STC

Environmental Services Inc.
Geologists and Environmental Scientists

4754 RESEARCH DRIVE **SAN ANTONIO, TEXAS 78240** **Office (210) 696-6286 / FAX (210) 696-8761**

May 31, 2023

Ms. Corrina Green
Assoc. Vice President
Office of Real Estate, Construction, and Planning
University of Texas at San Antonio
One UTSA Circle
San Antonio, Texas 78249

Subject: Indoor Air Quality and Mold Assessment Report
The Institute of Texan Cultures
801 E César E. Chávez Blvd.
San Antonio, Texas 78205
STC Project No. 230171

Ms. Green,

This letter presents the results of an indoor air quality and mold assessment that was performed at the location noted above. The work involved a visual survey of the building and the testing for airborne fungi. Additional testing included the measurement of moisture content of building materials, carbon dioxide, carbon monoxide, temperature, and humidity. Additional details concerning the work are presented below.

PROJECT INFORMATION

The work was performed in the building known as the Institute of Texan Cultures located at 801 E César E. Chávez Blvd. in San Antonio, Texas. We understand that UTSA is evaluating options for remodeling or possibly relocating the Institute of Texan Cultures. As part of that evaluation, we understand that UTSA is seeking information regarding air quality in the building.

VISUAL SURVEY

On May 22, 2023, STC conducted a visual inspection of the building for any conditions which may impact indoor air quality. The visual inspection revealed the following conditions:

- No visible mold was identified during the assessment.
- Minor water-stained ceiling tiles were noted in Break Room 1.03.09 and in Corridor 1.03A.
- Minor dust accumulations were noted on the HVAC supply air registers located in Corridor 1.03A.

SAMPLING

Airborne Mold Sampling

A total of six (20) air samples were collected on May 22, 2023. Nineteen (19) of these air samples were collected from the normal breathing zone inside the building (See Photographs 1 through 19). These normal breathing zone samples included the following locations:

- First Floor – Corridor 1.01 by Restroom 1.01.04A
- First Floor – Room 1.02.05B
- First Floor – Corridor 1.03A
- First Floor – Corridor 1.05 by 1.05.19
- First Floor – Corridor 1.04 by 1.04.09
- First Floor – Room 1.01.03
- Third Floor – Room 3.01.02
- Third Floor – Room 3.12.04C
- Third Floor – Room 3.08.07
- Third Floor – Room 3.07.02
- Third Floor – Room 3.04.20
- Third Floor – Room 3.04.10
- Third Floor – Room 3.01.05
- Second Floor – Corridor 2.01 by 2.01.02A
- Second Floor – Corridor 2.02 (exhibit area)
- Second Floor – Exhibit area north of Room 2.08A
- Second Floor – Corridor 2.08 (Dome Theater)
- Second Floor – Corridor 2.04 (exhibit area)
- Second Floor – Corridor 2.05 (exhibit area)

The normal breathing zone is defined as a height of three to five feet from the floor of the building. This is the height at which occupants normally sit or stand. Breathing zone samples are designed to evaluate the typical airborne mold spore concentrations inhaled by building occupants. The twentieth and final air sample was taken outside of the building for purposes of comparison (See Photograph 20 in Appendix A).

The samples were collected by Mr. Michael Treviño of STC Environmental Services, Inc. Mr. Treviño is a licensed Mold Assessment Consultant in the State of Texas (License No. 1210, expiration January 18, 2024). The samples referenced above were sent to EM Lab of Ft. Lauderdale, Florida (Lab License No. 1014) for analysis. Each of the samples were analyzed using a microscopy method that evaluates the genus or type of mold spores present.

A summary of the test results is presented in the attached Table I. Floor plans showing the sample locations are presented in the attached Figures 1 through 3. Site photographs showing the areas sampled are presented in Appendix A. The analytical laboratory report and chain of custody record are presented in Appendix B.

Moisture Readings

During the assessment, a meter was also used to measure moisture levels in sheetrock throughout the building. Sheetrock testing produced moisture levels ranging from 0.1 to 0.4 percent. This range is considered to be normal and dry.

Infrared Imaging

Infrared imaging of the project area was conducted using a FLIR MR176. This camera has the ability to detect subtle temperature differences in building materials which could reveal potential wet areas or areas where condensate may accumulate. Infrared imaging of the project area did not detect any temperature differentials that would suggest the presence of wet building materials.

Other Handheld Meter Readings

Handheld meters were used to measure temperature, humidity, carbon dioxide, and carbon monoxide throughout the building. One (1) measurement was also recorded outside the building for comparative purposes. For each sample location, the handheld meters collected data for approximately five (5) minutes. The data are presented as the minimum and maximum values of each parameter detected within the five (5) minute sample time. The results of this testing are shown in the attached Table II.

GENERAL DISCUSSION OF TESTING

A general discussion of testing and methods used to interpret the results are presented below.

Airborne Mold

The results of airborne mold testing should always include a comparison to outside levels at the time of indoor sampling. This comparison is desirable because outdoor mold levels vary widely, and outdoor air is mixed with indoor air through various mechanisms.

Outdoor air is deliberately brought into many commercial buildings through adjustable valve devices on the air handling machinery. This action is taken to reduce buildup of carbon dioxide and other indoor pollutants. Outdoor air will also enter a building through doors and windows.

It is generally accepted that "normal" outside levels of mold vary widely depending on laboratory protocol, weather, and seasonal conditions. Many of the spores are made up of the common genus of Aspergillus/Penicillium and Cladosporium. Total fungi concentrations of 500 to 5,000 counts per cubic meter are common during many parts of the year in much of Texas but can be significantly higher during periods when mold growth is abundant. Additional details regarding average mold levels in Texas is presented in the Extended Outdoor Comparison Report presented in Appendix B.

It should be noted that there are **NO** specific levels of mold/fungi that are considered harmful to humans. Individuals have a different tolerance level. In addition, different geographical locations exhibit a wide variance of air quality levels. Therefore, what is typical outside in one geographical area may never occur in another.

When evaluating indoor air quality, the types of mold present are also examined. Certain types of mold, such as Stachybotrys, tend to thrive indoors and are rarely found outdoors. When the concentration of mold types such as Stachybotrys is elevated, it often indicates damp conditions or water leaks creating indoor mold growth.

Evaluation of Airborne Mold through use of the Mold Score

A Mold Score can also be used to evaluate airborne mold. The Mold Score is a ranking system developed by EM Lab that evaluates the potential for indoor mold growth.

Mold scores range from 100 to 300. A rating less than 150 is considered low and indicative of a low probability of spores originating inside. A score from 150 to 250 is considered moderate, and a score greater than 250 suggests a high probability of indoor mold growth.

Temperature and Humidity

The Occupational Safety and Health Administration has published guidelines which indicate that occupants will be most comfortable when air handling systems maintain temperature in the range of 68 to 76 degrees Fahrenheit and humidity is kept to a range of 20 to 60%.

Mold growth can be promoted with higher temperatures and humidity. For some species of mold, growth is significantly enhanced when humidity exceeds 55%.

Carbon Dioxide and Carbon Monoxide

According to OSHA, carbon dioxide levels are a useful screening technique to ascertain whether adequate quantities of outside fresh air are being introduced into an indoor environment. Without the introduction of fresh air, carbon dioxide will naturally accumulate due to human respiration.

At 600 ppm, OSHA indicates there are minimal air quality complaints. OSHA indicates carbon dioxide (CO_2) levels above 1,000 ppm may indicate inadequate ventilation. When CO_2 is elevated above 1,000 ppm, occupants may complain of symptoms such as headaches, fatigue and eye and throat irritation. However, OSHA states that exceeding the 1,000 ppm CO_2 guideline does not necessarily indicate that the building is hazardous. Rather this level should be used as a guideline.

OSHA also recognizes a permissible exposure limit of 5,000 ppm for CO_2. This is the maximum allowable concentration, for a work environment, without respiratory protection.

Carbon monoxide (CO) is typically indicative of a combustion source. Average levels in homes without gas stoves vary from 0.5 to 5 ppm. Levels near properly adjusted gas stoves are often 5 to 15 ppm and those near poorly adjusted stoves may be 30 ppm or higher.

OSHA also recognizes a permissible exposure limit of 50 ppm for CO. This is the maximum allowable concentration, for a work environment, without respiratory protection.

Moisture Testing

Moisture readings were collected with a Delmhorst BD-2100 moisture meter. This meter evaluates moisture content of sheetrock and wood by measuring the electrical resistance between two electrodes, or metal pins, that are inserted into the building material. When moisture is present, electricity flows more easily. Conversely, dryer materials will resist electrical flow.

For sheetrock, the meter readings may range between 0.2% and 50% moisture content. The meter manufacturer manual indicates that moisture content readings should be interpreted as follows:

- 0 to 0.5% Moisture – Normal
- 0.5 to 1% Moisture – Borderline
- >1% Moisture - too wet for painting or wallpaper

CONCLUSIONS

The results of the assessment are summarized in the following outline.

1. The indoor air sample collected in Corridor 1.03A (Sample BZ-03) produced a Mold Score value of 300 (See Table I). A Mold Score value of 300 indicates the highest possible obtainable score and suggests a *high* probability that the spores found were derived from an indoor source of mold growth. When individual mold types are examined, *Penicillium / Aspergillus sp.* was detected inside at a concentration of 15,000 spores/m^3 (See Table I). This level exceeded the 90th percentile of 1,100 spores/m^3 found outdoors throughout the entire year in the Southern United States (See Table I). These findings indicate an indoor source of *Penicillium / Aspergillus sp.* is likely present in Corridor 1.03A.

2. The underlying source of the elevated levels of *Penicillium / Aspergillus sp.* detected in Corridor 1.03A is unknown. The visual inspection noted minor water-stained ceiling tiles and dust accumulations on HVAC supply air registers in this area. Testing of building materials did not indicate the presence of elevated moisture. Therefore, based on the assessment, we suspect the high level of airborne mold found in Corridor 1.03A is caused by one of or combination of two sources, which are:

    a. Mold growth inside the HVAC system and/or;
    b. Hidden mold growth from past water leaks

3. The remaining eighteen (18) indoor air samples collected from the normal breathing zones located throughout the building produced Mold Score values ranging between 101 to 122 (See Table I). Mold Score values ranging from 100 to 150 suggest a low probability that the spores found were derived from an indoor source of mold growth. Further, none of the individual mold types exceeded the 50th percentile for outdoor air. Therefore, this finding indicates that the mold spores found in these samples were likely derived from an outdoor source and indoor mold growth is not likely occurring in these areas.

4. Relative humidity levels in approximately one half of the areas exceeded 55 percent (See Table II). These areas included the following:

    a. First Floor – Corridor 1.01 by Restroom 1.01.04A
    b. Third Floor – Room 3.01.02
    c. Third Floor – Room 3.12.04C
    d. Third Floor – Room 3.04.10
    e. Third Floor – Room 3.01.05
    f. Second Floor – Corridor 2.01 by 2.01.02A
    g. Second Floor – Corridor 2.02 (exhibit area)
    h. Second Floor – Exhibit area north of Room 2.08A
    i. Second Floor – Corridor 2.08 (Dome Theater)
    j. Second Floor – Corridor 2.04 (exhibit area)

Mold may spontaneously grow on surfaces when humidity levels are sustained for long periods above 55 percent. This finding suggests that the HVAC systems serving the above-listed areas may not be controlling humidity levels as efficiently as the HVAC systems serving other areas of the building.

5. The outdoor level of humidity on the day of testing was approximately 45% (See Table II). HVAC systems typically reduce humidity to levels that are 10% to 20% below outside levels. However, with an outside level of only 45%, a reduction of 10% may not be feasible. Rather, a 10% to 20% reduction may only be feasible when outside humidity levels are higher. In addition, if outdoor humidity levels were significantly higher 24 to 48 hours prior to testing, that condition could account for indoor humidity levels found on the day of testing, and indicate the HVAC system has not yet compensated for the change in equilibrium.

6. Testing of the sheetrock throughout the building produced moisture concentrations ranging from 0.1 to 0.4 percent. This range is normal. Further, infrared imaging of the building did not detect any temperature differentials that would suggest the presence of wet building materials.

7. Carbon monoxide was not detected inside the building. Therefore, the carbon monoxide levels inside were within the mandatory OSHA limits and suggested OSHA guidelines for best comfort (See Table II).

8. Carbon dioxide levels inside were within the mandatory OSHA limits and the suggested OSHA guidelines for best comfort (See Table II).

9. Temperature levels were generally within the range for best comfort with two exceptions. Temperatures were colder than the targeted comfort level in Rooms 3.01.02, 3.08.07, and 3.07.02 (See Table II). We understand that temperature levels are deliberately colder in those rooms due to the optimal temperature for document storage. Temperature was higher than the target comfort zone in the corridor by Room 1.04.09. The higher temperature in that area is apparently due to poor HVAC supply air distribution in that area.

RECOMMENDATIONS

Based on the air testing, STC has the following recommendations to improve the indoor air quality:

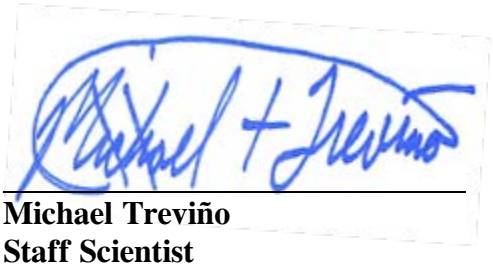
1. We recommend that the quality of the HVAC supply air be tested in Corridor 1.03A. That can be accomplished by testing of the air in the HVAC supply registers. If that testing indicates the HVAC system is not the likely source of the elevated spore counts, then additional actions will be recommended to locate the source.

2. We recommend that the HVAC systems be evaluated for proper humidity control. That evaluation should include additional indoor measurements when outdoor humidity levels are higher. If additional measurements confirm the HVAC system is not properly controlling humidity, then modifications to the system may be warranted. If plans for the building involve air handler replacement or significant remodeling that will involve rearrangement of the HVAC duct system, then an evaluation of humidity control can be deferred until those actions are completed.

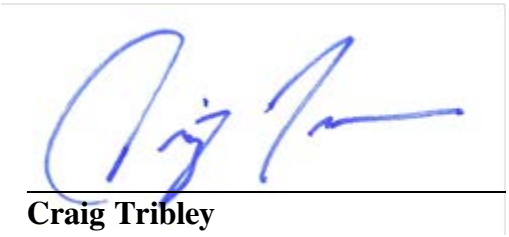
3. It is recommended that the HVAC system be evaluated for proper balancing in the corridor near Room 1.04.09. Adjustments to the supply air flow rates could potentially lower the temperatures in that corridor. Again, if plans for the building involve air handler replacement or significant remodeling that will involve rearrangement of the HVAC duct system, then an evaluation of the system balance can be deferred until those actions are completed.

If you have any questions concerning the assessment, please do not hesitate to call our office at (210) 696-6288.

Respectfully,



Michael Treviño
Staff Scientist
Mold Consultant 1210



Craig Tribley
President
Mold Consultant 1104

Attachments:
Table I – Summary of Airborne Mold Testing
Table II – Handheld Meter Readings
Figure 1 – First Floor - Sample Locations
Figure 2 – Second Floor - Sample Locations
Figure 3 – Third Floor - Sample Locations
Appendix A – Site Photographs
Appendix B – Analytical Laboratory Report and Chain of Custody

Table I
Summary of Airborne Mold Tests

Sample Name/Reference Standard	Sample Type	Total Spores Spores/m³	*Penicillium/ Aspergillus sp.* Spores/m³	*Cladosporium sp.* Spores/m³	Mold Score
OUT-20: Outside the building	Outside	5,300	190	610	N/A
BZ-01: Corridor 1.01 by 1.01.04A	Breathing Zone	360	80	120	111
BZ-02: Room 1.02.05B	Breathing Zone	110	27	27	104
BZ-03: Corridor 1.03A	Breathing Zone	15,000	15,000	67	300
BZ-04: Corridor by Rm. 1.05.19	Breathing Zone	67	67	0	110
BZ-05: Corridor 1.04 by Rm. 1.04.09	Breathing Zone	130	93	27	114
BZ-06: Room 1.01.03	Breathing Zone	240	150	0	122
BZ-07: Room 3.01.02	Breathing Zone	53	27	13	105
BZ-08: Room 3.12.04C	Breathing Zone	110	67	27	110
BZ-09: Room 3.08.07	Breathing Zone	40	0	0	105
BZ-10: Room 3.07.02	Breathing Zone	13	0	0	101
BZ-11: Room 3.04.20	Breathing Zone	150	53	27	108
BZ-12: Room 3.04.10	Breathing Zone	250	0	190	122
BZ-13: Room 3.01.05	Breathing Zone	40	13	0	102
BZ-14: Corridor 2.01 by Room 2.01.02A	Breathing Zone	40	0	0	105
BZ-15: Corridor 2.02 (Exhibit)	Breathing Zone	40	0	13	105
BZ-16: North of Room 2.08A (Exhibit)	Breathing Zone	40	27	0	104
BZ-17: Room 2.08 (Dome Theater)	Breathing Zone	27	13	0	102
BZ-18: Corridor 2.04 (Exhibit)	Breathing Zone	27	13	0	102
BZ-19: Corrdior 2.05 (Exhibit)	Breathing Zone	27	13	0	102
Normal Outside Level - 50th percentile - Entire Year in Southern U. S.	N/A	N/A	210	1,100	N/A
Very High Outside Level - 90th percentile - Entire year in Southern U. S.	N/A	N/A	1,100	6,800	N/A
Frequency of Detection in Outdoor Air	N/A	N/A	67%	97%	N/A

LEGEND

	Green shaded cells show indoor values that exceeded the normal outdoor value (50th percentile)
	Red shaded cells show Mold Scores greater than 150

Mold Score Ratings

100 to 150 - Low probability of indoor mold growth
150 to 250 - Moderate probability of indoor mold growth
250 to 300 - High probability of indoor mold growth

NOTES: Only selected mold types are shown. For a complete list of all mold types found, see lab report.

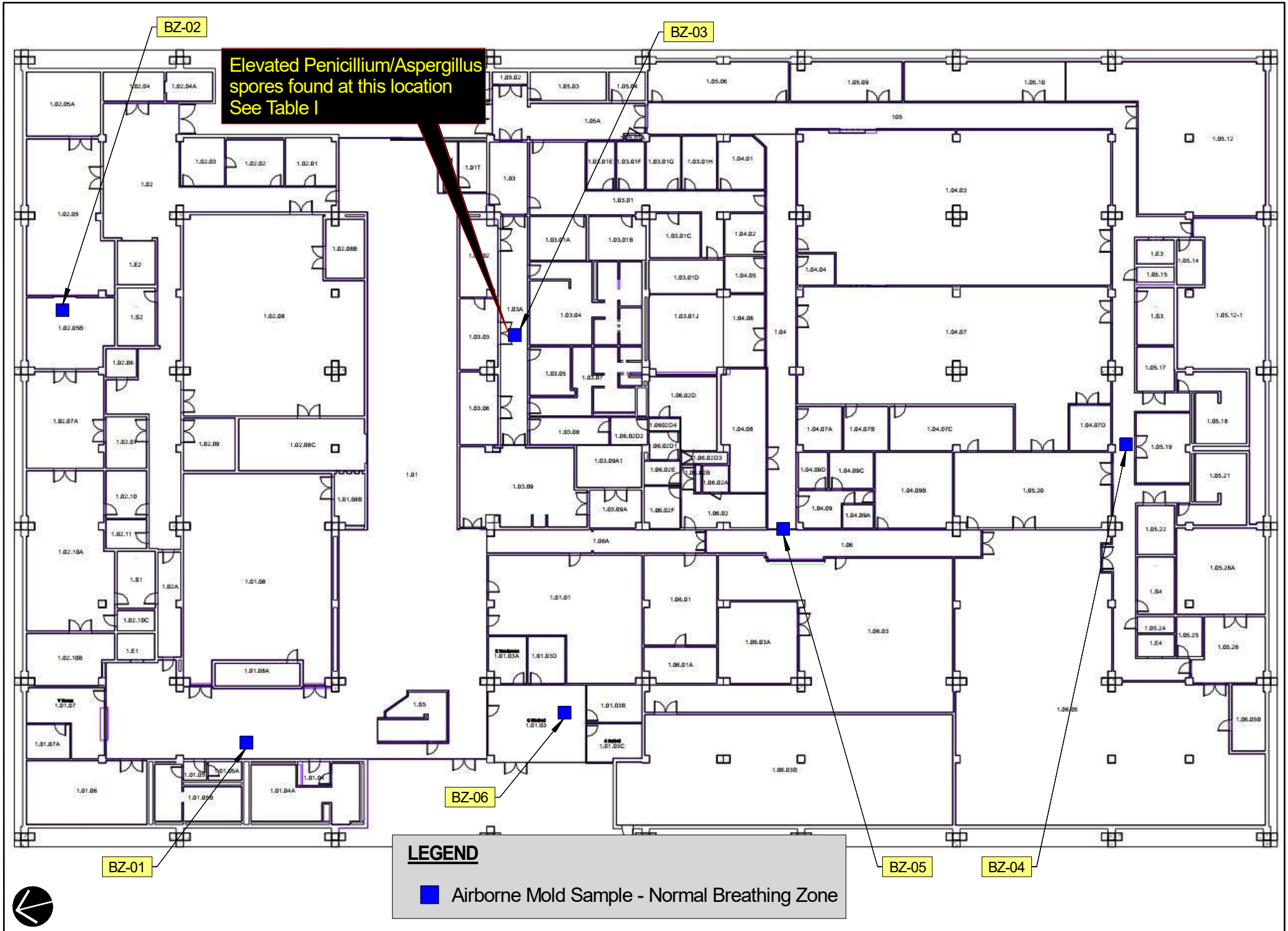
Table II
Handheld Meter Readings

Location	Temperature (°F)	Relative Humidity (%)	CO_2 (ppm)	CO_2 (ppm)
Outside the building	89.2 - 89.8	44.3 - 45.1	400 - 408	0
Corridor 1.01 by 1.01.04A	70.1 - 70.5	56.7 - 58.7	411 - 417	0
Room 1.02.05B	70.4 - 70.8	43.3 - 44.4	433 - 439	0
Corridor 1.03A	72.0 - 72.2	44.9 - 50.5	417 - 444	0
Corridor by Rm. 1.05.19	74.5 - 76.0	47.0 - 49.5	407 - 459	0
Corridor 1.04 by Rm. 1.04.09	78.4 - 80.9	41.8 - 46.1	405 - 418	0
Room 1.01.03	72.1 - 75.8	43.8 - 49.4	420 - 448	0
Room 3.01.02	63.7 - 67.0	54.4 - 62.5	401 - 425	0
Room 3.12.04C	68.0 - 69.0	54.3 - 56.5	416 - 421	0
Room 3.08.07	61.7 - 63.6	48.1 - 52.5	392 - 400	0
Room 3.07.02	60.5 - 62.4	45.5 - 48.9	398 - 407	0
Room 3.04.20	69.0 - 72.3	45.8 - 51.4	413 - 431	0
Room 3.04.10	68.8 - 69.5	60.1 - 62.5	410 - 415	0
Room 3.01.05	67.8 - 68.4	60.7 - 62.1	400 - 427	0
Corridor 2.01 by Room 2.01.02A	68.6 - 69.9	60.0 - 62.6	394 - 508	0
Corridor 2.02 (Exhibit)	67.3 - 69.3	61.1 - 65.0	395 - 459	0
North of Room 2.08A (Exhibit)	68.7 - 69.3	62.2 - 69.3	392 - 425	0
Room 2.08 (Dome Theater)	70.3 - 70.8	59.4 - 60.6	390 - 431	0
Corridor 2.04 (Exhibit)	71.6 - 72.2	55.4 - 56.8	397 - 416	0
Corrdior 2.05 (Exhibit)	73.8 - 74.0	48.9 - 49.2	451 - 558	0
Mandatory OSHA Limits	N/A	N/A	5,000 ppm	50 ppm
Best Indoor Comfort Range	68 to 76°F	20 to 60%	<1,000 ppm	<0 ppm

NOTES:

1. Green shaded cells indicate humidity levels above 55%. Mold may spontaneously grow on surfaces when this threshold is exceeded for extended periods.

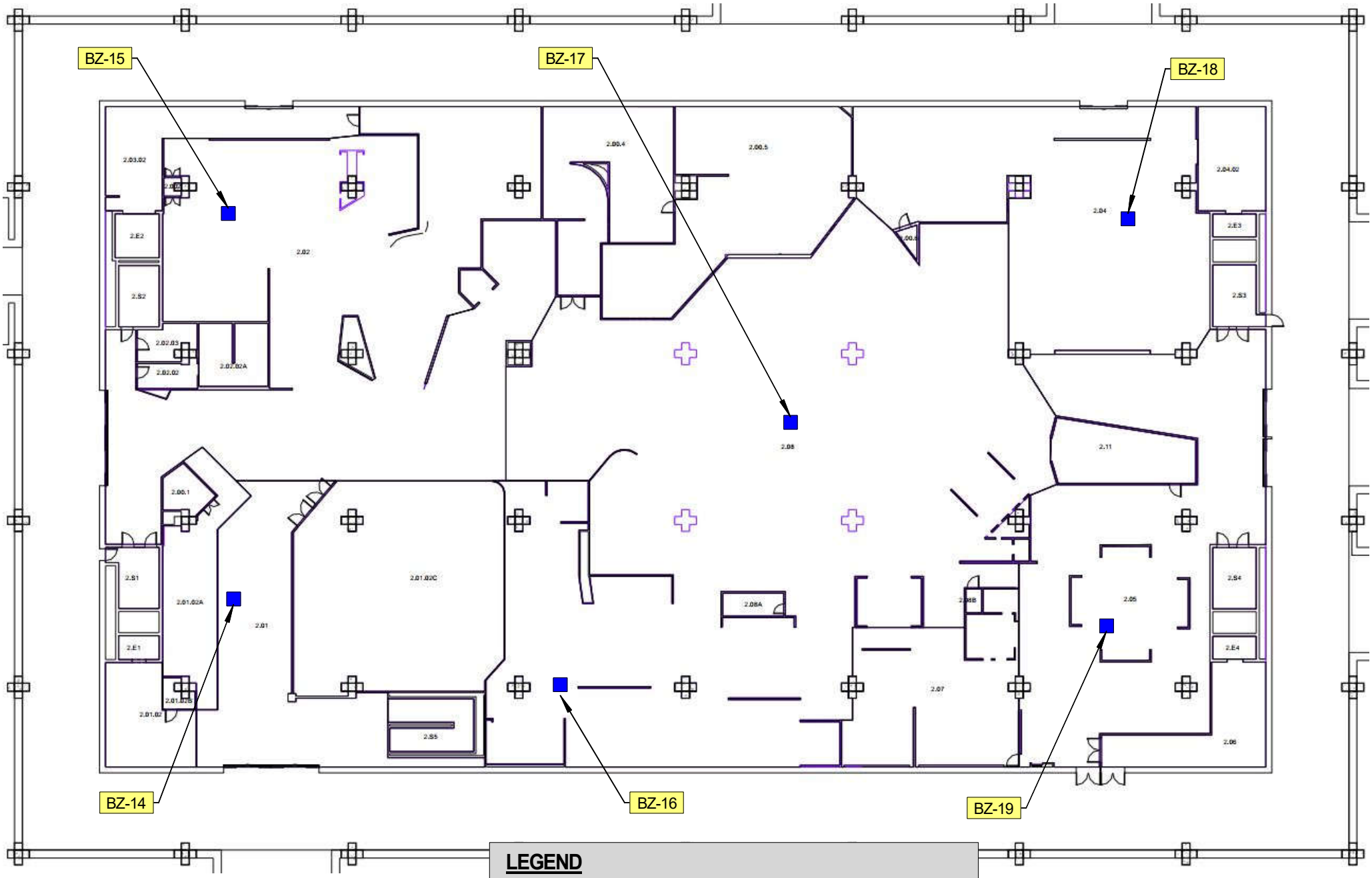
2. Red shaded cells show valaues outside the best indoor comfort range



STC Environmental Services Inc.. Geologists and Environmental Scientists

UTSA - ITC Bldg. - 1st Floor - Airborne Mold Sample Locations - 5/22/23

FIG. 1



STC Environmental Services Inc.. Geologists and Environmental Scientists	UTSA - ITC Bldg. - 2nd Floor - Airborne Mold Sample Locations - 5/22/23	FIG. 2

STC Environmental Services Inc.. Geologists and Environmental Scientists	UTSA - ITC Bldg. - 3rd Floor - Airborne Mold Sample Locations - 5/22/23	FIG. 3

APPENDIX A
SITE PHOTOGRAPHS

PHOTO 1: **Shows airborne mold sample BZ-01, indicated by the red arrow, collected from the normal breathing zone in Corridor 1.01 by Restroom 1.01.04A.**

PHOTO 2: **Shows airborne mold sample BZ-02, indicated by the red arrow, collected from the normal breathing zone in Room 1.02.05B.**

PHOTO 3: **Shows airborne mold sample BZ-03, indicated by the red arrow, collected from the normal breathing zone in Corridor 1.03A by Room 1.03.03.**

PHOTO 4: **Shows airborne mold sample BZ-04, indicated by the red arrow, collected from the normal breathing zone in Corridor 1.05 by Room 1.05.19.**

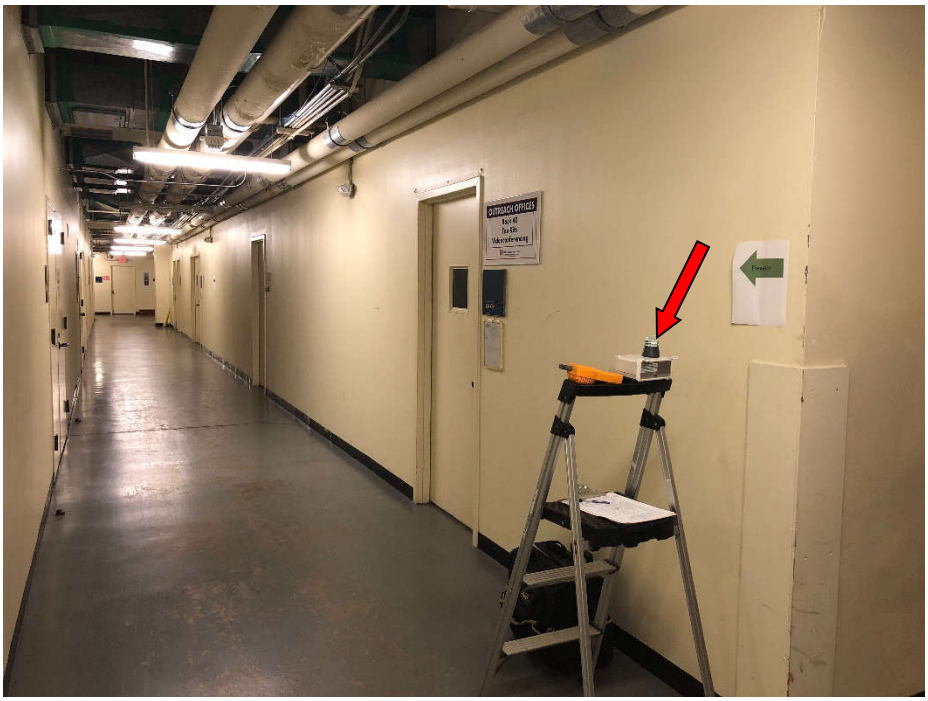


PHOTO 5: **Shows airborne mold sample BZ-05, indicated by the red arrow, collected from the normal breathing zone in Corridor 1.04 by Restroom 1.04.09.**

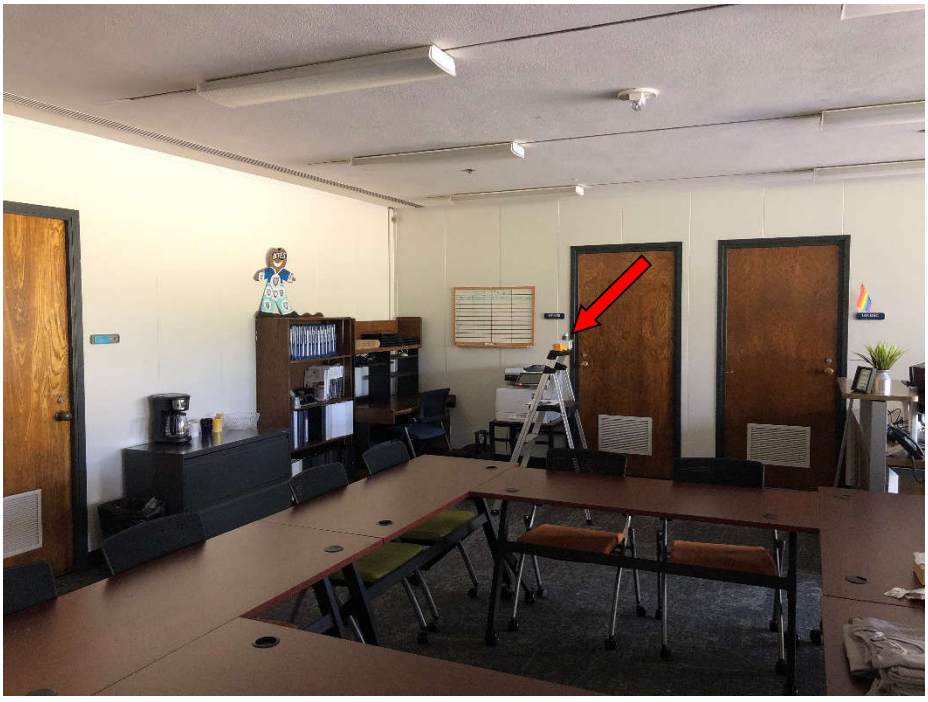


PHOTO 6: **Shows airborne mold sample BZ-06, indicated by the red arrow, collected from the normal breathing zone in Room 1.01.03.**

PHOTO 7: **Shows airborne mold sample BZ-07, indicated by the red arrow, collected from the normal breathing zone in Room 3.01.02.**

PHOTO 8: **Shows airborne mold sample BZ-08, indicated by the red arrow, collected from the normal breathing zone in Room 3.12.04C.**

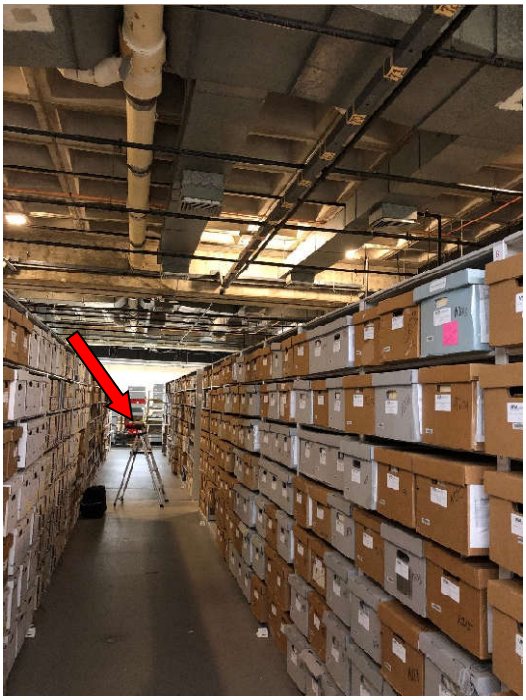


PHOTO 9: **Shows airborne mold sample BZ-09, indicated by the red arrow, collected from the normal breathing zone in Room 3.08.07.**

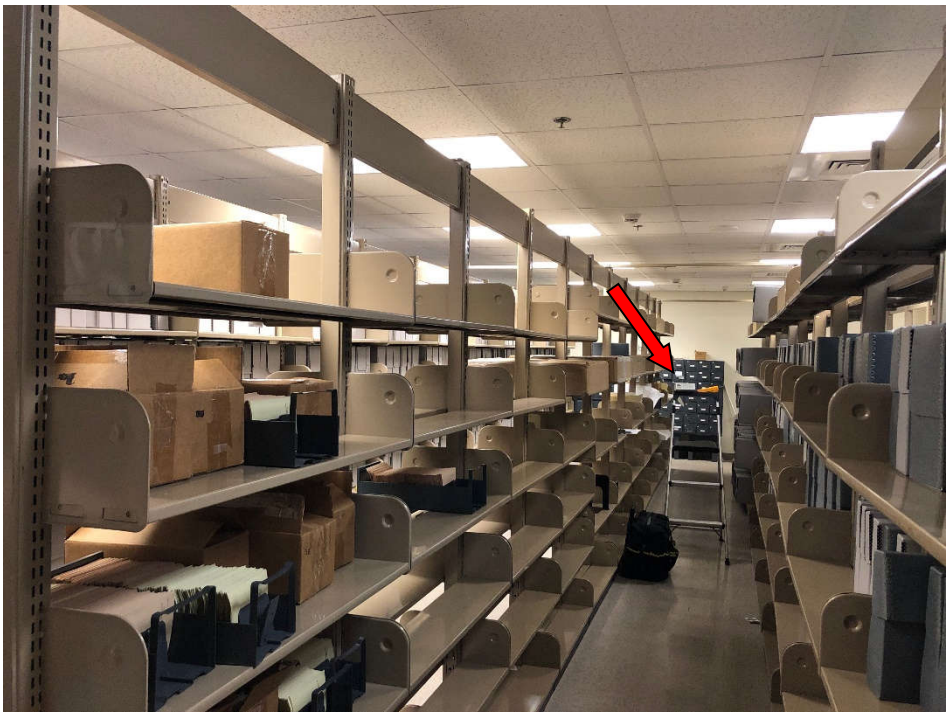


PHOTO 10: **Shows airborne mold sample BZ-10, indicated by the red arrow, collected from the normal breathing zone in Room 3.07.02.**

PHOTO 11: **Shows airborne mold sample BZ-11, indicated by the red arrow, collected from the normal breathing zone in Room 3.04.20.**

PHOTO 12: **Shows airborne mold sample BZ-12, indicated by the red arrow, collected from the normal breathing zone in Room 3.04.10.**

PHOTO 13: **Shows airborne mold sample BZ-13, indicated by the red arrow, collected from the normal breathing zone in Room 3.01.05.**

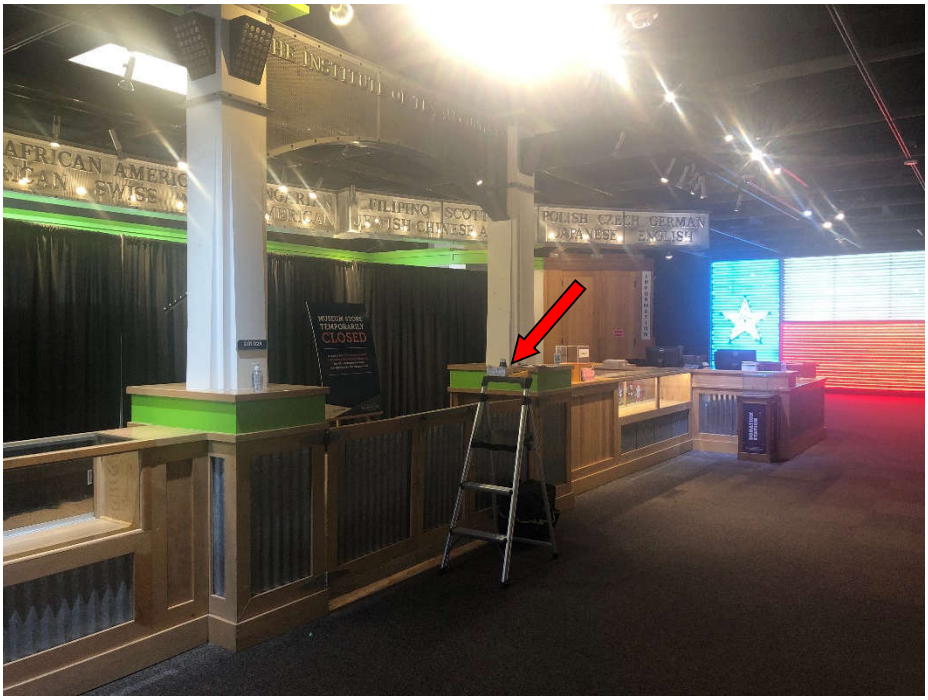


PHOTO 14: **Shows airborne mold sample BZ-14, indicated by the red arrow, collected from the normal breathing zone in Corridor 2.01 by Room 2.01.02A.**

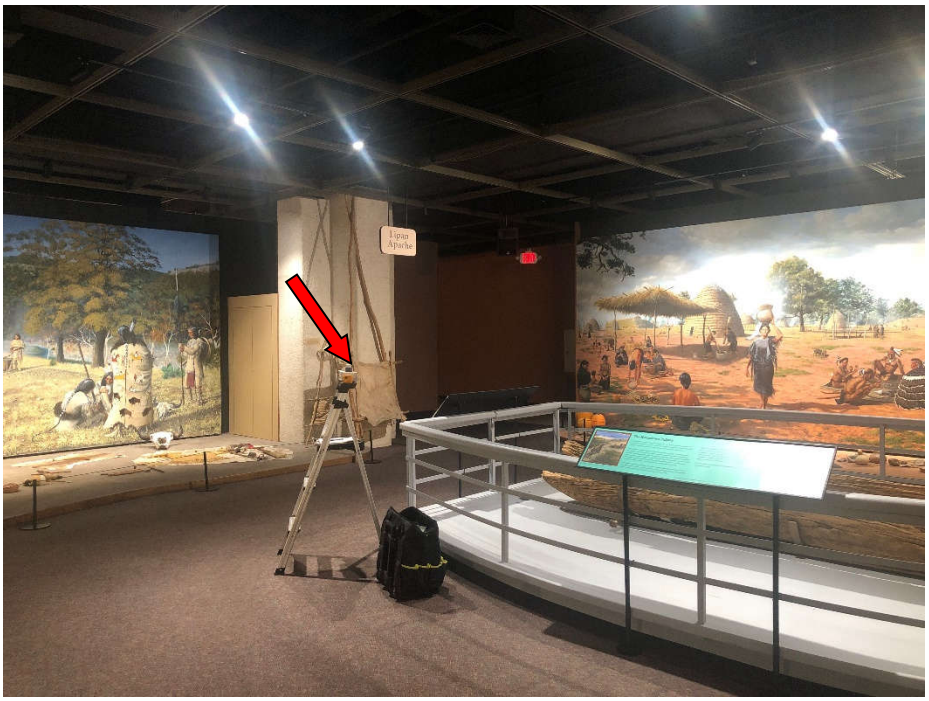


PHOTO 15: **Shows airborne mold sample BZ-15, indicated by the red arrow, collected from the normal breathing zone in Corridor 2.02.**

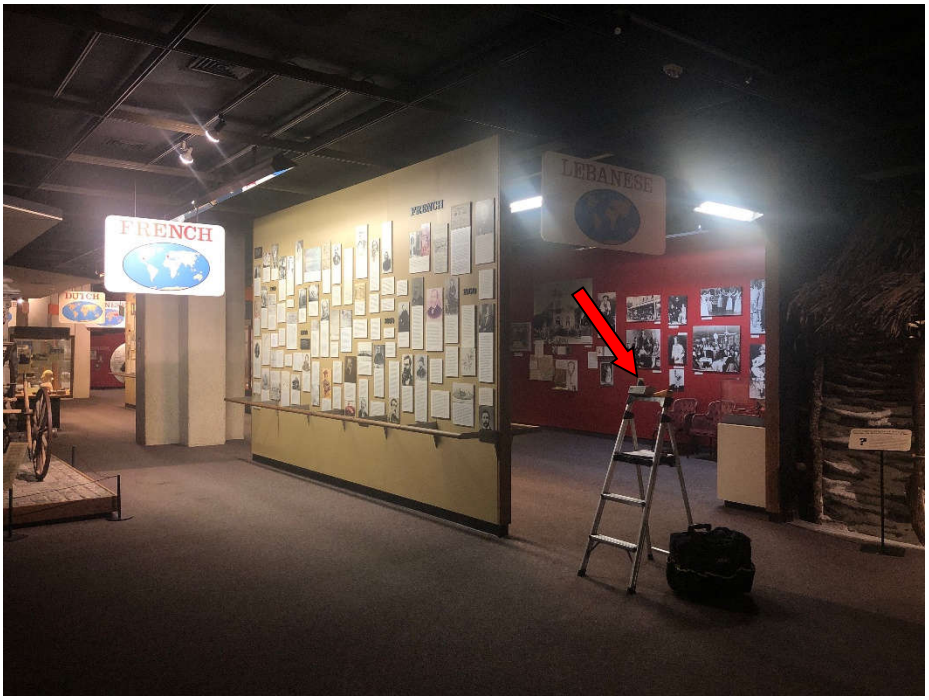


PHOTO 16: **Shows airborne mold sample BZ-16, indicated by the red arrow, collected from the normal breathing zone in the exhibit area north of Room 2.08A.**

PHOTO 17: **Shows airborne mold sample BZ-17, indicated by the red arrow, collected from the normal breathing zone in Room 2.08 (Dome Theater).**

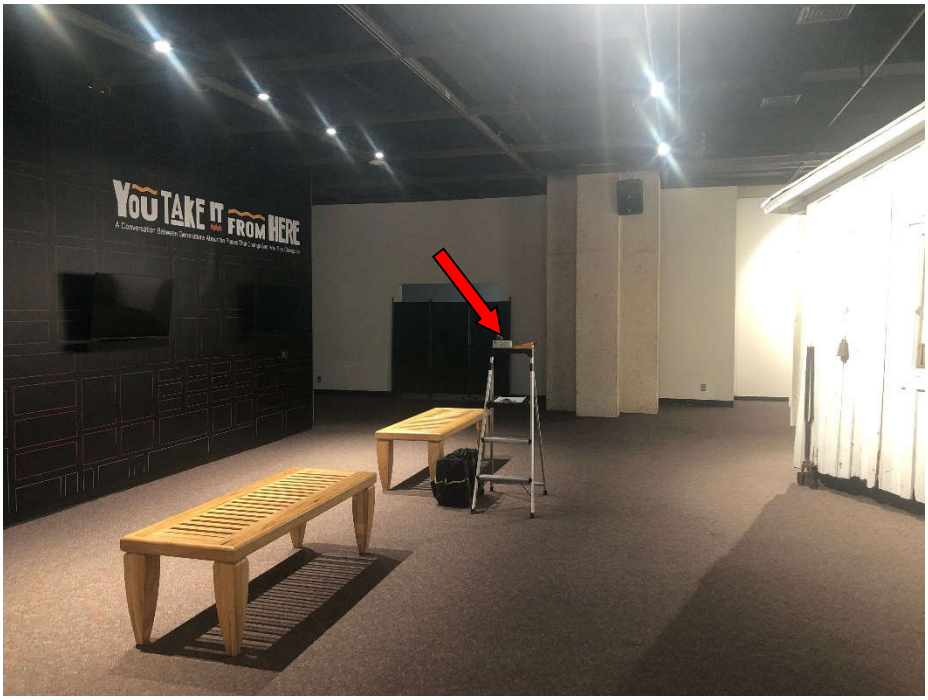


PHOTO 18: **Shows airborne mold sample BZ-18, indicated by the red arrow, collected from the normal breathing zone in Corridor 2.04.**

PHOTO 19: **Shows airborne mold sample BZ-19, indicated by the red arrow, collected from the normal breathing zone in Corridor 2.05.**

PHOTO 20: **Shows airborne mold sample BZ-20, indicated by the red arrow, collected from outside the UTSA Institute of Texan Cultures located at 801 E. Cesar E. Chavez Blvd. in San Antonio, Texas on May 22, 2023.**

APPENDIX B
ANALYTICAL LABORATORY REPORT AND CHAIN OF CUSTODY

eurofins | Built Environment Testing

Report for:

Michael T. Trevino
STC Environmental Services Inc.
4754 Research Dr
San Antonio, TX 78240

Regarding: Eurofins EPK Built Environment Testing, LLC
Project: 230171; ITC Bldg.
EML ID: 3269888

Approved by:

Business Unit Manager
Balu Krishnan

Dates of Analysis:
Spore trap analysis: 05-24-2023 and 05-25-2023

Service SOPs: Spore trap analysis (EM-MY-S-1038)
AIHA-LAP, LLC accredited service, Lab ID #173067

All samples were received in acceptable condition unless noted in the Report Comments portion in the body of the report. Due to the nature of the analyses performed, field blank correction of results is not applied. The results relate only to the samples as received and tested. Information supplied by the client which can affect the validity of results: sample air volume.

Eurofins EPK Built Environment Testing, LLC ("the Company"), a member of the Eurofins Built Environment Testing group of companies, shall have no liability to the client or the client's customer with respect to decisions or recommendations made, actions taken or courses of conduct implemented by either the client or the client's customer as a result of or based upon the Test Results. In no event shall the Company be liable to the client with respect to the Test Results except for the Company's own willful misconduct or gross negligence nor shall the Company be liable for incidental or consequential damages or lost profits or revenues to the fullest extent such liability may be disclaimed by law, even if the Company has been advised of the possibility of such damages, lost profits or lost revenues. In no event shall the Company's liability with respect to the Test Results exceed the amount paid to the Company by the client therefor.
Eurofins EPK Built Environment Testing, LLC's LabServe® reporting system includes automated fail-safes to ensure that all AIHA-LAP, LLC quality requirements are met and notifications are added to reports when any quality steps remain pending.

Eurofins EPK Built Environment Testing, LLC
6301 NW 5th way, Suite#: 1410, Ft. Lauderdale, FL 33309
(866) 871-1984 www.eurofinsus.com/Built

Client: STC Environmental Services Inc.
C/O: Michael T. Trevino
Re: 230171; ITC Bldg.

Date of Sampling: 05-22-2023
Date of Receipt: 05-24-2023
Date of Report: 05-25-2023

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	BZ-01: 1.01 By 1.01.04A			BZ-02: 1.02.05B		
Comments (see below)	None			None		
Lab ID-Version‡:	15859545-1			15859546-1		
Analysis Date:	05/24/2023			05/24/2023		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria						
Ascospores	6	100	80			
Basidiospores	6	100	80	4	100	53
Bipolaris/Drechslera group						
Cercospora						
Chaetomium						
Cladosporium	9	100	120	2	100	27
Curvularia						
Epicoccum						
Fusarium						
Other brown						
Other colorless						
Penicillium/Aspergillus types†	6	100	80	2	100	27
Pyricularia						
Rusts						
Smuts, Periconia, Myxomycetes						
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)††	1+			1+		
Hyphal fragments/m3	27			< 13		
Pollen/m3	13			< 13		
Skin cells (1-4+)	< 1+			< 1+		
Sample volume (liters)	75			75		
§ TOTAL SPORES/m3			360			110

Comments:

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† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.
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The analytical sensitivity is the spores/m^3 divided by the raw count, expressed in spores/m^3, per spore and per sample.

For more information regarding analytical sensitivity, please contact QA by calling the laboratory.
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Date of Receipt: 05-24-2023
Date of Report: 05-25-2023

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	BZ-03: 1.03A			BZ-04: Hall By 1.06.19		
Comments (see below)	None			None		
Lab ID-Version‡:	15859547-1			15859548-1		
Analysis Date:	05/24/2023			05/24/2023		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria	1	100	13			
Ascospores						
Basidiospores						
Bipolaris/Drechslera group						
Cercospora						
Chaetomium						
Cladosporium	5	100	67			
Curvularia						
Epicoccum						
Fusarium						
Other brown	1	100	13			
Other colorless						
Penicillium/Aspergillus types†	275	25	15,000	5	100	67
Pyricularia						
Rusts						
Smuts, Periconia, Myxomycetes						
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)††	1+			1+		
Hyphal fragments/m3	67			< 13		
Pollen/m3	< 13			< 13		
Skin cells (1-4+)	1+			< 1+		
Sample volume (liters)	75			75		
§ TOTAL SPORES/m3			15,000			67

Comments:

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SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	BZ-05: 1.04 By 1.04.09			BZ-06: 1.01.03		
Comments (see below)	None			None		
Lab ID-Version‡:	15859549-1			15859550-1		
Analysis Date:	05/24/2023			05/24/2023		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria						
Ascospores	1	100	13	3	100	40
Basidiospores				3	100	40
Bipolaris/Drechslera group						
Cercospora						
Chaetomium						
Cladosporium	2	100	27			
Curvularia						
Epicoccum						
Fusarium						
Other brown				1	100	13
Other colorless						
Penicillium/Aspergillus types†	7	100	93	11	100	150
Pyricularia						
Rusts						
Smuts, Periconia, Myxomycetes						
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)††	1+			1+		
Hyphal fragments/m3	< 13			27		
Pollen/m3	< 13			< 13		
Skin cells (1-4+)	< 1+			< 1+		
Sample volume (liters)	75			75		
§ TOTAL SPORES/m3			130			240

Comments:

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SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	BZ-07: 3.01.02			BZ-08: 3.12.04C		
Comments (see below)	None			None		
Lab ID-Version‡:	15859551-1			15859552-1		
Analysis Date:	05/24/2023			05/24/2023		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria						
Ascospores						
Basidiospores				1	100	13
Bipolaris/Drechslera group						
Cercospora						
Chaetomium						
Cladosporium	1	100	13	2	100	27
Curvularia						
Epicoccum						
Fusarium						
Other brown	1	100	13			
Other colorless						
Penicillium/Aspergillus types†	2	100	27	5	100	67
Pyricularia						
Rusts						
Smuts, Periconia, Myxomycetes						
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)††	< 1+			1+		
Hyphal fragments/m3	< 13			< 13		
Pollen/m3	< 13			< 13		
Skin cells (1-4+)	< 1+			< 1+		
Sample volume (liters)	75			75		
§ TOTAL SPORES/m3			53			110

Comments:

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.
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SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	BZ-09: 3.08.07			BZ-10: 3.07.02		
Comments (see below)	None			None		
Lab ID-Version‡:	15859553-1			15859554-1		
Analysis Date:	05/24/2023			05/24/2023		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria						
Ascospores	1	100	13			
Basidiospores	1	100	13	1	100	13
Bipolaris/Drechslera group						
Cercospora						
Chaetomium						
Cladosporium						
Curvularia						
Epicoccum						
Fusarium						
Other brown	1	100	13			
Other colorless						
Penicillium/Aspergillus types†						
Pyricularia						
Rusts						
Smuts, Periconia, Myxomycetes						
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)††	< 1+			< 1+		
Hyphal fragments/m3	27			< 13		
Pollen/m3	< 13			< 13		
Skin cells (1-4+)	< 1+			< 1+		
Sample volume (liters)	75			75		
§ TOTAL SPORES/m3			40			13

Comments:

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SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	BZ-11: 3.04.20			BZ-12: 3.04.10		
Comments (see below)	None			None		
Lab ID-Version‡:	15859555-1			15859556-1		
Analysis Date:	05/24/2023			05/24/2023		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria	1	100	13			
Ascospores	2	100	27			
Basidiospores	2	100	27			
Bipolaris/Drechslera group						
Cercospora						
Chaetomium						
Cladosporium	2	100	27	14	100	190
Curvularia				1	100	13
Epicoccum						
Fusarium						
Other brown				3	100	40
Other colorless						
Penicillium/Aspergillus types†	4	100	53			
Pyricularia						
Rusts						
Smuts, Periconia, Myxomycetes				1	100	13
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)††	1+			1+		
Hyphal fragments/m3	< 13			110		
Pollen/m3	< 13			< 13		
Skin cells (1-4+)	< 1+			< 1+		
Sample volume (liters)	75			75		
§ TOTAL SPORES/m3			150			250

Comments:

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SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	BZ-13: 3.01.05			BZ-14: 2.01 By 2.01.02A		
Comments (see below)	None			None		
Lab ID-Version‡:	15859557-1			15859558-1		
Analysis Date:	05/24/2023			05/24/2023		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria						
Ascospores						
Basidiospores	2	100	27	2	100	27
Bipolaris/Drechslera group						
Cercospora						
Chaetomium						
Cladosporium						
Curvularia						
Epicoccum						
Fusarium						
Other brown						
Other colorless				1	100	13
Penicillium/Aspergillus types†	1	100	13			
Pyricularia						
Rusts						
Smuts, Periconia, Myxomycetes						
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)††	1+			1+		
Hyphal fragments/m3	< 13			< 13		
Pollen/m3	< 13			< 13		
Skin cells (1-4+)	< 1+			< 1+		
Sample volume (liters)	75			75		
§ TOTAL SPORES/m3			40			40

Comments:

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SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	BZ-15: 2.02			BZ-16: North By 2.08A		
Comments (see below)	None			None		
Lab ID-Version‡:	15859559-1			15859560-1		
Analysis Date:	05/24/2023			05/24/2023		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria						
Ascospores						
Basidiospores	1	100	13	1	100	13
Bipolaris/Drechslera group						
Cercospora						
Chaetomium						
Cladosporium	1	100	13			
Curvularia						
Epicoccum						
Fusarium						
Other brown						
Other colorless	1	100	13			
Penicillium/Aspergillus types†				2	100	27
Pyricularia						
Rusts						
Smuts, Periconia, Myxomycetes						
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)††	1+			< 1+		
Hyphal fragments/m3	< 13			< 13		
Pollen/m3	< 13			< 13		
Skin cells (1-4+)	< 1+			< 1+		
Sample volume (liters)	75			75		
§ TOTAL SPORES/m3			40			40

Comments:

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SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	BZ-17: 2.08-Dome			BZ-18: 2.04		
Comments (see below)	None			None		
Lab ID-Version‡:	15859561-1			15859562-1		
Analysis Date:	05/24/2023			05/24/2023		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria						
Ascospores						
Basidiospores	1	100	13	1	100	13
Bipolaris/Drechslera group						
Cercospora						
Chaetomium						
Cladosporium						
Curvularia						
Epicoccum						
Fusarium						
Other brown						
Other colorless						
Penicillium/Aspergillus types†	1	100	13	1	100	13
Pyricularia						
Rusts						
Smuts, Periconia, Myxomycetes						
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)††	< 1+			1+		
Hyphal fragments/m3	< 13			< 13		
Pollen/m3	< 13			< 13		
Skin cells (1-4+)	< 1+			< 1+		
Sample volume (liters)	75			75		
§ TOTAL SPORES/m3			27			27

Comments:

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Client: STC Environmental Services Inc.
C/O: Michael T. Trevino
Re: 230171; ITC Bldg.

Date of Sampling: 05-22-2023
Date of Receipt: 05-24-2023
Date of Report: 05-25-2023

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	BZ-19: 2.05			OUT-20: Outside The Bldg.		
Comments (see below)	None			None		
Lab ID-Version‡:	15859563-1			15859564-1		
Analysis Date:	05/24/2023			05/25/2023		
	raw ct.	% read	spores/m3	raw ct.	% read	spores/m3
Alternaria				17	100	230
Ascospores				40	25	2,100
Basidiospores	1	100	13	35	25	1,900
Bipolaris/Drechslera group				1	100	13
Cercospora				2	100	27
Chaetomium						
Cladosporium				46	100	610
Curvularia				1	100	13
Epicoccum				1	100	13
Fusarium				5	100	67
Other brown				2	100	27
Other colorless				3	100	40
Penicillium/Aspergillus types†	1	100	13	14	100	190
Pyricularia				3	100	40
Rusts						
Smuts, Periconia, Myxomycetes						
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)††	< 1+			1+		
Hyphal fragments/m3	< 13			27		
Pollen/m3	< 13			40		
Skin cells (1-4+)	< 1+			< 1+		
Sample volume (liters)	75			75		
§ TOTAL SPORES/m3			27			5,300

Comments:

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.
† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.
††Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

The analytical sensitivity is the spores/m^3 divided by the raw count, expressed in spores/m^3, per spore and per sample.

For more information regarding analytical sensitivity, please contact QA by calling the laboratory.
‡ A "Version" indicated by -"x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x".
§ Total Spores/m3 has been rounded to two significant figures to reflect analytical precision.

Introduction

Molds are a natural and important part of our environment. They are ubiquitous and are found virtually everywhere. Molds produce tiny spores to reproduce. These spores can be found in both indoor and outdoor air and on indoor and outdoor surfaces. When mold spores land on a damp spot, they may begin growing and digesting whatever they are growing on in order to survive, leading to adverse conditions. In response to increasing public concern, a number of government authorities, including the United States EPA, California Department of Health Services and New York City Department of Health, have developed recommendations and guidelines for assessment and remediation of mold. Websites for these organizations can be found at the end of this report.

While it is generally accepted that molds can be allergenic and can lead to adverse health conditions in susceptible people, unfortunately there are no widely accepted or regulated interpretive standards or numerical guidelines for the interpretation of microbial data. The absence of standards often makes interpretation of microbial data difficult and controversial. This report has been designed to provide some basic interpretive information using certain assumptions and facts that have been extracted from a number of peer reviewed texts, such as the American Conference of Governmental Industrial Hygienists (ACGIH). In the absence of standards, the user must determine the appropriateness and applicability of this report to any given situation. Identification of the presence of a particular fungus in an indoor environment does not necessarily mean that the building occupants are or are not being exposed to antigenic or toxic agents.

None of the information contained herein should be construed as medical advice or a call to action for evacuation or remediation. Only a qualified physician should make any decision relative to medical significance.

EMLab P&K did not conduct the site investigation, provide consulting or collect the samples referenced in this report. EMLab P&K's primary involvement in this project is to provide analytical results for the samples submitted. The data presented in this report are based on the samples and accompanying information provided and represents concentrations at a point in time under the conditions sampled.

EMLab P&K's standard terms and conditions govern all aspects of this report.

Materials

Please refer to the chain of custody included with this report.

Methods

1. **Surface Samples – Swab, Dust, Tape and Bulk Samples**
   Swab, Dust and Tape samples are mounted on a glass slide and observed under a bright field microscope for either Qualitative or Quantitative Examination. A bulk sample is also simultaneously observed under a stereomicroscope to look for signs of any visible discoloration or fungal growth, which is then mounted and observed under a bright field microscope for either Qualitative or Quantitative Examination. The samples are analyzed at a minimum of 200X magnification and up to a 1000X magnification. In the qualitative

examination, the prepared samples are observed for the presence of any structures or skewing of spore distribution that may indicate growth in the sample being analyzed. In the quantitative examination, the mold spores detected in the sample are counted and reported as spores per cm^2, spores per gram (or 1000mg), or spores per swab/wipe, etc depending on the sample type. These methodologies do not differentiate between viable and non-viable fungal spores.

2. Air Samples- Spore Trap Device

Spore traps are a unique sampling device designed for the rapid collection and analysis of a wide range of airborne particulates, including fungal spores. While analyzing the sample, the analyst takes a number of variables into account to select the proper analytical method to accurately determine the densities of the various spores on the trace. The densities of the debris and the spores on the trace will determine the approach to analyzing the sample. In general, the sample is directly mounted under the microscope and the various airborne particles detected are counted at a minimum of 200X magnification and up to 1000X magnification, with the entire trace (100% of the sample) being analyzed at 200X or 600X. This method does not differentiate between viable and non-viable fungal spores. This technique does not allow for the differentiation between *Aspergillus* and *Penicillium* spores. Additionally, depending on morphology, other non-distinctive spores are reported in categories such as ascospores or basidiospores. All slides are graded with the following debris scale for data qualification.

Debris Rating	Description	Interpretation
None	No particles detected.	No particulates on slide. The absence of particulates could indicate improper sampling as most air samples typically capture some particles.
<1+	Good visibility. A few particles detected.	Reported values are not affected by debris.
1+	Good visibility. No crowding of particles.	
2+	Decent visibility. Particles beginning to crowd.	Non-microbial particulates can mask the presence of fungal spores. As a result, actual values could be higher than the numbers reported. Higher debris ratings increase the probability of this bias.
3+	Decent visibility. Particles beginning to crowd.	
4+	Poor visibility. Particles beginning to overlap.	Excessive debris detected in the sample. Counts reported may vary drastically and actual values could be higher than the numbers reported. The sample should be collected at a shorter time interval, or other measures taken to reduce the collection of non-microbial debris. In addition, a >4+ rating will only allow for a count from the perimeter of the slide.
>4+	Poor visibility. Particles overlapping.	

3. Comments

Comments identify issues or events that are relevant to your analytical results. A comment includes information about any peculiar observation or situation encountered while analyzing the sample. In each case, the comments provide significant information vital to the interpretation of the laboratory data.

4. Data Interpretation

According to ACGIH, "Data from individual sampling episodes is often interpreted with respect to baseline data from other environments or the same environment under anticipated low exposure conditions." In the absence of established acceptable exposure limits, it is often necessary to use a comparison standard when interpreting data. In this instance, it will be necessary to sample the suspect area as well as a non-suspect area.

According to ACGIH, "...active fungal growth in indoor environments is inappropriate and may lead to exposure and adverse health effects."

a. Total Fungal Spores

According to ACGIH, ".... differences that can detected with manageable sample sizes are likely to be in 10- fold multiplicative steps (e.g., 100 versus 1000...)". Following this logic, if total fungal spores are ten (10) times greater in the sample from a suspect area than in the negative control sample collected from a non-suspect area, then that sample area may be a fungal amplification site.

b. Mycelial Fragments

Mycelium is a fungal mass that constitutes the vegetative or living body of a fungus. Following the same logic above, if total mycelial fragments are ten (10) times greater in the suspect sample than in the negative control, then the sample area is considered to be a fungal amplification site. The presence of mycelial fragments provides evidence of microbial growth.

c. Mycotoxins

Molds can produce toxic substances called mycotoxins. More than 200 mycotoxins have been identified from common molds, and many more remain to be identified. Some of the molds that are known to produce mycotoxins are commonly found in moisture-damaged buildings. Exposure pathways for mycotoxins can include inhalation, ingestion, or skin contact. Although some mycotoxins are well known to affect humans and have been shown to be responsible for human health effects, for many mycotoxins, little information is available, and in some cases research is ongoing. Some molds can produce several toxins, and some molds produce mycotoxins only under certain environmental conditions. The presence of mold in a building does not necessarily mean that mycotoxins are present or that they are present in large quantities.

d. Water Indicator Molds

Certain authorities identify certain molds whose presence indicates excessive moisture. The presence of a few spores of indicator mold should be interpreted with caution. Additionally, it should be recognized that these named molds are not necessarily the only ones of potential significance.

e. Mold Glossary

Specific characteristics of the individual molds listed in the report are presented in Table 1.

f. Useful Resources

i. Guidelines on Assessment and Remediation of Fungi in Indoor Environments, New York City Department of Health. www1.nyc.gov/assets/doh/downloads/pdf/epi/epi-mold-guidelines.pdf

ii. Facts about Mold, New York City Department of Health.www1.nyc.gov/assets/doh/downloads/pdf/epi/mold-brochure.pdf

iii. Mold Resources, United States Environmental Protection Agency. http://www.epa.gov/mold/moldresources.html
iv. Mold in My Home, What do I do? California Department of Health Services. http://www.lapublichealth.org/eh/docs/housing/brochure/moldhome.pdf

Table 1: Summary of Specific Mold Characteristics

Fungi	Environmental Indicator		Typically Found
Alternaria			*Alternaria* is one of the more common fungi found in nature. It is found growing indoors on a variety of substrates including wallboards, painted walls, etc.
Arthrinium			*Arthrinium* is a saprobe and is found on plants. It is rarely found growing indoors.
Ascospores			Ascospores are ubiquitous in nature and are commonly found in the outdoor environment. Some fungi that belong to the ascomycete family include the sexual forms of *Penicillium/ Aspergillus*, *Chaetomium*, etc that may be frequently found growing on damp substrates.
Aureobasidium			*Aureobasidium* is commonly found in a variety of soils. Indoors, it is commonly found where moisture accumulates, especially bathrooms, and kitchens, on shower curtains, tile grout, windowsills, textiles, and liquid waste materials.
Basidiospores			Basidiospores are Saprophytes and plant pathogens and are commonly found in gardens, forests, and woodlands. They also include organisms that are the agent of "dry rot," and other fungi that cause white and brown wood rot, which may grow and destroy the structural wood of buildings.
Bipolaris/ Dreschlera			*Bipolaris* and *Dreschlera* are usually found associated with plant debris, and soil. They are plant pathogens of numerous plants, particularly grasses. *Bipolaris* and *Dreschlera* can grow indoors on a variety of substrates.
Botrytis			*Botrytis* is commonly found in tropical and temperate climates growing on vegetative matter. They may be found indoors in conjugation with indoor plants, fruits and vegetables.
Chaetomium			*Chaetomium* is often found on materials containing cellulose such as sheetrock paper, or other wet materials.
Cladosporium			*Cladosporium* is a common outdoor mold. They are commonly found on dead plants, food, textiles, and a variety of other surfaces. Indoors, they can grow on a variety of substrates including textiles, wood, moist windowsills, etc. It can grow at 0°C and is associated with refrigerated foods.
Curvularia			*Curvularia* is found on plant materials and is considered a saprobe. Indoors, they can grow on a variety of substrates.
Epicoccum			*Epicoccum* is a saprophyte and considered a weekly parasitic secondary invader of plants. They tend to colonize continuously damp materials such as damp wallboard and fabrics.
Fusarium			*Fusarium* requires very wet conditions and is frequently isolated from plants and grains. They colonize continuously damp materials such as damp wallboard and water reservoirs for humidifiers and drip pans.

Memnoniella			*Memnoniella* can be found growing on a variety of cellulose-containing materials.
Nigrospora			*Nigrospora* is especially abundant in warm climates and is rarely found growing indoors.
Oidium/ Peronospora			*Oidium* and *Peronospora* are plant pathogens and are not found growing indoors.
Penicillium/ Aspergillus			*Penicillium* and *Aspergillus* are ubiquitous in environment. *Aspergillus* tends to colonize continuously damp materials such as damp wallboard and fabrics. *Penicillium* is commonly found in house dusts, wallpaper, decaying fabrics, moist clipboards, etc.
Pithomyces/ Ulocladium			*Pithomyces* is commonly found on grass and decaying plant material and are rarely found growing indoors. *Ulocladium* has a high water requirement and therefore colonizes continuously damp materials such as damp wallboard and fabrics.
Rusts			Rusts are plant pathogens and only grow on host plants.
Smuts/ *Periconia*/ Myxomycetes			Smuts and Myxomycetes are parasitic plant pathogens that require a living host. Smuts do not usually grow indoors. *Periconia* are rarely found growing indoors. Myxomycetes are occasionally found indoors, but rarely growing.
Stachybotrys			*Stachybotrys* are commonly found indoors on wet materials containing cellulose, such as wallboard, jute, wicker, straw baskets, and other paper materials.
Stemphylium			*Stemphylium* is either parasitic or saprophytic and is rarely found growing indoors.
Torula			*Torula* can grow indoors on cellulose containing materials such as wallboard, jute, wicker, straw baskets, and other paper materials.
Other brown/ colorless			An uncharacteristic fungal spore that does not lend itself to classification via direct microscopy.

 Potential Water Intrusion/Indicator Mold Capable of Mycotoxin Production

 Potential Water Intrusion/Indicator Mold

Quality Programs

The EMLab P&K's laboratory network is staffed with highly trained analysts, the majority of which hold advanced degrees. The reliability of test results depends on many factors such as the personnel performing the tests, environmental conditions, selection and validation of test methods, equipment functioning, as well as the sampling, storage and handling of test items, all of which are a reflection of the overall quality system of the laboratory.

EMLab P&K has modeled its quality system after ISO 17025, General Requirements for the Competence of Testing and Calibration Laboratories, one of the most stringent sets of standards in the industry, to ensure that its customers receive the highest standard of accuracy, reliability, and impartiality that they have come to expect from the leader in the environmental industry. EMLab P&K's laboratories adherence to the standards set forth in ISO 17025 has been validated and formally recognized through accreditations granted by an independent outside agency, American Industrial Hygiene Association Laboratory Accreditation Program, LLC (AIHA-LAP, LLC), on a site by site basis. As an additional measure to demonstrate its competency to perform the analyses it offers to its clients, EMLab P&K laboratories

also participate in a variety of different proficiency testing programs, including the Environmental Microbiology Proficiency Analytical Testing Program (EMPAT) sponsored by the American Industrial Hygiene Association Proficiency Analytical Testing Programs.

As part of our continuous commitment to excellence, EMLab P&K laboratories are also inspected, licensed and/or accredited by a number of governmental agencies and independent associations in addition to those already mentioned above. The scope of services, accreditation certificates, and proficiency results can all be accessed at www.emlabpk.com.

References

1. Bioaerosols: Assessment and Control. Janet Macher, Ed., American Conference of Government Industrial Hygienists, Cincinnati, OH (1999).

2. EPA: The Inside Story. A Guide to Indoor Air Quality, United States Environmental Protection Agency and the United States Consumer Product Safety Commission, Washington DC (1995).

3. Health Canada: Exposure Guidelines for Residential Indoor Air Quality. Environmental Health Directorate. Health Protection Branch, Health Canada, Ottawa, Ontario (1989).

4. IIRC: Standard and Reference Guide for Professional Water Damage Restoration, 2nd Ed. Institute of Inspection, Cleaning and Restoration, Vancouver, WA (1999).

5. Field Guide for the Determination of Bio logical Contaminants in Environmental Samples. American Industrial Hygiene Association, Fairfax, VA (1996).

6. Standards of Practice for the Assessment of Indoor Environmental Quality, Volume I: Mold Sampling, Assessment of Mold Contamination. Indoor Environmental Standards Organization (2002).

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Re: 230171; ITC Bldg.

Date of Sampling: 05-22-2023
Date of Receipt: 05-24-2023
Date of Report: 05-25-2023

MoldRANGE™, Local Climate; Extended Outdoor Comparison

Outdoor Location: OUT-20, Outside The Bldg.

Fungi Identified	Outdoor data	Typical Outdoor Data for: May in South† EMLab Regional Climate code¹ A Annual Temp, A Elev., B Rain, B Temp. Range (n‡=629)						Typical Outdoor Data for: The entire year in South† EMLab Regional Climate code¹ A Annual Temp, A Elev., B Rain, B Temp. Range (n‡=6533)					
Project zip code 78205	spores/m3	very low	low	med	high	very high	freq %	very low	low	med	high	very high	freq %
Generally able to grow indoors*													
Alternaria	230	27	40	110	270	400	77	13	40	80	210	350	67
Bipolaris/Drechslera group	13	13	13	27	53	110	39	13	13	53	110	160	39
Chaetomium	-	13	13	13	200	380	6	13	13	13	27	40	6
Cladosporium	610	210	480	1,400	4,100	6,800	98	160	330	1,100	3,700	6,800	97
Curvularia	13	13	13	22	53	67	19	13	13	40	80	160	29
Epicoccum	13	13	13	40	110	160	44	13	13	40	67	120	27
Fusarium	67	13	13	40	67	87	4	13	13	53	150	200	9
Nigrospora	-	13	13	27	53	67	26	13	13	53	110	170	40
Other brown	27	13	13	24	53	110	23	13	13	40	53	110	22
Other colorless	40	13	16	53	90	160	5	13	13	53	120	190	5
Penicillium/Aspergillus types	190	53	67	210	750	1,100	64	53	80	210	640	1,100	67
Stachybotrys	-	-	-	-	-	-	1	13	13	13	27	63	1
Torula	-	13	13	27	67	170	14	13	13	40	67	120	12
Seldom found growing indoors**													
Ascospores	2,100	80	160	530	1,600	3,000	96	53	110	290	910	1,700	88
Basidiospores	1,900	110	240	800	2,500	5,100	98	67	160	480	2,000	4,200	93
Cercospora	27	13	13	50	110	160	39	13	22	53	160	270	31
Pyricularia	40	13	13	13	33	53	7	13	13	27	67	110	6
Rusts	-	13	13	27	53	110	28	13	13	27	53	110	14
Smuts, Periconia, Myxomycetes	-	27	40	110	270	480	82	27	40	110	270	440	77
§ TOTAL SPORES/m3	5,300												

¹EMLab Regional Climate codes are a climate classification scheme for regional geographic areas containing multiple states. The MoldRANGE™ Local Climate report uses the sampling location zip code to identify the EMLab Regional Climate code in that area. Using information available from the NOAA weather database, the EMLab Regional Climate code sharpens the precision of the MoldRANGE™reporting system, providing more reliable estimates of the range and average concentrations of the different airborne fungal spore types for each region. Additional information on the EMLab Regional Climate code system can be found on the last page of this report.

†The Typical Outdoor Data represents the typical outdoor spore levels across the region's group of states for the time period and EMLab Regional Climate code indicated. The last column represents the frequency of occurrence. The very low, low, med, high, and very high values represent the 10, 20, 50, 80, and 90 percentile values of the spore type when it is detected. For example, if the frequency of occurrence is 63% and the low value is 53, it would mean that the given spore type is detected 63% of the time and, when detected, 20% of the time it is present in levels above the detection limit and below 53 spores/m3. These values are updated periodically and if not enough data is available to make a statistically meaningful assessment, it is indicated with a dash.

‡ n is the sample size used to calculate the MoldRANGE™ Local Climate data summarized in the table.

* The spores in this category are generally capable of growing on wet building materials in addition to growing outdoors. Building related growth is dependent upon the fungal type, moisture level, type of material, and other factors. *Cladosporium* is one of the predominant spore types worldwide and is frequently present in high numbers. *Penicillium/Aspergillus* species colonize both outdoor and indoor wet surfaces rapidly and are very easily dispersed. Other genera are usually present in lesser numbers.

** These fungi are generally not found growing on wet building materials. For example, the rusts and smuts are obligate plant pathogens. However, in each group there are notable exceptions. For example, agents of wood decay are members of the basidiomycetes and high counts of a single morphological type of basidiospore on an inside sample should be considered significant.

§ Total Spores/m3 has been rounded to two significant figures to reflect analytical precision.

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Understanding EMLab Regional Climate Codes

Outdoor airborne spore concentrations are strongly influenced by climate and weather patterns, often resulting in pronounced seasonal and diurnal cycles (Burge 1995). The seasonal climatic changes directly affect the growth cycle of plants, thereby influencing fungal growth, spore maturation, and release cycles. By evaluating outdoor spore concentrations across similar climatic zones rather than for the state as a whole, it is possible to provide a more representative estimate of typical outdoor spore levels and frequency of occurrence for different airborne fungal spore types in a given area.

The EMLab Regional Climate code system is a novel classification system that uses data from the NOAA - National Oceanic and Atmospheric Administration database to define unique climate zones. The following climate variables, for each regional zip code, are obtained from NOAA and assigned a letter code of A (above the regional average for that variable) or B (below the regional average for that variable):

1. Annual High Temperature
2. Elevation
3. Rainfall/Precipitation
4. Monthly Temperature Range

The result is a 4-character code assigned to each statewide zip code, referred to as the Regional Climate Code. Below are some examples of decoded Regional Climate Codes:

AAAA = Above avg. Annual High Temperature, Above avg. Elevation, Above avg. Rainfall/Precipitation, Above avg. Monthly Temperature Range
AABB = Above avg. Annual High Temperature, Above avg. Elevation, Below avg. Rainfall/Precipitation, Below avg. Monthly Temperature Range
BBAA = Below avg. Annual High Temperature, Below avg. Elevation, Above avg. Rainfall/Precipitation, Above avg. Monthly Temperature Range

The actual outdoor air sample data from matching regional climate codes in each group of states are then compiled in a manner relating typical spore concentrations and frequency of occurrence.

The data presented in this report is from the South Region which includes the states of: AR, KS, LA, MS, OK, and TX

The NOAA regional climate variables were selected by mapping data points from a subset of approximately 145,000 weather and geographic database entries to over 80,000 outdoor spore trap samples with known zip codes and assessing them using orthogonal array experimental design techniques. The results were then compared to the typical ranges of spore types found when grouping zip codes using the Koppen-Geiger climatic classification system; a commonly used climatic system that provides an objective numerical definition in terms of climatic elements such as temperature, rainfall, and other seasonal characteristics . The EMLab Regional Climate codes showed improved granularity and refinement of the zip code groupings, implying a better representation of the expected range of spore types to be found within an individual zip code.

The values on this report were calculated by obtaining the four variables listed above from the over 585 million data points of weather and geographic information available in the NOAA database, and determining the frequencies and percentile values of spore types by utilizing over 180,000 Eurofins EMLab P&K outdoor spore trap samples with known zip codes.

This report groups regional zip codes in relation to these EMLab Regional Climate codes and summarizes MoldRANGE™ data by month and year within each EMLab Regional Climate code.

References:

Burge, Harriet, A. Bioaerosols: Boca Raton: Lewis Publishers, pp. 163-171, 1995.

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MoldSTAT™: Supplementary Statistical Spore Trap Report

Outdoor Summary: OUT-20: Outside The Bldg.

Species detected	Outdoor sample spores/m3 (<100 / 1K / 10K / >100K)	Typical outdoor ranges (North America)	Freq. %
Alternaria	230	7 - 27 - 390	38
Ascospores	2,100	13 - 210 - 6,000	74
Basidiospores	1,900	13 - 430 - 24,000	89
Bipolaris/Drechslera group	13	7 - 13 - 190	13
Cercospora	27	7 - 33 - 510	12
Cladosporium	610	27 - 430 - 7,500	87
Curvularia	13	7 - 27 - 600	18
Epicoccum	13	7 - 27 - 270	19
Fusarium	67	7 - 27 - 290	3
Other brown	27	7 - 20 - 160	27
Other colorless	40	7 - 27 - 530	4
Penicillium/Aspergillus types	190	17 - 190 - 2,700	61
Pyricularia	40	7 - 20 - 280	5
Smuts, Periconia, Myxomycetes	< 13	7 - 53 - 850	63
Total	5,300		

The "Typical outdoor ranges" and "Freq. %" columns show the typical low, medium, and high spore counts per cubic meter and the frequency of occurrence for the given spore type. The low, medium, and high values represent the 2.5, 50, and 97.5 percentile values when the spore type is detected. For example, if the low value is 53 and the frequency of occurence is 63%, it would mean that we typically detect the given spore type on 63 percent of all outdoor samples and, when detected, 2.5% of the time it is present in levels below 53 spores/m3.

Indoor Samples

Location: BZ-01: 1.01 By 1.01.04A

% of outdoor total spores/m3	Friedman chi-square* (indoor variation)	Agreement ratio** (indoor/outdoor)	Spearman rank correlation*** (indoor/outdoor)	MoldSCORE**** (indoor/outdoor)
Result: 6%	dF: 18 Result: 14.4982 Critical value: 28.8693 Inside Similar: Yes	Result: 0.4706	dF: 13 Result: 0.7830 Critical value: 0.4780 Outside Similar: Yes	Score: 111 Result: Low

Species Detected	Spores/m3 (<100 / 1K / 10K / >100K)
Ascospores	80
Basidiospores	80
Cladosporium	120
Penicillium/Aspergillus types	80
Total	360

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MoldSTAT™: Supplementary Statistical Spore Trap Report

Location: BZ-02: 1.02.05B

% of outdoor total spores/m3	Friedman chi-square* (indoor variation)	Agreement ratio** (indoor/outdoor)	Spearman rank correlation*** (indoor/outdoor)	MoldSCORE**** (indoor/outdoor)
Result: 2%	dF: 18 Result: 14.4982 Critical value: 28.8693 Inside Similar: Yes	Result: 0.3750	dF: 13 Result: 0.6401 Critical value: 0.4780 Outside Similar: Yes	Score: 104 Result: Low

Species Detected	Spores/m3 <100 1K 10K >100K	
Basidiospores		53
Cladosporium		27
Penicillium/Aspergillus types		27
Total		110

Location: BZ-03: 1.03A

% of outdoor total spores/m3	Friedman chi-square* (indoor variation)	Agreement ratio** (indoor/outdoor)	Spearman rank correlation*** (indoor/outdoor)	MoldSCORE**** (indoor/outdoor)
Result: 286%	dF: 18 Result: 14.4982 Critical value: 28.8693 Inside Similar: Yes	Result: 0.4706	dF: 13 Result: 0.4313 Critical value: 0.4780 Outside Similar: No	Score: 300 Result: High

Species Detected	Spores/m3 <100 1K 10K >100K	
Alternaria		13
Cladosporium		67
Other brown		13
Penicillium/Aspergillus types		15,000
Total		15,000

Location: BZ-04: Hall By 1.06.19

% of outdoor total spores/m3	Friedman chi-square* (indoor variation)	Agreement ratio** (indoor/outdoor)	Spearman rank correlation*** (indoor/outdoor)	MoldSCORE**** (indoor/outdoor)
Result: 1%	dF: 18 Result: 14.4982 Critical value: 28.8693 Inside Similar: Yes	Result: 0.1429	dF: 13 Result: 0.4725 Critical value: 0.4780 Outside Similar: No	Score: 110 Result: Low

Species Detected	Spores/m3 <100 1K 10K >100K	
Penicillium/Aspergillus types		67
Total		67

Eurofins EPK Built Environment Testing, LLC
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(866) 871-1984 www.eurofinsus.com/Built

Client: STC Environmental Services Inc.
C/O: Michael T. Trevino
Re: 230171; ITC Bldg.

Date of Sampling: 05-22-2023
Date of Receipt: 05-24-2023
Date of Report: 05-25-2023

MoldSTAT™: Supplementary Statistical Spore Trap Report

Location: BZ-05: 1.04 By 1.04.09

% of outdoor total spores/m3	Friedman chi-square* (indoor variation)	Agreement ratio** (indoor/outdoor)	Spearman rank correlation*** (indoor/outdoor)	MoldSCORE**** (indoor/outdoor)
Result: 2%	dF: 18 Result: 14.4982 Critical value: 28.8693 Inside Similar: Yes	Result: 0.3750	dF: 13 Result: 0.6415 Critical value: 0.4780 Outside Similar: Yes	Score: 114 Result: Low

Species Detected	Spores/m3 (<100 / 1K / 10K / >100K)
Ascospores	13
Cladosporium	27
Penicillium/Aspergillus types	93
Total	130

Location: BZ-06: 1.01.03

% of outdoor total spores/m3	Friedman chi-square* (indoor variation)	Agreement ratio** (indoor/outdoor)	Spearman rank correlation*** (indoor/outdoor)	MoldSCORE**** (indoor/outdoor)
Result: 4%	dF: 18 Result: 14.4982 Critical value: 28.8693 Inside Similar: Yes	Result: 0.4706	dF: 13 Result: 0.5865 Critical value: 0.4780 Outside Similar: Yes	Score: 122 Result: Low

Species Detected	Spores/m3 (<100 / 1K / 10K / >100K)
Ascospores	40
Basidiospores	40
Other brown	13
Penicillium/Aspergillus types	150
Total	240

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C/O: Michael T. Trevino
Re: 230171; ITC Bldg.

Date of Sampling: 05-22-2023
Date of Receipt: 05-24-2023
Date of Report: 05-25-2023

MoldSTAT™: Supplementary Statistical Spore Trap Report

Location: BZ-07: 3.01.02

% of outdoor total spores/m3	Friedman chi-square* (indoor variation)	Agreement ratio** (indoor/outdoor)	Spearman rank correlation*** (indoor/outdoor)	MoldSCORE**** (indoor/outdoor)
Result: 1%	dF: 18 Result: 14.4982 Critical value: 28.8693 Inside Similar: Yes	Result: 0.3750	dF: 13 Result: 0.3681 Critical value: 0.4780 Outside Similar: No	Score: 105 Result: Low

Species Detected	Spores/m3 <100 1K 10K >100K
Cladosporium	13
Other brown	13
Penicillium/Aspergillus types	27
Total	53

Location: BZ-08: 3.12.04C

% of outdoor total spores/m3	Friedman chi-square* (indoor variation)	Agreement ratio** (indoor/outdoor)	Spearman rank correlation*** (indoor/outdoor)	MoldSCORE**** (indoor/outdoor)
Result: 2%	dF: 18 Result: 14.4982 Critical value: 28.8693 Inside Similar: Yes	Result: 0.3750	dF: 13 Result: 0.6113 Critical value: 0.4780 Outside Similar: Yes	Score: 110 Result: Low

Species Detected	Spores/m3 <100 1K 10K >100K
Basidiospores	13
Cladosporium	27
Penicillium/Aspergillus types	67
Total	110

Location: BZ-09: 3.08.07

% of outdoor total spores/m3	Friedman chi-square* (indoor variation)	Agreement ratio** (indoor/outdoor)	Spearman rank correlation*** (indoor/outdoor)	MoldSCORE**** (indoor/outdoor)
Result: < 1%	dF: 18 Result: 14.4982 Critical value: 28.8693 Inside Similar: Yes	Result: 0.3750	dF: 13 Result: 0.5440 Critical value: 0.4780 Outside Similar: Yes	Score: 105 Result: Low

Species Detected	Spores/m3 <100 1K 10K >100K
Ascospores	13
Basidiospores	13
Other brown	13
Total	40

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C/O: Michael T. Trevino
Re: 230171; ITC Bldg.

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Date of Receipt: 05-24-2023
Date of Report: 05-25-2023

MoldSTAT™: Supplementary Statistical Spore Trap Report

Location: BZ-10: 3.07.02

% of outdoor total spores/m3	Friedman chi-square* (indoor variation)	Agreement ratio** (indoor/outdoor)	Spearman rank correlation*** (indoor/outdoor)	MoldSCORE**** (indoor/outdoor)
Result: < 1%	dF: 18 Result: 14.4982 Critical value: 28.8693 Inside Similar: Yes	Result: 0.1429	dF: 13 Result: 0.5797 Critical value: 0.4780 Outside Similar: Yes	Score: 101 Result: Low

Species Detected	Spores/m3 <100 1K 10K >100K
Basidiospores	13
Total	13

Location: BZ-11: 3.04.20

% of outdoor total spores/m3	Friedman chi-square* (indoor variation)	Agreement ratio** (indoor/outdoor)	Spearman rank correlation*** (indoor/outdoor)	MoldSCORE**** (indoor/outdoor)
Result: 2%	dF: 18 Result: 14.4982 Critical value: 28.8693 Inside Similar: Yes	Result: 0.5556	dF: 13 Result: 0.8324 Critical value: 0.4780 Outside Similar: Yes	Score: 108 Result: Low

Species Detected	Spores/m3 <100 1K 10K >100K
Alternaria	13
Ascospores	27
Basidiospores	27
Cladosporium	27
Penicillium/Aspergillus types	53
Total	150

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C/O: Michael T. Trevino
Re: 230171; ITC Bldg.

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Date of Receipt: 05-24-2023
Date of Report: 05-25-2023

MoldSTAT™: Supplementary Statistical Spore Trap Report

Location: BZ-12: 3.04.10

% of outdoor total spores/m3	Friedman chi-square* (indoor variation)	Agreement ratio** (indoor/outdoor)	Spearman rank correlation*** (indoor/outdoor)	MoldSCORE**** (indoor/outdoor)
Result: 4%	dF: 18 Result: 14.4982 Critical value: 28.8693 Inside Similar: Yes	Result: 0.3529	dF: 14 Result: 0.0011 Critical value: 0.4593 Outside Similar: No	Score: 122 Result: Low

Species Detected	Spores/m3 (<100 / 1K / 10K / >100K)
Cladosporium	190
Curvularia	13
Other brown	40
Smuts, Periconia, Myxomycetes	13
Total	250

Location: BZ-13: 3.01.05

% of outdoor total spores/m3	Friedman chi-square* (indoor variation)	Agreement ratio** (indoor/outdoor)	Spearman rank correlation*** (indoor/outdoor)	MoldSCORE**** (indoor/outdoor)
Result: < 1%	dF: 18 Result: 14.4982 Critical value: 28.8693 Inside Similar: Yes	Result: 0.2667	dF: 13 Result: 0.5687 Critical value: 0.4780 Outside Similar: Yes	Score: 102 Result: Low

Species Detected	Spores/m3 (<100 / 1K / 10K / >100K)
Basidiospores	27
Penicillium/Aspergillus types	13
Total	40

Location: BZ-14: 2.01 By 2.01.02A

% of outdoor total spores/m3	Friedman chi-square* (indoor variation)	Agreement ratio** (indoor/outdoor)	Spearman rank correlation*** (indoor/outdoor)	MoldSCORE**** (indoor/outdoor)
Result: < 1%	dF: 18 Result: 14.4982 Critical value: 28.8693 Inside Similar: Yes	Result: 0.2667	dF: 13 Result: 0.4863 Critical value: 0.4780 Outside Similar: Yes	Score: 105 Result: Low

Species Detected	Spores/m3 (<100 / 1K / 10K / >100K)
Basidiospores	27
Other colorless	13
Total	40

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C/O: Michael T. Trevino
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MoldSTAT™: Supplementary Statistical Spore Trap Report

Location: BZ-15: 2.02

% of outdoor total spores/m3	Friedman chi-square* (indoor variation)	Agreement ratio** (indoor/outdoor)	Spearman rank correlation*** (indoor/outdoor)	MoldSCORE**** (indoor/outdoor)
Result: < 1%	dF: 18 Result: 14.4982 Critical value: 28.8693 Inside Similar: Yes	Result: 0.3750	dF: 13 Result: 0.5440 Critical value: 0.4780 Outside Similar: Yes	Score: 105 Result: Low

Species Detected	Spores/m3 <100 1K 10K >100K
Basidiospores	13
Cladosporium	13
Other colorless	13
Total	40

Location: BZ-16: North By 2.08A

% of outdoor total spores/m3	Friedman chi-square* (indoor variation)	Agreement ratio** (indoor/outdoor)	Spearman rank correlation*** (indoor/outdoor)	MoldSCORE**** (indoor/outdoor)
Result: < 1%	dF: 18 Result: 14.4982 Critical value: 28.8693 Inside Similar: Yes	Result: 0.2667	dF: 13 Result: 0.5522 Critical value: 0.4780 Outside Similar: Yes	Score: 104 Result: Low

Species Detected	Spores/m3 <100 1K 10K >100K
Basidiospores	13
Penicillium/Aspergillus types	27
Total	40

Location: BZ-17: 2.08-Dome

% of outdoor total spores/m3	Friedman chi-square* (indoor variation)	Agreement ratio** (indoor/outdoor)	Spearman rank correlation*** (indoor/outdoor)	MoldSCORE**** (indoor/outdoor)
Result: < 1%	dF: 18 Result: 14.4982 Critical value: 28.8693 Inside Similar: Yes	Result: 0.2667	dF: 13 Result: 0.5618 Critical value: 0.4780 Outside Similar: Yes	Score: 102 Result: Low

Species Detected	Spores/m3 <100 1K 10K >100K
Basidiospores	13
Penicillium/Aspergillus types	13
Total	27

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C/O: Michael T. Trevino
Re: 230171; ITC Bldg.

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Date of Report: 05-25-2023

MoldSTAT™: Supplementary Statistical Spore Trap Report

Location: BZ-18: 2.04

% of outdoor total spores/m3	Friedman chi-square* (indoor variation)	Agreement ratio** (indoor/outdoor)	Spearman rank correlation*** (indoor/outdoor)	MoldSCORE**** (indoor/outdoor)
Result: < 1%	dF: 18 Result: 14.4982 Critical value: 28.8693 Inside Similar: Yes	Result: 0.2667	dF: 13 Result: 0.5618 Critical value: 0.4780 Outside Similar: Yes	Score: 102 Result: Low

Species Detected	Spores/m3 <100 1K 10K >100K
Basidiospores	13
Penicillium/Aspergillus types	13
Total	27

Location: BZ-19: 2.05

% of outdoor total spores/m3	Friedman chi-square* (indoor variation)	Agreement ratio** (indoor/outdoor)	Spearman rank correlation*** (indoor/outdoor)	MoldSCORE**** (indoor/outdoor)
Result: < 1%	dF: 18 Result: 14.4982 Critical value: 28.8693 Inside Similar: Yes	Result: 0.2667	dF: 13 Result: 0.5618 Critical value: 0.4780 Outside Similar: Yes	Score: 102 Result: Low

Species Detected	Spores/m3 <100 1K 10K >100K
Basidiospores	13
Penicillium/Aspergillus types	13
Total	27

* The Friedman chi-square statistic is a non-parametric test that examines variation in a set of data (in this case, all indoor spore counts). The null hypothesis (H0) being tested is that there is no meaningful difference in the data for all indoor locations. The alternative hypothesis (used if the test disproves the null hypothesis) is that there is a difference between the indoor locations. The null hypothesis is rejected when the result of the test is greater than the critical value. The critical value that is displayed is based on the degrees of freedom (dF) of the test and a significance level of 0.05.

** An agreement ratio is a simple method for assessing the similarity of two samples (in this case the indoor sample and the outdoor summary) based on the spore types present. A score of one indicates that the types detected in one location are the same as that in the other. A score of zero indicates that none of the types detected indoors are present outdoors. Typically, an agreement of 0.8 or higher is considered high.

*** The Spearman rank correlation is a non-parametric test that examines correlation between two sets of data (in this case the indoor location and the outdoor summary). The null hypothesis (H0) being tested is that the indoor and outdoor samples are unrelated. The alternative hypothesis (used if the test disproves the null hypothesis) is that the samples are similar. The null hypothesis is rejected when the result of the test is greater than the critical value. The critical value that is displayed is based on the degrees of freedom (dF) of the test and a significance level of 0.05.

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MoldSTAT™: Supplementary Statistical Spore Trap Report

**** MoldSCORE™ is a specialized method for examining air sampling data. It is a score between 100 and 300, with 100 indicating a greater likelihood that the airborne indoor spores originated from the outside, and 300 indicating a greater likelihood that they originated from an inside source. The Result displayed is based on the numeric score given and will be either Low, Medium, or High, indicating a low, medium, or high likelihood that the spores detected originated from an indoor source. Eurofins EMLab P&Kreserves the right to, and may at anytime, modify or change the MoldScore algorithm without notice.

Interpretation of the data contained in this report is left to the client or the persons who conducted the field work. This report is provided for informational and comparative purposes only and should not be relied upon for any other purpose. "Typical outdoor ranges" are based on the results of the analysis of samples delivered to and analyzed by Eurofins EMLab P&K and assumptions regarding the origins of those samples. Sampling techniques, contaminants infecting samples, unrepresentative samples and other similar or dissimilar factors may affect these results. With the statistical analysis provided, as with all statistical comparisons and analyses, false-positive and false-negative results can and do occur. Eurofins EMLab P&K hereby disclaims any liability for any and all direct, indirect, punitive, incidental, special or consequential damages arising out of the data contained in, or any actions taken or omitted in reliance upon, this report.

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Date of Sampling: 05-22-2023
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MoldSCORE™: Spore Trap Report

Outdoor Sample: OUT-20 Outside The Bldg.

Fungi Identified	Outdoor sample spores/m3 <100 1K 10K >100K	Raw count	Spores/ m3
Generally able to grow indoors*			
Alternaria		17	230
Bipolaris/Drechslera group		1	13
Chaetomium		ND	< 13
Cladosporium		46	610
Curvularia		1	13
Epicoccum		1	13
Fusarium		5	67
Nigrospora		ND	< 13
Other brown		2	27
Other colorless		3	40
Penicillium/Aspergillus types†		14	190
Stachybotrys		ND	< 13
Torula		ND	< 13
Seldom found growing indoors**			
Ascospores		40	2,100
Basidiospores		35	1,900
Cercospora		2	27
Pyricularia		3	40
Rusts		ND	< 13
Smuts, Periconia, Myxomycetes		ND	< 13
Total			**5,267**

Location: BZ-01 1.01 By 1.01.04A

Fungi Identified	Indoor sample spores/m3 <100 1K 10K >100K	Raw count	Spores/ m3	MoldSCORE‡ 100 200 300	Score
Generally able to grow indoors*					
Alternaria		ND	< 13		100
Bipolaris/Drechslera group		ND	< 13		100
Chaetomium		ND	< 13		100
Cladosporium		9	120		105
Curvularia		ND	< 13		100
Nigrospora		ND	< 13		100
Penicillium/Aspergillus types†		6	80		111
Stachybotrys		ND	< 13		100
Torula		ND	< 13		100
Seldom found growing indoors**					
Ascospores		6	80		100
Basidiospores		6	80		100
Rusts		ND	< 13		100
Smuts, Periconia, Myxomycetes		ND	< 13		100
Total			**360**	**Final MoldSCORE**	**111**

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MoldSCORE™: Spore Trap Report

Location: BZ-02 1.02.05B

Fungi Identified	Indoor sample spores/m3 <100 1K 10K >100K	Raw count	Spores/ m3	MoldSCORE‡ 100 200 300	Score
Generally able to grow indoors*					
Alternaria		ND	< 13		100
Bipolaris/Drechslera group		ND	< 13		100
Chaetomium		ND	< 13		100
Cladosporium		2	27		101
Curvularia		ND	< 13		100
Nigrospora		ND	< 13		100
Penicillium/Aspergillus types†		2	27		104
Stachybotrys		ND	< 13		100
Torula		ND	< 13		100
Seldom found growing indoors**					
Ascospores		ND	< 13		100
Basidiospores		4	53		101
Rusts		ND	< 13		100
Smuts, Periconia, Myxomycetes		ND	< 13		100
Total			**107**	**Final MoldSCORE**	**104**

Location: BZ-03 1.03A

Fungi Identified	Indoor sample spores/m3 <100 1K 10K >100K	Raw count	Spores/ m3	MoldSCORE‡ 100 200 300	Score
Generally able to grow indoors*					
Alternaria		1	13		100
Bipolaris/Drechslera group		ND	< 13		100
Chaetomium		ND	< 13		100
Cladosporium		5	67		100
Curvularia		ND	< 13		100
Nigrospora		ND	< 13		100
Other brown		1	13		100
Penicillium/Aspergillus types†		275	15,000		300
Stachybotrys		ND	< 13		100
Torula		ND	< 13		100
Seldom found growing indoors**					
Ascospores		ND	< 13		100
Basidiospores		ND	< 13		100
Rusts		ND	< 13		100
Smuts, Periconia, Myxomycetes		ND	< 13		100
Total			**14,760**	**Final MoldSCORE**	**300**

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Date of Report: 05-25-2023

MoldSCORE™: Spore Trap Report

Location: BZ-04 Hall By 1.06.19

Fungi Identified	Indoor sample spores/m3 <100 1K 10K >100K	Raw count	Spores/ m3	MoldSCORE‡ 100 200 300	Score
Generally able to grow indoors*					
Alternaria		ND	< 13		100
Bipolaris/Drechslera group		ND	< 13		100
Chaetomium		ND	< 13		100
Cladosporium		ND	< 13		100
Curvularia		ND	< 13		100
Nigrospora		ND	< 13		100
Penicillium/Aspergillus types†		5	67		110
Stachybotrys		ND	< 13		100
Torula		ND	< 13		100
Seldom found growing indoors**					
Ascospores		ND	< 13		100
Basidiospores		ND	< 13		100
Rusts		ND	< 13		100
Smuts, Periconia, Myxomycetes		ND	< 13		100
Total			**67**	**Final MoldSCORE**	**110**

Location: BZ-05 1.04 By 1.04.09

Fungi Identified	Indoor sample spores/m3 <100 1K 10K >100K	Raw count	Spores/ m3	MoldSCORE‡ 100 200 300	Score
Generally able to grow indoors*					
Alternaria		ND	< 13		100
Bipolaris/Drechslera group		ND	< 13		100
Chaetomium		ND	< 13		100
Cladosporium		2	27		101
Curvularia		ND	< 13		100
Nigrospora		ND	< 13		100
Penicillium/Aspergillus types†		7	93		114
Stachybotrys		ND	< 13		100
Torula		ND	< 13		100
Seldom found growing indoors**					
Ascospores		1	13		100
Basidiospores		ND	< 13		100
Rusts		ND	< 13		100
Smuts, Periconia, Myxomycetes		ND	< 13		100
Total			**133**	**Final MoldSCORE**	**114**

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Date of Report: 05-25-2023

MoldSCORE™: Spore Trap Report

Location: BZ-06 1.01.03

Fungi Identified	Indoor sample spores/m3 <100 1K 10K >100K	Raw count	Spores/ m3	MoldSCORE‡ 100 200 300	Score
Generally able to grow indoors*					
Alternaria		ND	< 13		100
Bipolaris/Drechslera group		ND	< 13		100
Chaetomium		ND	< 13		100
Cladosporium		ND	< 13		100
Curvularia		ND	< 13		100
Nigrospora		ND	< 13		100
Other brown		1	13		105
Penicillium/Aspergillus types†		11	150		122
Stachybotrys		ND	< 13		100
Torula		ND	< 13		100
Seldom found growing indoors**					
Ascospores		3	40		100
Basidiospores		3	40		100
Rusts		ND	< 13		100
Smuts, Periconia, Myxomycetes		ND	< 13		100
Total			**240**	**Final MoldSCORE**	**122**

Location: BZ-07 3.01.02

Fungi Identified	Indoor sample spores/m3 <100 1K 10K >100K	Raw count	Spores/ m3	MoldSCORE‡ 100 200 300	Score
Generally able to grow indoors*					
Alternaria		ND	< 13		100
Bipolaris/Drechslera group		ND	< 13		100
Chaetomium		ND	< 13		100
Cladosporium		1	13		100
Curvularia		ND	< 13		100
Nigrospora		ND	< 13		100
Other brown		1	13		105
Penicillium/Aspergillus types†		2	27		104
Stachybotrys		ND	< 13		100
Torula		ND	< 13		100
Seldom found growing indoors**					
Ascospores		ND	< 13		100
Basidiospores		ND	< 13		100
Rusts		ND	< 13		100
Smuts, Periconia, Myxomycetes		ND	< 13		100
Total			**53**	**Final MoldSCORE**	**105**

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Date of Report: 05-25-2023

MoldSCORE™: Spore Trap Report

Location: BZ-08 3.12.04C

Fungi Identified	Indoor sample spores/m3 <100 1K 10K >100K	Raw count	Spores/ m3	MoldSCORE‡ 100 200 300	Score
Generally able to grow indoors*					
Alternaria		ND	< 13		100
Bipolaris/Drechslera group		ND	< 13		100
Chaetomium		ND	< 13		100
Cladosporium		2	27		101
Curvularia		ND	< 13		100
Nigrospora		ND	< 13		100
Penicillium/Aspergillus types†		5	67		110
Stachybotrys		ND	< 13		100
Torula		ND	< 13		100
Seldom found growing indoors**					
Ascospores		ND	< 13		100
Basidiospores		1	13		100
Rusts		ND	< 13		100
Smuts, Periconia, Myxomycetes		ND	< 13		100
Total			**107**	**Final MoldSCORE**	**110**

Location: BZ-09 3.08.07

Fungi Identified	Indoor sample spores/m3 <100 1K 10K >100K	Raw count	Spores/ m3	MoldSCORE‡ 100 200 300	Score
Generally able to grow indoors*					
Alternaria		ND	< 13		100
Bipolaris/Drechslera group		ND	< 13		100
Chaetomium		ND	< 13		100
Cladosporium		ND	< 13		100
Curvularia		ND	< 13		100
Nigrospora		ND	< 13		100
Other brown		1	13		105
Penicillium/Aspergillus types†		ND	< 13		100
Stachybotrys		ND	< 13		100
Torula		ND	< 13		100
Seldom found growing indoors**					
Ascospores		1	13		100
Basidiospores		1	13		100
Rusts		ND	< 13		100
Smuts, Periconia, Myxomycetes		ND	< 13		100
Total			**40**	**Final MoldSCORE**	**105**

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(866) 871-1984 www.eurofinsus.com/Built

Client: STC Environmental Services Inc.
C/O: Michael T. Trevino
Re: 230171; ITC Bldg.

Date of Sampling: 05-22-2023
Date of Receipt: 05-24-2023
Date of Report: 05-25-2023

MoldSCORE™: Spore Trap Report

Location: BZ-10 3.07.02

Fungi Identified	Indoor sample spores/m3 <100 1K 10K >100K	Raw count	Spores/ m3	MoldSCORE‡ 100 200 300	Score
Generally able to grow indoors*					
Alternaria		ND	< 13		100
Bipolaris/Drechslera group		ND	< 13		100
Chaetomium		ND	< 13		100
Cladosporium		ND	< 13		100
Curvularia		ND	< 13		100
Nigrospora		ND	< 13		100
Penicillium/Aspergillus types†		ND	< 13		100
Stachybotrys		ND	< 13		100
Torula		ND	< 13		100
Seldom found growing indoors**					
Ascospores		ND	< 13		100
Basidiospores		1	13		101
Rusts		ND	< 13		100
Smuts, Periconia, Myxomycetes		ND	< 13		100
Total			**13**	**Final MoldSCORE**	**101**

Location: BZ-11 3.04.20

Fungi Identified	Indoor sample spores/m3 <100 1K 10K >100K	Raw count	Spores/ m3	MoldSCORE‡ 100 200 300	Score
Generally able to grow indoors*					
Alternaria		1	13		103
Bipolaris/Drechslera group		ND	< 13		100
Chaetomium		ND	< 13		100
Cladosporium		2	27		101
Curvularia		ND	< 13		100
Nigrospora		ND	< 13		100
Penicillium/Aspergillus types†		4	53		108
Stachybotrys		ND	< 13		100
Torula		ND	< 13		100
Seldom found growing indoors**					
Ascospores		2	27		100
Basidiospores		2	27		100
Rusts		ND	< 13		100
Smuts, Periconia, Myxomycetes		ND	< 13		100
Total			**147**	**Final MoldSCORE**	**108**

Eurofins EPK Built Environment Testing, LLC
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(866) 871-1984 www.eurofinsus.com/Built

Client: STC Environmental Services Inc.
C/O: Michael T. Trevino
Re: 230171; ITC Bldg.

Date of Sampling: 05-22-2023
Date of Receipt: 05-24-2023
Date of Report: 05-25-2023

MoldSCORE™: Spore Trap Report

Location: BZ-12 3.04.10

Fungi Identified	Indoor sample spores/m3 <100 1K 10K >100K	Raw count	Spores/ m3	MoldSCORE‡ 100 200 300	Score
Generally able to grow indoors*					
Alternaria		ND	< 13		100
Bipolaris/Drechslera group		ND	< 13		100
Chaetomium		ND	< 13		100
Cladosporium		14	190		110
Curvularia		1	13		105
Nigrospora		ND	< 13		100
Other brown		3	40		116
Penicillium/Aspergillus types†		ND	< 13		100
Stachybotrys		ND	< 13		100
Torula		ND	< 13		100
Seldom found growing indoors**					
Ascospores		ND	< 13		100
Basidiospores		ND	< 13		100
Rusts		ND	< 13		100
Smuts, Periconia, Myxomycetes		1	13		103
Total			**253**	**Final MoldSCORE**	**122**

Location: BZ-13 3.01.05

Fungi Identified	Indoor sample spores/m3 <100 1K 10K >100K	Raw count	Spores/ m3	MoldSCORE‡ 100 200 300	Score
Generally able to grow indoors*					
Alternaria		ND	< 13		100
Bipolaris/Drechslera group		ND	< 13		100
Chaetomium		ND	< 13		100
Cladosporium		ND	< 13		100
Curvularia		ND	< 13		100
Nigrospora		ND	< 13		100
Penicillium/Aspergillus types†		1	13		102
Stachybotrys		ND	< 13		100
Torula		ND	< 13		100
Seldom found growing indoors**					
Ascospores		ND	< 13		100
Basidiospores		2	27		101
Rusts		ND	< 13		100
Smuts, Periconia, Myxomycetes		ND	< 13		100
Total			**40**	**Final MoldSCORE**	**102**

Eurofins EPK Built Environment Testing, LLC
6301 NW 5th way, Suite#: 1410, Ft. Lauderdale, FL 33309
(866) 871-1984 www.eurofinsus.com/Built

Client: STC Environmental Services Inc.
C/O: Michael T. Trevino
Re: 230171; ITC Bldg.

Date of Sampling: 05-22-2023
Date of Receipt: 05-24-2023
Date of Report: 05-25-2023

MoldSCORE™: Spore Trap Report

Location: BZ-14 2.01 By 2.01.02A

Fungi Identified	Indoor sample spores/m3 <100 1K 10K >100K	Raw count	Spores/ m3	MoldSCORE‡ 100 200 300	Score
Generally able to grow indoors*					
Alternaria		ND	< 13		100
Bipolaris/Drechslera group		ND	< 13		100
Chaetomium		ND	< 13		100
Cladosporium		ND	< 13		100
Curvularia		ND	< 13		100
Nigrospora		ND	< 13		100
Other colorless		1	13		105
Penicillium/Aspergillus types†		ND	< 13		100
Stachybotrys		ND	< 13		100
Torula		ND	< 13		100
Seldom found growing indoors**					
Ascospores		ND	< 13		100
Basidiospores		2	27		101
Rusts		ND	< 13		100
Smuts, Periconia, Myxomycetes		ND	< 13		100
Total			**40**	**Final MoldSCORE**	**105**

Location: BZ-15 2.02

Fungi Identified	Indoor sample spores/m3 <100 1K 10K >100K	Raw count	Spores/ m3	MoldSCORE‡ 100 200 300	Score
Generally able to grow indoors*					
Alternaria		ND	< 13		100
Bipolaris/Drechslera group		ND	< 13		100
Chaetomium		ND	< 13		100
Cladosporium		1	13		101
Curvularia		ND	< 13		100
Nigrospora		ND	< 13		100
Other colorless		1	13		105
Penicillium/Aspergillus types†		ND	< 13		100
Stachybotrys		ND	< 13		100
Torula		ND	< 13		100
Seldom found growing indoors**					
Ascospores		ND	< 13		100
Basidiospores		1	13		100
Rusts		ND	< 13		100
Smuts, Periconia, Myxomycetes		ND	< 13		100
Total			**40**	**Final MoldSCORE**	**105**

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(866) 871-1984 www.eurofinsus.com/Built

Client: STC Environmental Services Inc.
C/O: Michael T. Trevino
Re: 230171; ITC Bldg.

Date of Sampling: 05-22-2023
Date of Receipt: 05-24-2023
Date of Report: 05-25-2023

MoldSCORE™: Spore Trap Report

Location: BZ-16 North By 2.08A

Fungi Identified	Indoor sample spores/m3 <100 1K 10K >100K	Raw count	Spores/ m3	MoldSCORE‡ 100 200 300	Score
Generally able to grow indoors*					
Alternaria		ND	< 13		100
Bipolaris/Drechslera group		ND	< 13		100
Chaetomium		ND	< 13		100
Cladosporium		ND	< 13		100
Curvularia		ND	< 13		100
Nigrospora		ND	< 13		100
Penicillium/Aspergillus types†		2	27		104
Stachybotrys		ND	< 13		100
Torula		ND	< 13		100
Seldom found growing indoors**					
Ascospores		ND	< 13		100
Basidiospores		1	13		100
Rusts		ND	< 13		100
Smuts, Periconia, Myxomycetes		ND	< 13		100
Total			**40**	**Final MoldSCORE**	**104**

Location: BZ-17 2.08-Dome

Fungi Identified	Indoor sample spores/m3 <100 1K 10K >100K	Raw count	Spores/ m3	MoldSCORE‡ 100 200 300	Score
Generally able to grow indoors*					
Alternaria		ND	< 13		100
Bipolaris/Drechslera group		ND	< 13		100
Chaetomium		ND	< 13		100
Cladosporium		ND	< 13		100
Curvularia		ND	< 13		100
Nigrospora		ND	< 13		100
Penicillium/Aspergillus types†		1	13		102
Stachybotrys		ND	< 13		100
Torula		ND	< 13		100
Seldom found growing indoors**					
Ascospores		ND	< 13		100
Basidiospores		1	13		100
Rusts		ND	< 13		100
Smuts, Periconia, Myxomycetes		ND	< 13		100
Total			**27**	**Final MoldSCORE**	**102**

Eurofins EPK Built Environment Testing, LLC
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Client: STC Environmental Services Inc.
C/O: Michael T. Trevino
Re: 230171; ITC Bldg.

Date of Sampling: 05-22-2023
Date of Receipt: 05-24-2023
Date of Report: 05-25-2023

MoldSCORE™: Spore Trap Report

Location: BZ-18 2.04

Fungi Identified	Indoor sample spores/m3 <100 1K 10K >100K	Raw count	Spores/ m3	MoldSCORE‡ 100 200 300	Score
Generally able to grow indoors*					
Alternaria		ND	< 13		100
Bipolaris/Drechslera group		ND	< 13		100
Chaetomium		ND	< 13		100
Cladosporium		ND	< 13		100
Curvularia		ND	< 13		100
Nigrospora		ND	< 13		100
Penicillium/Aspergillus types†		1	13		102
Stachybotrys		ND	< 13		100
Torula		ND	< 13		100
Seldom found growing indoors**					
Ascospores		ND	< 13		100
Basidiospores		1	13		100
Rusts		ND	< 13		100
Smuts, Periconia, Myxomycetes		ND	< 13		100
Total			**27**	**Final MoldSCORE**	**102**

Location: BZ-19 2.05

Fungi Identified	Indoor sample spores/m3 <100 1K 10K >100K	Raw count	Spores/ m3	MoldSCORE‡ 100 200 300	Score
Generally able to grow indoors*					
Alternaria		ND	< 13		100
Bipolaris/Drechslera group		ND	< 13		100
Chaetomium		ND	< 13		100
Cladosporium		ND	< 13		100
Curvularia		ND	< 13		100
Nigrospora		ND	< 13		100
Penicillium/Aspergillus types†		1	13		102
Stachybotrys		ND	< 13		100
Torula		ND	< 13		100
Seldom found growing indoors**					
Ascospores		ND	< 13		100
Basidiospores		1	13		100
Rusts		ND	< 13		100
Smuts, Periconia, Myxomycetes		ND	< 13		100
Total			**27**	**Final MoldSCORE**	**102**

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Client: STC Environmental Services Inc.
C/O: Michael T. Trevino
Re: 230171; ITC Bldg.

Date of Sampling: 05-22-2023
Date of Receipt: 05-24-2023
Date of Report: 05-25-2023

MoldSCORE™: Spore Trap Report

* The spores in this category are generally capable of growing on wet building materials in addition to growing outdoors. Building related growth is dependent upon the fungal type, moisture level, type of material, and other factors. *Cladosporium* is one of the predominant spore types worldwide and is frequently present in high numbers. *Penicillium/Aspergillus* species colonize both outdoor and indoor wet surfaces rapidly and are very easily dispersed. Other genera are usually present in lesser numbers.

** These fungi are generally not found growing on wet building materials. For example, the rusts and smuts are obligate plant pathogens. However, in each group there are notable exceptions. For example, agents of wood decay are members of the basidiomycetes and high counts of a single morphological type of basidiospore on an inside sample should be considered significant.

†The spores of Aspergillus and Penicillium (and others such as Acremonium, Paecilomyces) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods.

‡Rated on a scale from 100 to 300. A rating less than 150 is low and indicates a low probability of spores originating inside. A rating greater than 250 is high and indicates a high probability that the spores originated from inside, presumably from indoor mold growth. A rating between 150 and 250 indicates a moderate likelihood of indoor fungal growth. MoldSCORE is NOT intended for wall cavity samples. It is intended for ambient air samples in residences. Using the analysis on other samples (like wall cavity samples) will lead to misleading results.

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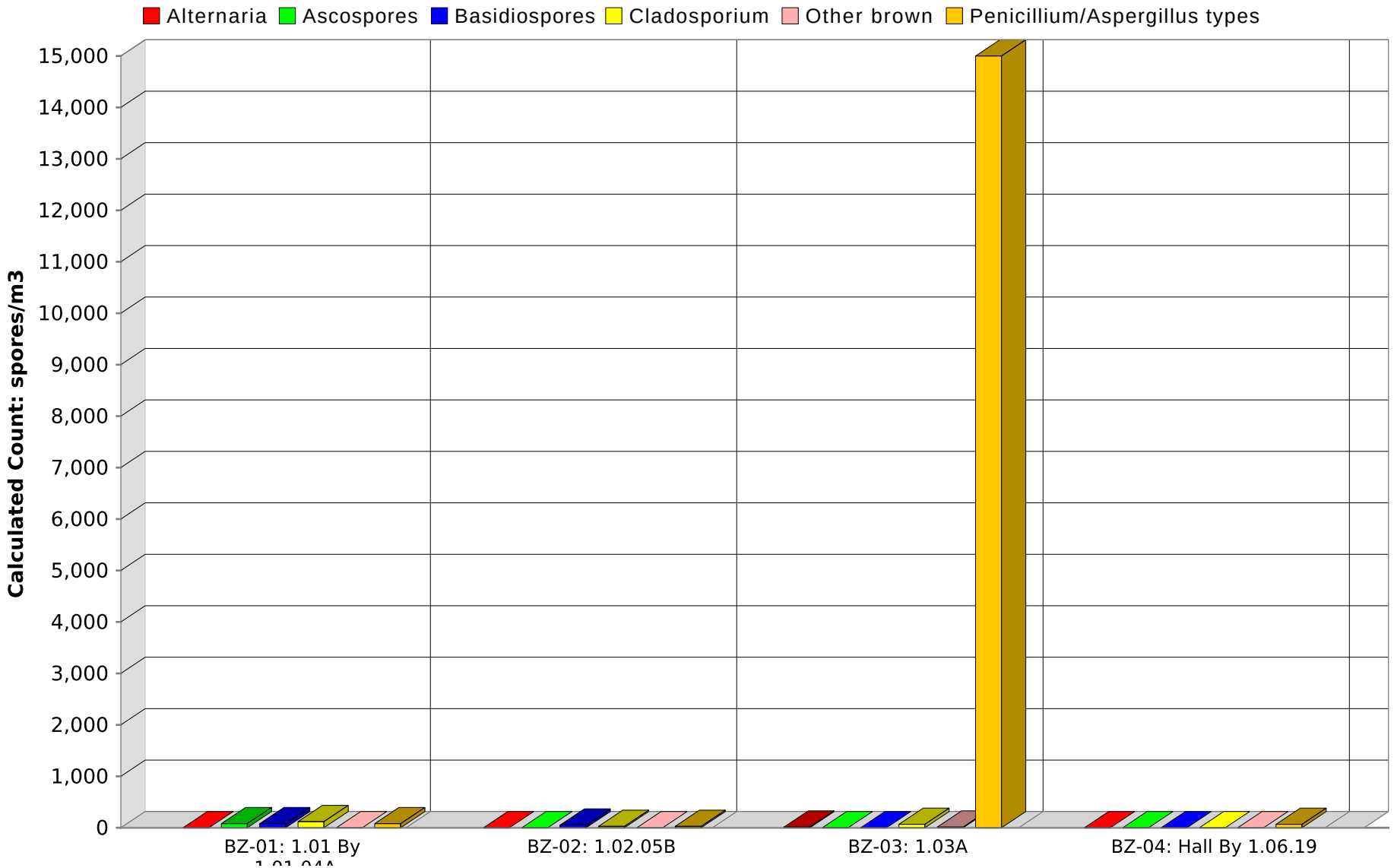
SPORE TRAP REPORT: NON-VIABLE METHODOLOGY



Comments:

Note: Graphical output may understate the importance of certain "marker" genera.

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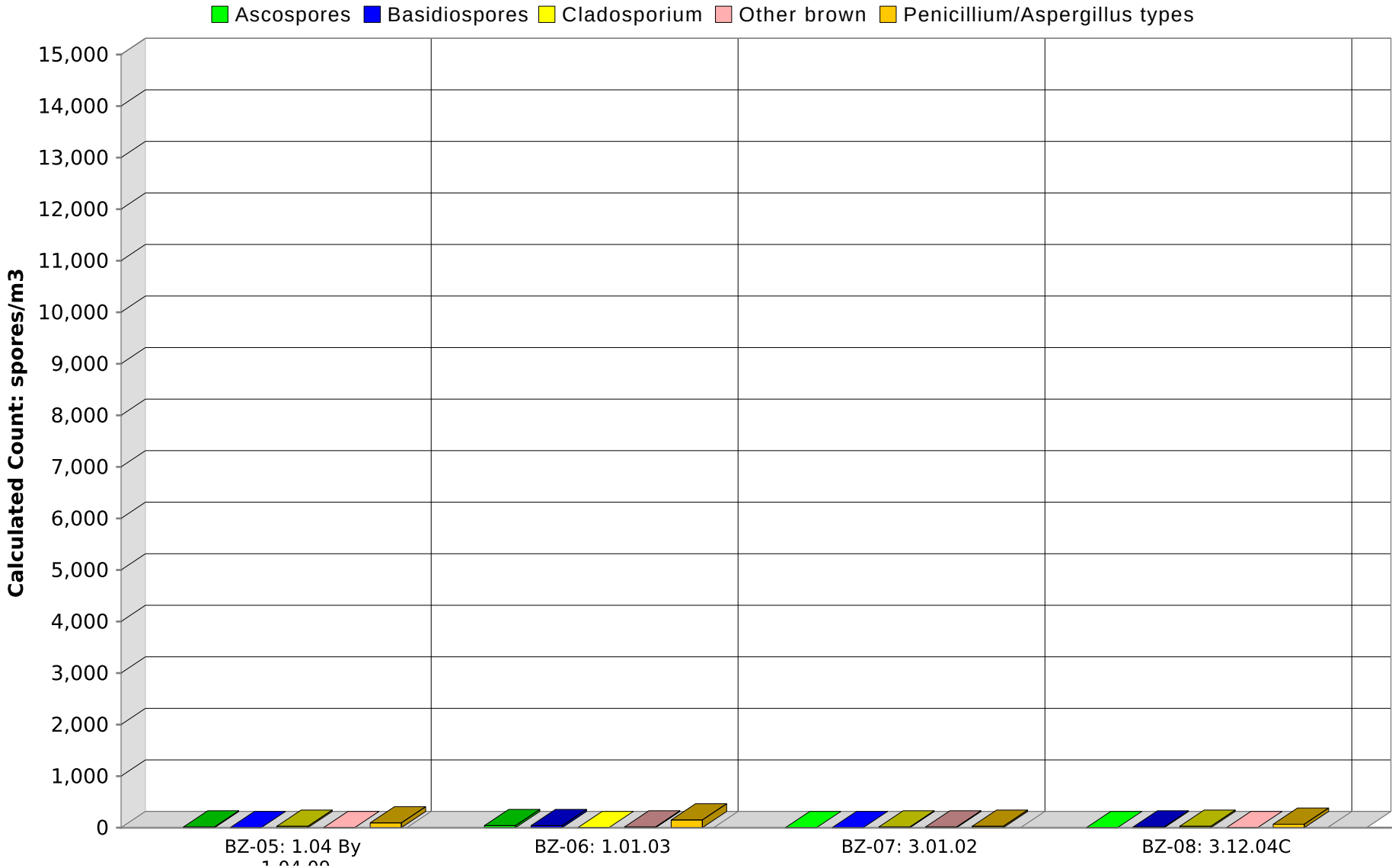
SPORE TRAP REPORT: NON-VIABLE METHODOLOGY



Comments:

Note: Graphical output may understate the importance of certain "marker" genera.

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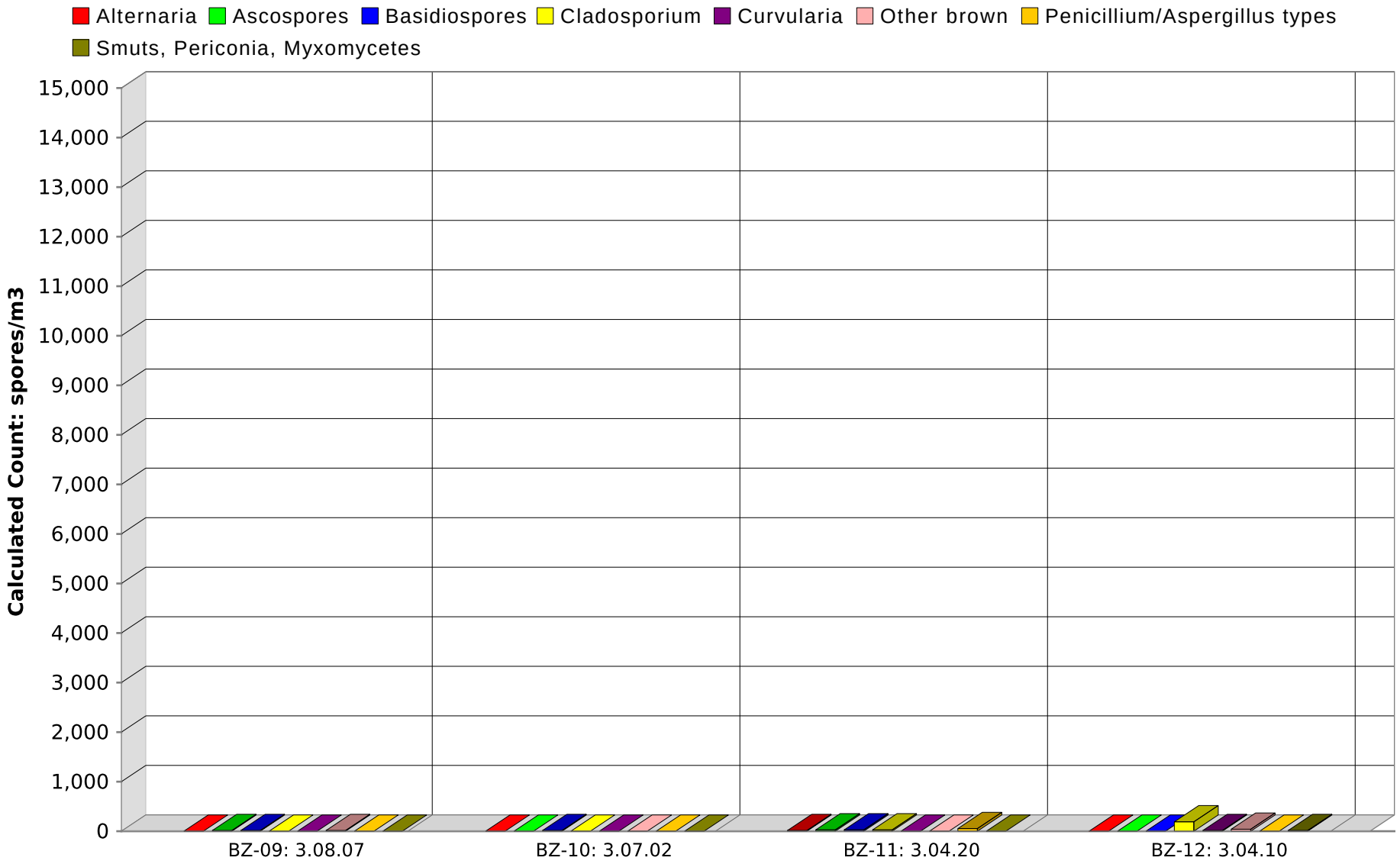
SPORE TRAP REPORT: NON-VIABLE METHODOLOGY



Comments:

Note: Graphical output may understate the importance of certain "marker" genera.

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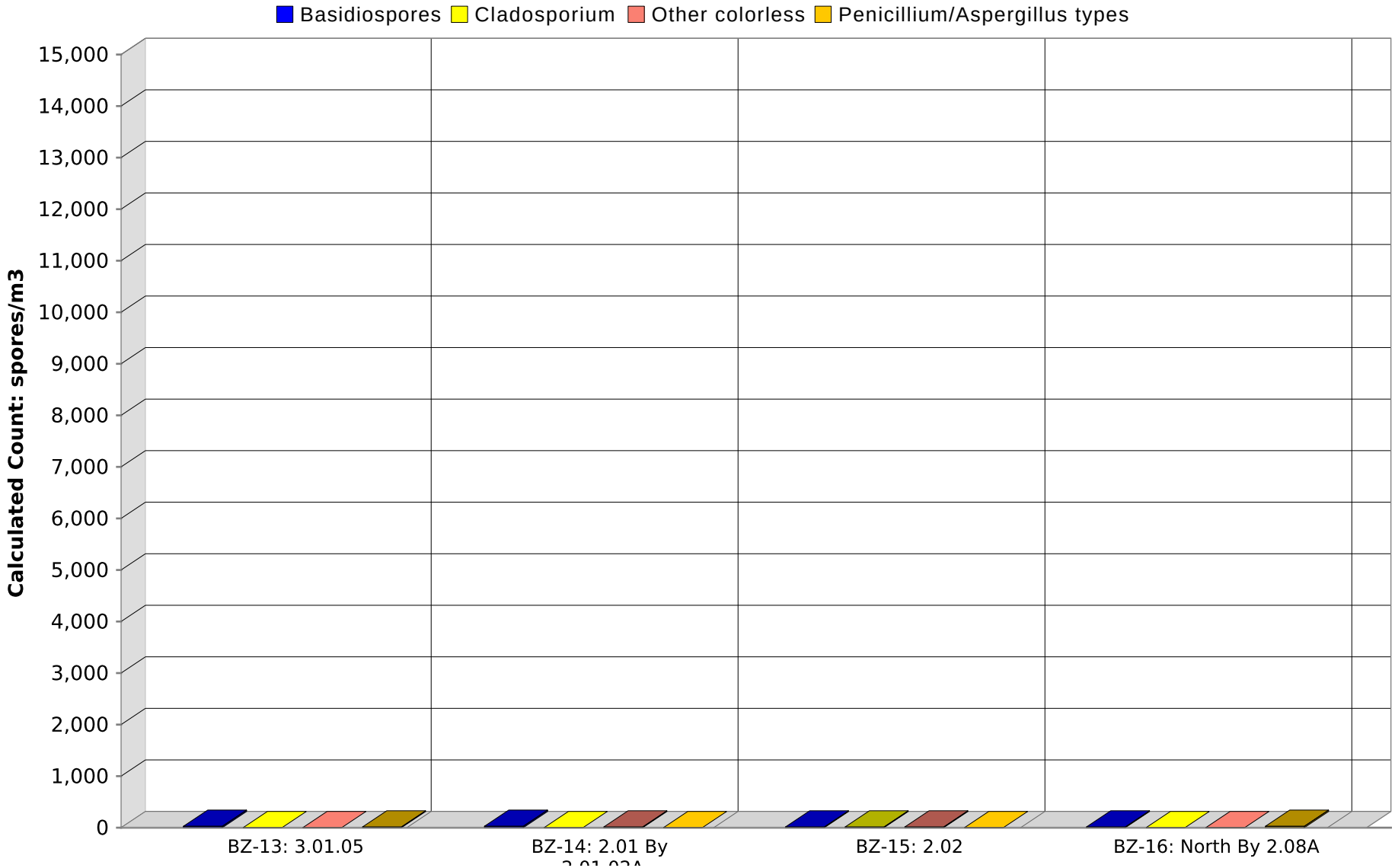
SPORE TRAP REPORT: NON-VIABLE METHODOLOGY



Comments:

Note: Graphical output may understate the importance of certain "marker" genera.

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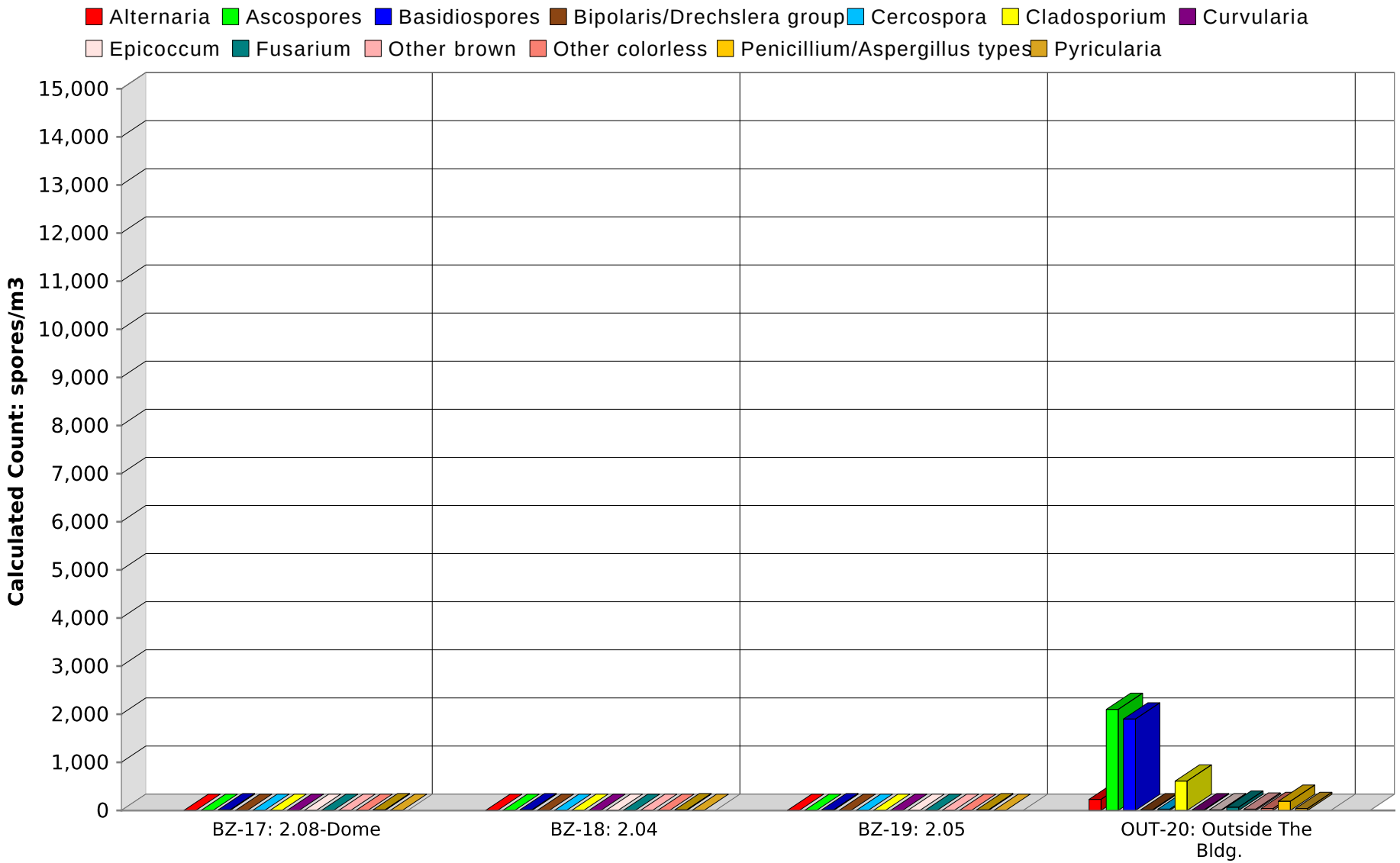
(866) 871-1984 www.eurofinsus.com/Built

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY



Comments:

Note: Graphical output may understate the importance of certain "marker" genera.

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SSF, CA: 6000 Shoreline Court, Suite 205, South San Francisco, CA 94080 * (866) 888-6653

Page 1 of 2

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CONTACT INFORMATION

Company:	STC Environmental Services, Inc.	Address:	4754 Research Drive., San Antonio, Texas 78240
Contact:	Michael Trevino	Special Instructions:	
Phone:	210-696-6288		Email report to: mtrevino@stces.com

PROJECT INFORMATION

Project ID:	230171		
Project Description:	ITC Bldg.		
Project Zip Code:	78205	Sampling Date & Time:	5/22/23
PO Number:	0167-23	Sampled By:	MTT

TURN AROUND TIME CODES (TAT)

- (STD) – Standard (DEFAULT) [circled]
- ND – Next Business Day
- SD – Same Business Day Rush*
- *Please call Client Services for locations with Rush services

Rushes received after 2pm or on weekends, will be considered received the next business day. Please alert us in advance of weekend analysis needs.

Sample ID	Description	Sample Type (Below)	TAT (Above)	Total Volume (Air Samples only)	Notes
BZ-01	1.01 5z 1.01.04A	O	STD	75L	4917960
BZ-02	1.02.05B	↓	↓	↓	4917970
BZ-03	1.03A	↓	↓	↓	4917928
BZ-04	Hall 5z 1.06.19	↓	↓	↓	4917691
BZ-05	1.04 5z 1.04.09	↓	↓	↓	MTT ~~4917601~~ 4917708
BZ-06	1.01.03	↓	↓	↓	4917601
BZ-07	3.01.02	↓	↓	↓	4917962
BZ-08	3.12.04C	↓	↓	↓	4917939
BZ-09	3.08.07	↓	↓	↓	4917560
BZ-10	3.07.02	↓	↓	↓	4917640
BZ-11	3.04.20	↓	↓	↓	4917600

REQUESTED SERVICES (Check boxes below)

PCM Air: Fiber Count (NIOSH 7400)	PCM Air: OSHA with TWA	PLM Bulk: Asbestos Bulk PLM	PLM Bulk: EPA Point Count (200 Point Count)	PLM Bulk: EPA Point Count (400 Point Count)	PLM Bulk: EPA Point Count (1000 Point Count)	PLM Bulk: Gravimetric Point Count (400 Pt Count)	PLM Bulk: Gravimetric Point Count (1000 Pt Count)	Rock & Soil: CARB 435 Method (400 Point Count)	Rock & Soil: CARB 435 Method (1000 Point Count)	Other Requests: Lead Analysis – Flame AA	Other Requests: Fungi Spore Trap	Other Requests
☐	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐	☒	☐
☐	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐	↓	☐
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☐	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐	↓	☐

SAMPLE TYPE CODES

A – Air	W – Wipe
B – Bulk	T – Tape
D – Dust	R – Rock
SO – Soil	O – Other:

RELINQUISHED BY	DATE & TIME	RECEIVED BY	DATE & TIME
[signature]	5/22/23	GA	5/23/24 10 am

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CHAIN OF CUSTODY

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003269888

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CONTACT INFORMATION

Company:	STC Environmental Services, Inc.	Address:	4754 Research Drive., San Antonio, Texas 78240
Contact:	Michael Trevino	Special Instructions:	
Phone:	210-696-6288		Email report to: mtrevino@stces.com

PROJECT INFORMATION

Project ID:	See Page 1		
Project Description:			
Project Zip Code:		Sampling Date & Time:	
PO Number:		Sampled By:	

TURN AROUND TIME CODES (TAT)

(STD) – Standard (DEFAULT)

ND – Next Business Day

SD – Same Business Day Rush*

*Please call Client Services for locations with Rush services

Rushes received after 2pm or on weekends, will be considered received the next business day. Please alert us in advance of weekend analysis needs.

REQUESTED SERVICES (Check boxes below)

Sample ID	Description	Sample Type (Below)	TAT (Above)	Total Volume (Air Samples only)	Notes	PCM Air: Fiber Count (NIOSH 7400)	PCM Air: OSHA with TWA	PLM Bulk: Asbestos Bulk PLM	PLM Bulk: EPA Point Count (200 Point Count)	PLM Bulk: EPA Point Count (400 Point Count)	PLM Bulk: EPA Point Count (1000 Point Count)	PLM Bulk: Gravimetric Point Count (400 Pt Count)	PLM Bulk: Gravimetric Point Count (1000 Pt Count)	PLM Rock & Soil: CARB 435 Method (400 Point Count)	PLM Rock & Soil: CARB 435 Method (1000 Point Count)	Other Requests: Lead Analysis – Flame AA	Other Requests: Fungal Spore Trap	Other Requests:
BZ-12	3.04.10	O	STD	75L	4917637	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐	☒	☐
BZ13	3.01.05				4917952	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐
BZ-14	2.01 by 2.01.02A				4917577	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐
BZ-15	2.02				4917930	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐
BZ-16	North of 2.08A				4917938	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐
BZ-17	2.08-Done				4917576	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐
BZ-18	2.04				4917597	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐
BZ-19	2.05				4917655	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐
BZT-20	Outside the Bldg.				4917951	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐
						☐	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐
						☐	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐	☐

SAMPLE TYPE CODES	
A – Air	W – Wipe
B – Bulk	T – Tape
D – Dust	R – Rock
SO – Soil	(O) – Other:

RELINQUISHED BY	DATE & TIME	RECEIVED BY	DATE & TIME
	5/22/23	GT	5/24/23
			10 AM