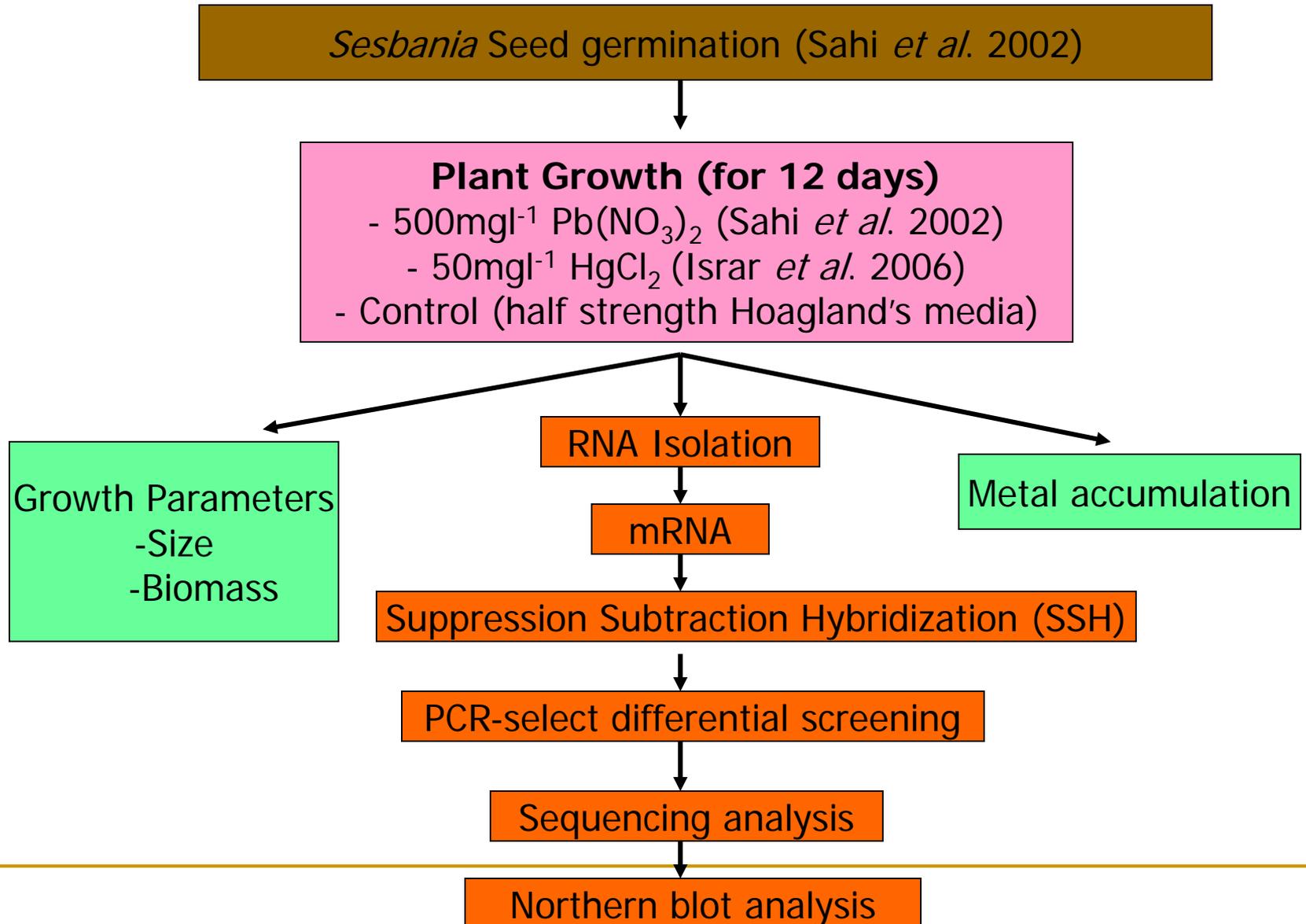


Glutathione content



Identification of lead and mercury responsive genes

Experimental Design



Suppression subtraction hybridization (SSH)

- Based on the technique called suppression PCR
 - Compare two populations of mRNA
 - Obtain clones of genes that are expressed in one population but not in the other
-

Sequencing results for Pb samples

- 63 clones corresponds to unigenes
 - 49 (78 %) identified as segments of cDNAs contained in GenBank database
 - 14 (22 %) were unknown (no similarity)
 - Clone # 7 exhibited homology to **type 2 metallothionein** sequences
-

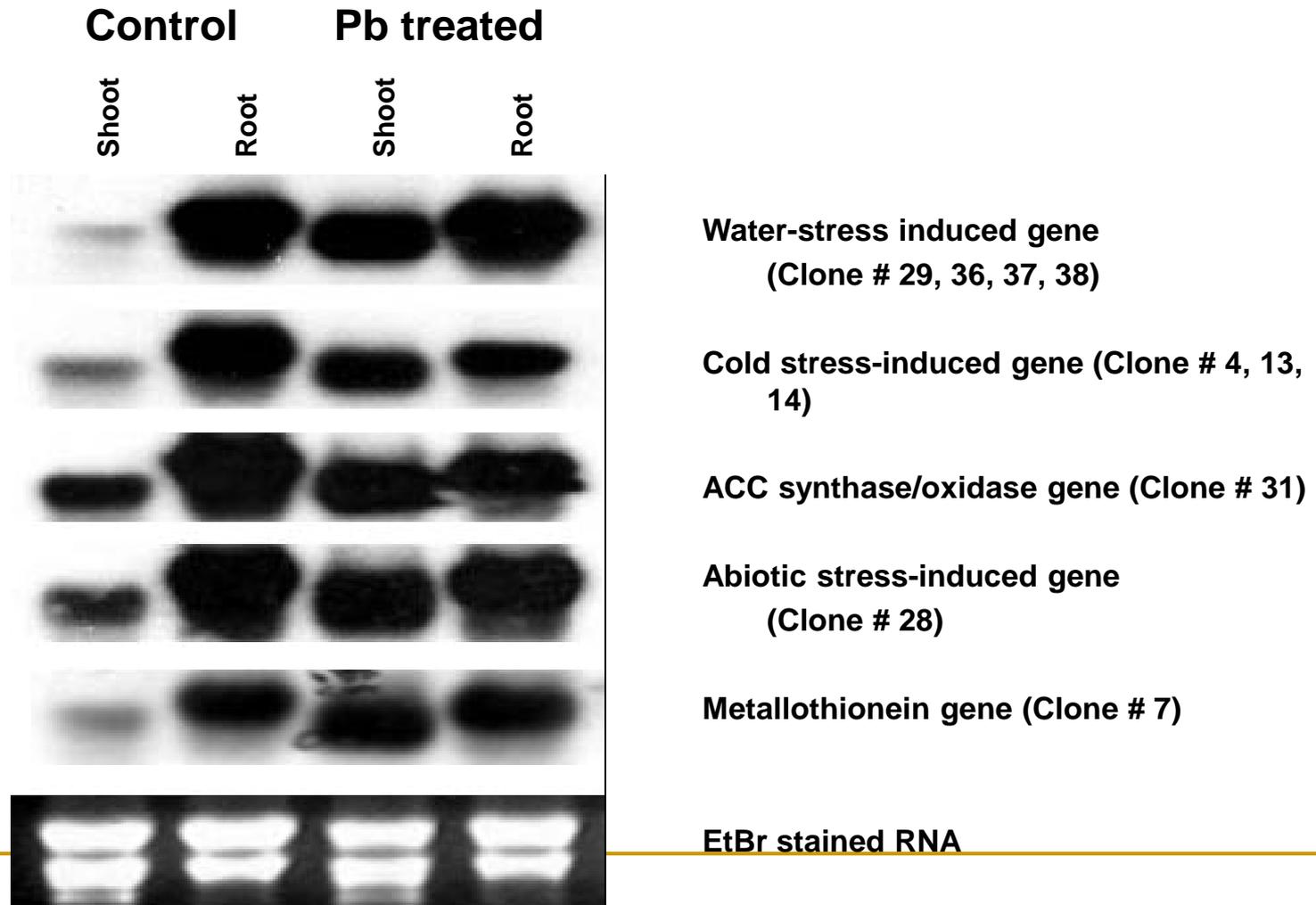
Clone	Accession number	Length (bp)	Homology ^a	E-value
SSH-1	DQ465754	183	<i>Acanthopanax sessiliflorus</i> cDNA library <i>Eleutherococcus sessiliflorus</i> cDNA, mRNA sequence (CF923918)	0.0
SSH-2	DQ465755	293	Apple_EST_Mdas Malus x domestica cDNA similar to dbj BAB33421.1 putative senescence-associated protein [<i>Pisum sativum</i>], mRNA sequence (DR993778)	1e ⁻¹⁵⁹
SSH-3	DQ465756	282	CabSau Flower Stage 12 (FLOu0012) <i>Vitis vinifera</i> cDNA clone VVI101F09 5, mRNA sequence (DT015551)	0.0
SSH-4	DQ465757	666	Cold stressed <i>Glycine clandestina</i> SSH cDNA clone Gc02_03a05, mRNA sequence (BG838800)	0.0
SSH-5	DQ465758	569	<i>Phaseolus vulgaris</i> seedling EST Library inoculated with anthracnose-PVEPSE3029E14 5', mRNA sequence (CB543340)	0.0
SSH-6	DQ465759	565	<i>Phaseolus vulgaris</i> seedling EST Library inoculated with anthracnose-cDNA clone PVEPSE3030N16 5', mRNA sequence (CB543682)	0.0
SSH-7	DQ465760	414	Type 2 Metallothionein-Cytochrome P450 like_TBP [<i>Citrullus lanatus</i>] (AB182926)	0.0
SSH-8	DQ465761	620	Cytochrome P450 like_TBP [<i>Nicotiana tabacum</i>] (BAA10929)	0.0
SSH-9	DQ465762	313	<i>Glycine max</i> cDNA clone Gm-c1086-27 5' similar to CYTOCHROME P450 LIKE_TBP mRNA sequence (BM091724)	1e ⁻¹⁷⁴
SSH-10	DQ465763	366	<i>Glycine max</i> cDNA, mRNA sequence (BE660497)	0.0
SSH-11	DQ465764	478	<i>Glycine max</i> cDNA, mRNA sequence (BU927378)	0.0
SSH-12	DQ465765	692	<i>Glycine soja</i> cDNA clone SOYBEAN CLONE ID: Gm-c1056-3170 5', mRNA sequence (CA799399)	0.0
SSH-13	DQ465766	739	<i>Glycine max</i> cold stressed leaves cDNA clone Gm01_16d09, mRNA sequence (BG839363)	0.0
SSH-14	DQ465767	674	<i>Glycine max</i> cold stressed leaves cDNA clone Gm01_17a09, mRNA sequence (BG839403)	0.0
SSH-15	DQ465768	471	Gmax SC <i>Glycine max</i> cDNA, mRNA sequence (BE660497)	0.0
SSH-16	DQ465769	875	<i>Gossypium hirsutum</i> cDNA clone GH_CHX12C18 3', mRNA sequence (DT462491)	0.0
SSH-17	DQ465770	840	hemolysin [<i>Acanthamoeba polyphaga</i>] (AAA58585)	0.0
SSH-18	DQ465771	666	<i>Heterobasidion annosum</i> - Scots pine infection stage (HAGE) subtraction cDNA clone hage001aD09, mRNA (BQ789710)	2e ⁻⁸⁵
SSH-19	DQ465772	295	Leafy spurge subtractive cDNA libraries <i>Euphorbia esula</i> cDNA clone RTP5O15 5', mRNA sequence (DT639472)	1e ⁻¹⁵⁸
SSH-20	DQ465773	633	<i>Lotus japonicus</i> nodule library 5 and 7 week-old <i>Lotus corniculatus</i> var. japonicus cDNA 5', mRNA sequence (AW720640)	0.0
SSH-21	DQ465774	299	<i>Medicago truncatula</i> cDNA clone MfTA01F19S6, mRNA sequence (AJ847433)	1e ⁻¹⁴⁹
SSH-22	DQ465775	341	<i>Medicago truncatula</i> cDNA clone MfTA09L24S6, mRNA sequence (AJ847823)	1e ⁻¹⁷⁷
SSH-23	DQ465776	522	Methyl Jasmonate-Elicited mRNA sequence from Root Cell Suspension Culture <i>Medicago truncatula</i> (CX533136)	0.0
SSH-24	DQ465777	137	<i>Mimulus guttatus</i> cDNA clone 0048P0008Z, mRNA sequence (CV515336)	9e ⁻⁴⁵
SSH-25	DQ465778	314	<i>Phaseolus vulgaris</i> leaf EST library cDNA clone PV_GEA0013C_C03.b1 5', mRNA sequence (CV530371)	1e ⁻¹⁸⁵
SSH-26	DQ465779	628	<i>Phaseolus vulgaris</i> leaf EST library cDNA clone PV_GEA0015C_G10.b1 5', mRNA sequence (CV531021)	0.0
SSH-27	DQ465780	229	<i>Populus trichocarpa</i> cDNA clone WS02553_I06 3', mRNA sequence (DT493138)	1e ⁻¹²⁷
SSH-28	DQ465781	899	Potato abiotic stress cDNA library <i>Solanum tuberosum</i> cDNA clone POAD792 5' end, mRNA sequence (CK272883)	0.0
SSH-29	DQ465782	900	Water stressed gmutDrNS01_32 <i>Glycine max</i> cDNA 3', mRNA sequence (CX711410)	0.0

SSH-30	DQ465783	265	Probable cytochrome P450 monooxygenase - maize (fragment) (T02955)	0.0
SSH-31	DQ465784	666	Putative ACC synthase/oxidase gene (BAB33421)	0.0
SSH-32	DQ465785	255	rRNA promoter binding protein [<i>Rattus norvegicus</i>] (NM147136)	1e ⁻¹⁴³
SSH-33	DQ465786	657	<i>Sesbania rostrata</i> root primordia cDNA clone SSH-10, mRNA sequence (AJ301742)	0.0
SSH-34	DQ465787	162	Subtracted cDNA library of maize inbred line H95-Rp1-Kr1N <i>Zea mays</i> cDNA clone Kr1N-4_D09, mRNA sequence (CA452627)	1e ⁻²⁶
SSH-35	DQ465788	531	Unknown protein (<i>Schistosoma japonicum</i>) (AA330301)	0.0
SSH-36	DQ465789	889	Water stressed gmrtDrNS01_28 Glycine max cDNA 3', mRNA sequence (CX711160)	0.0
SSH-37	DQ465790	874	Water stressed gmrtDrNS01_30 Glycine max cDNA 3', mRNA sequence (CX548993)	0.0
SSH-38	DQ465791	446	Water stressed gmrtDrNS01_31 Glycine max cDNA 3', mRNA sequence (CX707998)	0.0
SSH-39	DQ465792	289	Unnamed protein product [<i>Kluyveromyces lactis</i> NRRL Y-1140] (CAH00932)	5e ⁻⁹¹
SSH-40	DQ465793	648	CYTOCHROME P450 monooxygenase (EC 1.14.14.1) - common tobacco (0.0
SSH-41	DQ465794	881	Hypothetical protein [<i>Oryza sativa</i> (japonica cultivar-group)] (BAD46202)	0.0
SSH-42	DQ465795	357	26S ribosomal protein	0.0
SSH-43	DQ465796	288	Hypothetical protein GLP_748_1200_211 [<i>Giardia lamblia</i> ATCC 50803] (XP767406)	1e ⁻¹³⁷
SSH-44	DQ465797	371	Hypothetical protein UM05244.1 [<i>Ustilago maydis</i> 521] (XP761391)	1e ⁻¹⁸²
SSH-45	DQ465798	293	Unknown protein	1e ⁻¹⁷³
SSH-46	DQ465799	286	Unknown protein	1e ⁻¹⁶⁵
SSH-47	DQ465800	330	Unknown protein	1e ⁻¹⁶²
SSH-48	DQ465801	285	Unknown protein	1e ⁻¹⁵³
SSH-49	DQ465802	292	Unknown protein	1e ⁻¹⁶⁷
SSH-50	DQ465803	178	No homology ^b	---
SSH-51	DQ465804	499	No homology	---
SSH-52	DQ465805	404	No homology	---
SSH-53	DQ465806	472	No homology	---
SSH-54	DQ465807	478	No homology	---
SSH-55	DQ465808	561	No homology	---
SSH-56	DQ465809	578	No homology	---
SSH-57	DQ465810	646	No homology	---
SSH-58	DQ465811	377	No homology	---
SSH-59	DQ465812	547	No homology	---
SSH-60	DQ465813	293	No homology	---
SSH-61	DQ465814	352	No homology	---
SSH-62	DQ465815	368	No homology	---
SSH-63	DQ465816	630	No homology	---

^bNo significant sequence homology found in genome, EST, and protein database.

Northern blot analysis (Pb)

Planta 2007 (in press)



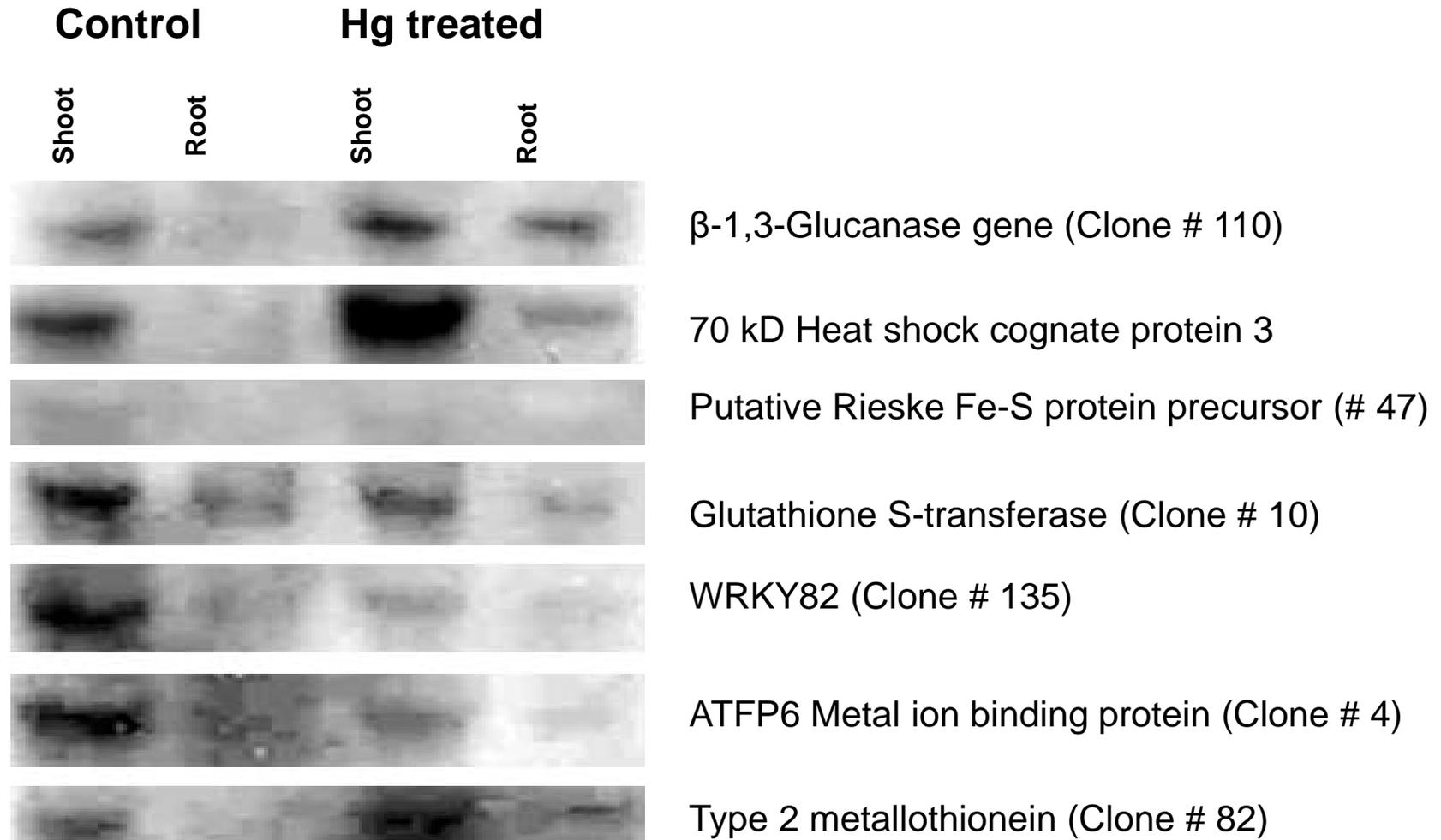
Sequencing results (Hg)

- 87 clones corresponds to unigenes
 - Clone # 31 had homology to **type 2 metallothionein (MT)**
 - Clone # 252- 70 kD Heat shock cognate protein 3
 - Clone # 4- ATFP6-metal ion binding protein
-

1. Vses-forward_016- gb|AAM94615.1|putative hydrolase [Glycine max] 333 7e-90
2. Vses-forward_031- ref|NP_190489.1|unknown protein [Arabidopsis thaliana] 58.5 7e-08
3. Vses-forward_032- ref|NP_195570.1|ATFP6 (FARNESYLATED PROTEIN 6); metal ion binding [Arabidopsis thaliana] 92.4 5e-18
4. Vses-forward_047- gb|AAG24883.1|ribulose-1,5-bisphosphate carboxylase small subunit rbcS2 [Glycine max] 206 6e-52
5. Vses-forward_048- emb|CAA68848.1|hin1 [Nicotiana tabacum] 186 1e-45
6. Vses-forward_063- dbj|BAA77604.1|plastidic aldolase NPALDP1 [Nicotiana paniculata] 394 1e-108
7. Vses-forward_064- gb|AAT80649.1|lipid transfer protein precursor [Malus x domestica] 139 3e-32
8. Vses-forward_079- emb|CAA85354.1|alpha-1,4 glucan phosphorylase, L isoform precursor [Vicia faba var. minor] 353 7e-96
9. Vses-forward_080- gb|AAG34805.1|glutathione S-transferase GST 15 [Glycine max] 337 2e-91
10. Vses-forward_095- emb|CAB17075.1|cysteine proteinase precursor [Phaseolus vulgaris] 297 3e-79
11. Vses-forward_030- sp|O65194|RBS_MEDSA Ribulose bisphosphate carboxylase small chain, chloroplast precursor (RuBisCO small subunit) 276 6e-73
12. Vses-forward_045- emb|CAA32197.1|chlorophyll a/b-binding protein [Lycopersicon esculentum] 376 1e-103
13. Vses-forward_046- gb|AAG24882.1|ribulose-1, 5-bisphosphate carboxylase small subunit rbcS1 [Glycine max] 283 6e-75
14. Vses-forward_061- gb|AAC16403.1|early light-induced protein [Glycine max] 192 3e-48
15. Vses-forward_062- ref|NP_190490.1|unknown protein [Arabidopsis thaliana] 42.4 0.006
16. Vses-forward_043- emb|CAA96570.1|CP12 [Pisum sativum] 189 5e-47
17. Vses-forward_060- emb|CAA05979.1|adenine nucleotide translocator [Lupinus albus] 117 1e-25
18. Vses-forward_091- ref|NP_974774.1|HAP2A; transcription factor [Arabidopsis thaliana] 152 1e-35
19. Vses-forward_025- sp|P93508|CRTC_RICCO Calreticulin precursor 358 9e-98
20. Vses-forward_058- sp|O22518|RSSA_SOYBN 40S ribosomal protein SA (p40) 170 1e-41
21. Vses-forward_089- emb|CAA45151.1|chloroplast Rieske FeS protein [Pisum sativum] 300 2e-80
22. Vses-forward_090- ref|NP_182172.1|unknown protein [Arabidopsis thaliana] 88.2 9e-17
23. Vses-forward_023- emb|CAA67696.1|23 kDa oxygen evolving protein of photosystem II [Solanum tuberosum] 260 1e-68
24. Vses-forward_055- ref|XP_467654.1|MutT/nudix-like [Oryza sativa (japonica cultivar-group)] 84.0 4e-15
25. Vses-forward_087- gb|AAD28640.2|geranylgeranyl hydrogenase [Glycine max] 372 1e-102
26. Vses-forward_037- ref|NP_186761.1|MTO1 (METHIONINE OVERACCUMULATION 1) [Arabidopsis thaliana] 155 6e-37
27. Vses-forward_085- ref|XP_469854.1|putative dehydrogenase precursor [Oryza sativa (japonica cultivar-group)] 293 3e-78
28. Vses-forward_003- emb|CAA33557.1|unnamed protein product [Pisum sativum] 124 8e-28
29. Vses-forward_020- emb|CAA43590.1|Type I (26 kD) CP29 polypeptide [Lycopersicon esculentum] 211 7e-54
30. Vses-forward_067- emb|CAB79860.1|putative zinc finger protein [Arabidopsis thaliana] 166 2e-40
31. Vses-forward_068- dbj|BAD18379.1|type 2 metallothionein [Vigna angularis] 110 2e-23
32. Vses-forward_083- ref|NP_177596.1|DNA binding [Arabidopsis thaliana] 88.6 7e-17
33. Vses-forward_084- emb|CAA32429.1|unnamed protein product [Arabidopsis thaliana] 43.5 0.002
34. Vses-forward_001- gb|AAM94806.1|rubisco activase alpha [Gossypium hirsutum] 105 5e-22
35. Vses-forward_002- emb|CAA81078.1|glycine hydroxymethyltransferase [Flaveria pringlei] 399 1e-109
36. Vses-forward_034- gb|AAQ72789.1|60S ribosomal protein L5 [Cucumis sativus] 358 1e-97
37. Vses-forward_050- dbj|BAB86847.1|elongation factor EF-2 [Pisum sativum] 183 2e-45
38. Vses-forward_066- gb|AAK25800.1|rubisco activase [Zantedeschia aethiopica] 43.9 0.002
39. Vses-forward_081- emb|CAA81082.1|glycine hydroxymethyltransferase [Solanum tuberosum] 408 1e-112
40. Vses-forward_031- dbj|D78130.1|Homo sapiens mRNA for squalene epoxidase, complete cds 89.7 6e-15
41. Vses-forward_032- ref|NM_120019.1|Arabidopsis thaliana ATFP6 (FARNESYLATED PROTEIN 6); metal ion binding AT4G38580 (ATFP6) mRNA, complete cds 79.8 6e-12
42. Vses-forward_047- gb|AF303941.1|AF303941 Glycine max ribulose-1,5-bisphosphate carboxylase small subunit rbcS3 mRNA, complete cds 454 1e-124
43. Vses-forward_048- gb|AY279310.1|Malus x domestica enolase-like mRNA, partial sequence 63.9 9e-07

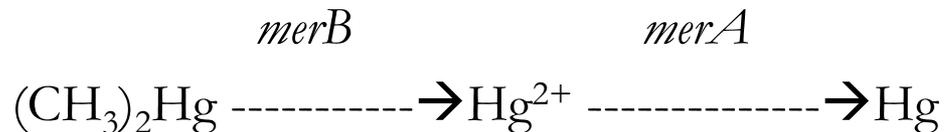
44. Vses-forward_063-gb|M97476.1|PEAALDIA Pisum sativum L. 3' end cds 694 0.0
45. Vses-forward_064-gb|DQ122797.1|Medicago sativa clone QB12 lipid transfer protein precursor, mRNA, partial cds 105 1e-19
46. Vses-forward_080-gb|AC161864.9|Medicago truncatula clone mth2-69d21, complete sequence 256 9e-65
47. Vses-forward_095-dbj|AP006110.1|Lotus corniculatus var. japonicus genomic DNA, chromosome 1, clone:LJT39G11, TM0195b, complete sequence 500 1e-138
48. Vses-forward_014-emb|V00458.1|GMRUBP Glycine max gene encoding ribulose-1,5-bisphosphate carboxylase small subunit 240 3e-60
49. Vses-forward_030-gb|AF056315.1|AF056315 Medicago sativa ribulose-1,5-bisphosphate carboxylase small subunit mRNA, nuclear gene encoding chloroplast protein, complete cds 252 1e-63
50. Vses-forward_045-emb|X81962.1|PSLHAB P.sativum mRNA for type II chlorophyll a/b binding protein 400 1e-108
51. Vses-forward_060-emb|AJ003197.1|LAAJ3197 Lupinus albus mRNA for adenine nucleotide translocator 260 2e-66
52. Vses-forward_009-gb|AC125095.3|Mus musculus BAC clone RP24-179H12 from chromosome 14, complete sequence 52.0 0.003
53. Vses-forward_025-gb|U74630.1|RCU74630 Ricinus communis calreticulin mRNA, complete cds 466 1e-128
54. Vses-forward_058-emb|AJ006759.1|CAR6759 Cicer arietinum mRNA for ribosome-associated protein p40 216 3e-53
55. Vses-forward_089-dbj|AB025003.1|Cicer arietinum mRNA for plastoquinol-plastocyanin reductase, partial cds 317 2e-83
56. Vses-forward_090-gb|AC109247.14|Mus musculus chromosome 9, clone RP23-462C14, complete sequence 56.0 9e-05
57. Vses-forward_008-gb|U74630.1|RCU74630 Ricinus communis calreticulin mRNA, complete cds 513 1e-142
58. Vses-forward_087-gb|DQ013361.1|Lotus corniculatus var. japonicus geranylgeranyl hydrogenase (GGH) mRNA, complete cds 652 0.0
59. Vses-forward_005-gb|AF220405.1|AF220405 Vitis riparia transcription factor (Rev136-2) mRNA, complete cds 75.8 2e-10
60. Vses-forward_037-gb|AF097180.1|AF097180 Nicotiana tabacum cystathionine gamma-synthase precursor (metB) mRNA, complete cds 69.9 8e-09
61. Vses-forward_085-gb|BT009463.1|Triticum aestivum clone wr1.pk0004.c11.fis, full insert mRNA sequence 198 1e-47
62. Vses-forward_003-dbj|AB236819.1|Trifolium pratense RNA for putative PSII-P protein, partial cds, clone: C214 244 2e-61
63. Vses-forward_020-gb|BT014450.1|Lycopersicon esculentum clone 133776F, mRNA sequence 230 2e-57
64. Vses-forward_067-dbj|AP004913.1|Lotus corniculatus var. japonicus genomic DNA, chromosome 5, clone:LJT02A14, TM0072a, complete sequence 206 4e-50
65. Vses-forward_083-ref|XM_466397.1|Oryza sativa (japonica cultivar-group), mRNA 75.8 7e-11
66. Vses-forward_084-gb|AC129090.21|Medicago truncatula clone mth2-14i8, complete sequence 113 2e-22
67. Vses-forward_002-gb|M87649.1|PEASHMTA Pisum sativum serine hydroxymethyltransferase mRNA, complete cds 763 0.0
68. Vses-forward_034-dbj|AB049724.1|Pisum sativum ssa-15 mRNA for putative senescence-associated protein, complete cds 460 1e-126
69. Vses_06_A06_T3 gb|AAQ87663.1|translationally controlled tumor protein [Elaeis guineensis] 224 1e-57
70. Vses_06_A08_T3 gb|AAX94836.1|Major Facilitator Superfamily, putative [Oryza sativa (japonica cultivar-group)] 115 4e-25
71. Vses_06_A12_T3 sp|P27774|KPPR_MESCR Phosphoribulokinase, chloroplast precursor (Phosphopentokinase) (PRKase) (PRK) 64.7 1e-09
72. Vses_06_B02_T3 ref|NP_192718.1|RPS18C (S18 RIBOSOMAL PROTEIN); RNA binding / structural constituent of ribosome [Arabidopsis thaliana] 93.6 2e-18
73. Vses_06_C01_T3 gb|AAB36543.1|DnaJ-like protein [Phaseolus vulgaris] 52.0 7e-06
74. Vses_06_C07_T3 ref|NP_187886.2|oxidoreductase [Arabidopsis thaliana] 170 1e-41
75. Vses_06_C08_T3 sp|P34921|G3PC_DIACA Glyceraldehyde-3-phosphate dehydrogenase, cytosolic 219 2e-56
76. Vses_06_C10_T3 gb|AAM93434.1|40S ribosomal S4 protein [Glycine max] 142 3e-33
77. Vses_06_C11_T3 gb|AAG33924.1|auxin-repressed protein [Robinia pseudoacacia] 169 5e-41
78. Vses_06_E08_T3 gb|AAS57914.1|70 kDa heat shock cognate protein 3 [Vigna radiata] 343 6e-93
79. Vses_06_D03-g_025-1672 gb|DQ322696.1|Glycine max WRKY82 mRNA, complete cds 119 8e-24
80. Vses_06_D08_T3 gb|AAA65011.1|similar to Atriplex nummularia chaperone ANJ1 protein, Swiss-Prot Accession Number JQ2142 76.6 3e-13
81. Vses_06_B09_T3 emb|AJ749797.1|Photobacterium damsela subsp. piscicida trpB gene for putative transposase, clone pRDA16 50.1 0.005
82. Vses_06_C08_T3 gb|DQ355800.1|Glycine max glyceraldehyde-3-phosphate dehydrogenase (GAPC1) mRNA, complete cds 323 3e-85
83. Vses_06_C11_T3 gb|AY009094.1|Robinia pseudoacacia auxin-repressed protein mRNA, complete cds 375 1e-100
84. Vses_06_C12_T3 emb|AJ006764.1|CAR6764 Cicer arietinum mRNA for putative cytidine or deoxycytidylate deaminase, partial 176 1e-41
85. Vses_06_D03_T3 gb|AY109342.1|Zea mays CL3469 4 mRNA sequence 167 4e-38
86. Vses_06_D04_T3 dbj|AB089677.1|Prunus persica PpNRT1 mRNA for nitrate transporter, complete cds 107 5e-20
87. Vses_06_F08-g_054-1672 dbj|AB242265.1|Sesbania rostrata Srglu64 mRNA for beta-1,3-glucanase, complete cds 795 0.0

Northern blot analysis (Hg)



Bioengineering of Plants for Efficient Remediation

- *Arabidopsis* transgenics constructed to express bacterial genes *merB* and *merA*. (PNAS 93, 1996)

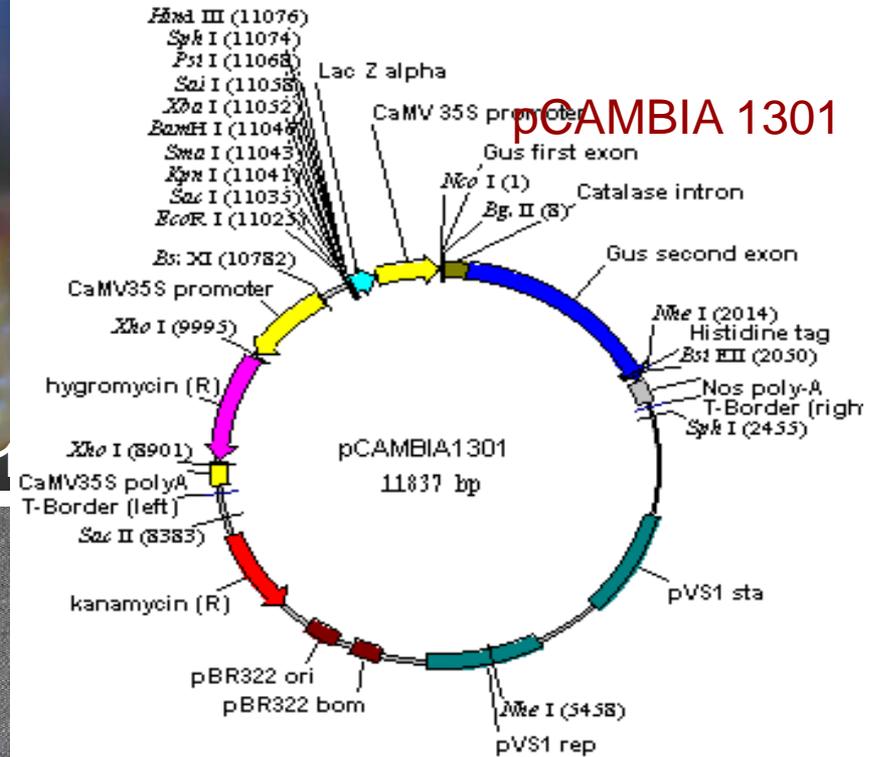
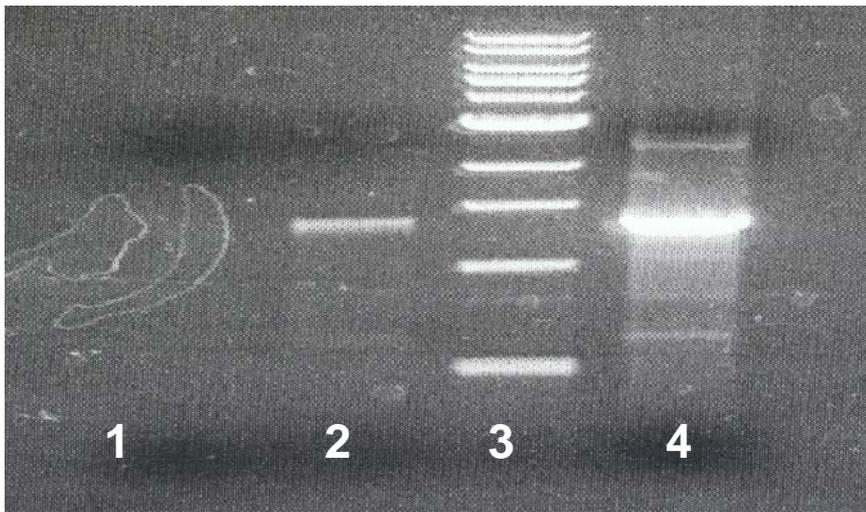
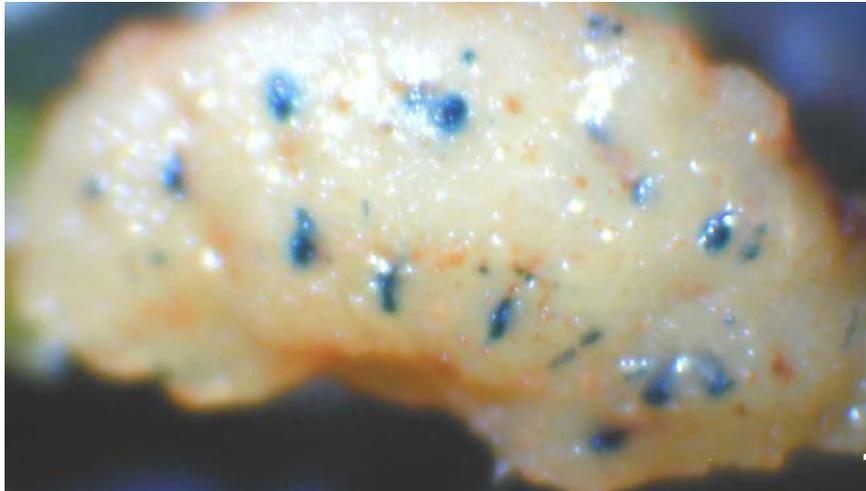


- **Indian mustard** transformed with *ATP Sulfurylase* (*APS*) genes demonstrate 4-fold increase in APS activity and accumulated 3X Se than the wild type.
- **Transgenic tomato** over expressing the bacterial gene *1-aminocyclopropane-1-carboxylic acid* (*ACC*) *deaminase* demonstrated enhanced tolerance for and accumulation of Co, Cu, Ni, Pb and Zn (J. Biotech 81, 2000).
- **Transgenic tobacco** expressing *citrate synthase* showed enhanced tolerance to Al toxicity (Science 276, 1997).
- **Transgenic *Arabidopsis*** expressing *phytochelatinsynthase* from wheat demonstrated enhanced accumulation of Cd (PNAS 100, 2003)

Sesbania Transformation

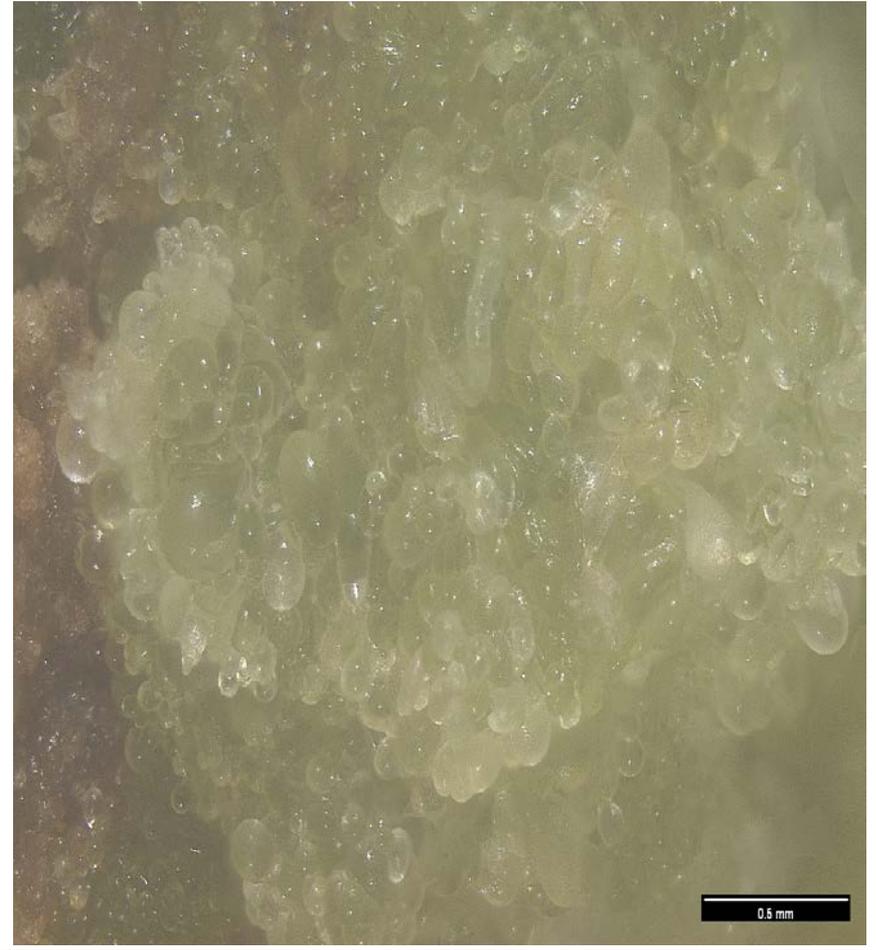
- Developed *in vitro* regeneration system using nodal explants
 - *Sesbania* callus infected with *Agrobacterium* containing pCambia 1305
 - pCambia has *GUS* gene which produces beta-glucuronidase
 - *GUS* histochemical assay to check gene expression.
 - Expression confirmed by PCR
-

Sesbania callus showing transformation



Lane 1: *S. drummondii* transformed tissue with strain K289 (without plasmid), **Lane 2:** transformed tissue with strain K289pCAMBIA 1305.1. **Lane 3:** 1 kb ladder; **Lane 4:** *GUS* gene amplified from pCAMBIA 1305.1 plasmid

Sesbania Regeneration



General Conclusion

- Phytoremediation by *Sesbania* is effective against a wide variety of contaminants.
 - Effective for sites with shallow contaminated soils.
 - A **type II metallothionein gene** identified - may be involved in heavy metal detoxification
 - Pb and Hg in *Sesbania* also induced other stress related genes
 - Slow process
 - Interdisciplinary approach
 - More research to manipulate metals accumulation efficiency of *Sesbania*.
-

ACKNOWLEDGEMENTS

Collaborators

Dr. J. Andersland, WKU
Dr. K. Raghothama, Purdue U
Dr. J. Jain, Univ. Notre Dam
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Financial Support

NSF-EPSCoR
Ogden College
